# Reviews of Physiology, Biochemistry and Pharmacology

Editors

R. H. Adrian, Cambridge · E. Helmreich, Würzburg H. Holzer, Freiburg · R. Jung, Freiburg O. Krayer, Boston · R. J. Linden, Leeds F. Lynen †, München · P. A. Miescher, Genève J. Piiper, Göttingen · H. Rasmussen, New Haven A. E. Renold, Genève · U. Trendelenburg, Würzburg K. Ullrich, Frankfurt/M. · W. Vogt, Göttingen A. Weber, Philadelphia

With 29 Figures

Springer-Verlag Berlin Heidelberg New York 1981

### **Reviews of Physiology, Biochemistry and Pharmacology**

formerly Ergebnisse der Physiologie, biologischen Chemie und experimentellen Pharmakologie

### ISBN 3-540-10408-9 Springer-Verlag Berlin Heidelberg New York ISBN 0-387-10408-9 Springer-Verlag New York Heidelberg Berlin

Library of Congress-Catalog-Card Number 74-3674

This work is subject to copyright. All rights are reserved, whether the whole or part of the material is concerned, specifically those of translation, reprinting, re-use of illustrations, broadcasting, reproduction by photocopying machine or similar means, and storage in data banks. Under § 54 of the German Copyright Law where copies are made for other than private use, a fee is payable to 'Verwertungsgesellschaft Wort', Munich.

© by Springer-Verlag Berlin Heidelberg 1981 Printed in Germany.

The use of registered names, trademarks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

Offsetprinting and Binding: Konrad Triltsch, Würzburg 2121/3130-543210

# Contents

Walter R. Hess (1881 – 1973). Von R. Jung, Freiburg/Bundesrepublik Deutschland. Mit 3 Abbildungen	1
Neural Organisation and Control of the Baroreceptor Reflex. By K. M. Spyer, London/Great Britain. With 13 Figures	23
Event-Related Slow (DC) Potentials in the Human Brain. By M. Haider, E. Groll-Knapp, and J. A. Ganglberger, Vienna/Austria. With 12 Figures	125
<ul> <li>α-Adrenoceptor Subclassification.</li> <li>By K. Starke, Freiburg/Federal Republic</li> <li>of Germany. With 1 Figure</li> </ul>	199
Author Index	237
Subject Index	259
Indexed in Current Contents	

# Walter R. Hess (1881-1973)

RICHARD JUNG \*

Am 12. August 1973 starb Walter Rudolf *Hess* im 93. Lebensjahr in Ascona nach einer über sechs Jahrzehnte währenden Tätigkeit als Physiologe. Mit dem Zürcher Physiologischen Institut war *Hess* 1912–1917 als Assistent und Privatdozent, 1917–1951 als Professor und Direktor und 1951–1960 als Emeritus eng verbunden. Dort hat er seine berühmten Arbeiten über das vegetative System, den Schlaf und die Motorik durchgeführt und war Lehrer vieler Generationen von Physiologen und Ärzten. Sein Lebensweg ist kurz zu schildern, seine Forschungsleistungen bedürfen einer ausführlicheren Darstellung.

### **Biographie und Persönlichkeit**

Der Lebensweg. Hess wurde am 17. März 1881 in Frauenfeld (Kanton Thurgau) als Sohn eines Gymnasialprofessors der Physik geboren, dem er die frühesten Anregungen zur naturwissenschaftlichen Forschung verdankt [40]. Er studierte 1899–1905 in Lausanne, Berlin, Kiel und Zürich Medizin. Nach ophthalmologischer Ausbildung und früher Heirat mit Luise Sandmeyer 1909 wurde er 1912 nach dreijähriger augenärztlicher Tätigkeit wieder Physiologe. An der Zürcher Universität, die ihm über 60 Jahre geistige Heimat blieb, wurde Hess 1913 Privatdozent und 1917 ordentlicher Professor der Physiologie und Direktor des Physiologischen Instituts. Von 1912–1951 hat er das Zürcher Institut nur kurz verlassen: für wenige Vortragsreisen, für die Ferien in Ascona, wo er seine Bücher schrieb, und nur einmal für ein Jahr Forschungsarbeit in Bonn als Gastassitent bei Verworn 1915/16. Für seine Entdeckung der Hirnstammreizeffekte auf das Verhalten der wachen Katze, die er 1927–1947 in systematischen Experimenten ausbaute, erhielt Hess 1949 den Nobelpreis für Physiologie und Medizin. Außer zahlreichen Ehrenmitgliedschaften Wissenschaftlicher Gesellschaften und Akademien erhielt er die Carl-Ludwig-Medaille und die Johannes-Müller-Medaille. 1951 wurde er emeritiert, arbeitete aber an der Auswertung seiner Experimente weiter, bis er sich im 85. Lebensjahr ganz in sein Ferienhaus in Ascona zurückzog.

<sup>\*</sup> Abteilung für Neurophysiologie der Universität Freiburg, Hansastr. 9a, D-7800 Freiburg

Der Physiologe. Sein langes Forscherleben in der Physiologie führte *Hess* vom Kreislauf [4, 5, 12] zu den vegetativen Hirnfunktionen [7, 14, 20], zur Atmung [13, 17, 18, 21], zum Schlaf [9–11, 16, 28], im dritten und sechsten Lebensjahrzehnt zur Motorik [3, 22–27] und schließlich zur Psychophysiologie [39].

Hess begann seine wissenschaftliche Arbeit 1903 als 22jähriger Student mit der eigenwilligen Beobachtung und Erklärung einer Gefäßanomalie, von der er mechanische Gesetzmäßigkeiten des Blutgefäßverlaufs mit ihrer Bedeutung für den Kreislauf ableitete [1]. Er erregte damit die Aufmerksamkeit W. Rouxs, des Begründers der Entwicklungsmechanik. Seitdem beschäftigten ihn die Kreislaufphysiologie und Neurophysiologie als Forschungsgebiet über fünf Jahrzehnte. Dennoch unterbrach er die geplante experimentelle Arbeit für fast sieben Jahre: 1905 begann er unter Einfluß des Ophthalmologen Haab eine klinisch-augenärztliche Fachausbildung an der Zürcher Augenklinik und arbeitete dort über Augenmuskellähmungen und ihre Doppelbildmessung [3]. Nach seiner Heirat eröffnete er eine Praxis als Augenarzt in Rapperswil am Zürcher See. Er wollte zunächst durch ärztliche Tätigkeit ein sicheres Familieneinkommen haben. Doch sein wissenschaftlicher Impetus und seine Liebe zur Physiologie setzten sich bald wieder durch: 1912 ging er an die Universität Zürich zurück, wo er sich im Physiologischen Institut bei Gaule schon im nächsten Jahre mit einer Arbeit über die Hämodynamik habilitierte [47]. 1915/16, während des ersten Weltkriegs, arbeitete er ein Jahr in Bonn bei Max Verworn, dem damals führenden deutschen Neurophysiologen. Verworns allgemeine Physiologie und sein weiter biologischer Aspekt, der von den Einzellern bis zum menschlichen Gehirn reichte, waren für Hess prägende Eindrücke. Doch die Erkrankung Verworns, der in Hess' Gegenwart seinen ersten cerebralen Schlaganfall erlitt, und die Kriegsverhältnisse beeinträchtigten die gemeinsame Arbeit.

Die Rückkehr nach Zürich brachte durch die Erkrankung Gaules für Hess eine schwere Zeit, die er mit Energie und Zähigkeit meisterte. Er mußte sich von 1916 bis 1919 fast ganz dem Unterricht und der Institutsorganisation widmen, mehrfach unterbrochen durch kurze Einberufungen zum Militärdienst. Hess konnte damals nur wenige Experimente über die Organdurchblutung fortführen, und die Forschung blieb bis zum Kriegsende liegen. Sein meist 15stündiger Arbeitstag reichte von morgens 7 bis abends 22 Uhr mit Vorlesungen, Kursen, Institutsordnung und Anleitung seiner Assistenten. Zwei tüchtige Mitarbeiter, die sich 1920/21 bei ihm habilitierten, der vorwiegend biophysikalisch ausgerichtete Alfred Fleisch und der mehr biochemisch-pharmakologisch interessierte Ernst Rothlin, halfen ihm, das Zürcher physiologische Institut trotz der ungünstigen Zeitverhältnisse zu neuer Aktivität zu führen. Nach Gaules vorzeitiger Emeritierung wurde der 36jährige Hess gegen manche Widerstände zum Professor



Abb. 1a, b. W.R. Hess als Privatdozent 1914 und als Professor 1936. a Der 33jährige Assistent von Gaule war damals seit 1912 am Zürcher Physiologischen Institut und seit 1913 habilititert, bevor er 1915–1916 neurophysiologisch bei Verworn in Bonn arbeitete. b Der 54jährige war seit 1917 Direktor des Physiologischen Instituts der Universität Zürich. 1936, als ich in sein Institut eintrat, bearbeitete Hess die Effekte der Zwischenhirnreizung auf Atmung und Kreislauf [20], nachdem er 1926–28 die intrakranielle Reiztechnik an der freien Katze entwickelt hatte [8, 9, 14]

der Physiologie und Institutsdirektor berufen. Die Chance, als junger Physiologe ein eigenes Institut aufzubauen und die Forschung zu fördern, hat *Hess* in 44 Jahren unermüdlicher Tätigkeit erfolgreich ausgenutzt. 1921 konzentrierte sich *Hess* nach einer orientierenden Reise durch die britischen Institute und den Besuch des Internationalen Physiologen-Kongresses in Edinburgh zunächst auf den Abschluß seiner Arbeiten zur Regulierung von Blutkreislauf und Atmung, die zu zwei Monographien [12, 13] erweitert wurden. 1924 schrieb er eine programmatische Arbeit über die Beziehung von psychischen und vegetativen Funktionen [7], die zur gedanklichen Grundlage seiner über zweieinhalb Jahrzehnte dauernden Experimente am zentral-vegetativen System des Zwischenhirns führte (vgl. S. 9).

1929 beim internationalen Physiologen-Kongreß in Boston lernte Hess die amerikanischen Physiologen kennen. Seine Hoffnung, W. Cannon für seine Konzeption der vegetativen Funktionen zu gewinnen, wurde enttäuscht. Eine gemeinsame Studienreise mit dem deutschen Physiologen Wachholder durch die nordamerikanischen Forschungsinstitute brachte bessere Kontakte, aber *Hess* blieb gegenüber den angloamerikanischen Physiologen reserviert und hielt die stereotaktischen Reizexperimente am narkotisierten Tier mit faradischer Reizung von *Ransom* und seiner Schule für unphysiologisch. Die deutschsprachige Physiologie war seine geistige Heimat, und er fühlte sich in der von ihm so genannten deutschen "Physiologenfamilie" zu Hause, in guter Freundschaft mit Gleichgesinnten und in kritischem Abstand zu anderen. Auch nach Gründung einer eigenen Schweizer Physiologischen Gesellschaft erschien ihm die Schweiz für wissenschaftliche Diskussion und produktiven Ideenaustausch zu klein. Die *Tagungen der Deutschen Physiologischen Gesellschaft* hat *Hess* über fünf Jahrzehnte regelmäßig besucht. Dort trug er seine experimentellen Ergebnisse vor und demonstrierte zuerst seine neue Methode der Zwischenhirn-



Abb. 2a-c. *Hess* auf Kongressen, im Institut und zu Hause 1948–1953. a Diskussion mit Fachkollegen auf der deutschen Physiologentagung in Frankfurt 1948. b Demonstration eines Experimentes vor Studenten 1949. c Im häuslichen Arbeitszimmer mit dem Papagei auf der Schulter 1953

reizung an der freilaufenden Katze auf der Deutschen Physiologentagung in Bonn 1927 [8]. Er blieb bis in sein hohes Alter ein anregender und kritischer Diskutant auf diesen Tagungen (Abb. 2a). Von 1928–1954 kam er oft nach Baden-Baden zu den Südwestdeutschen Neurologen und Psychiatern, in deren Versammlungen er 1928 und 1929 seine ersten Experimente über Schlafauslösung durch Zwischenhirnreiz bei der Katze im Film demonstrierte [9, 11]. Diese drei Kurzmitteilungen bei Neurologen- und Physiologentagungen [8, 9, 11] waren außer seiner Schlaftheorie [10, 16] über zehn Jahre seine einzigen Publikationen über den Schlaf durch Thalamusreiz. 1932 beschrieb er monographisch seine Methodik der Reizung und Ausschaltung lokalisierter Hirnstrukturen [14] und 1938 die Zwischenhirnbeeinflussung von Atmung und Kreislauf [20].

Hess organisierte Institut und Forschung nach funktionellen Gesichtspunkten. Auch bei Experimenten verwendete er die gleiche Regel, die er als Funktionsprinzip des lebenden Organismus ansah: die Kraft soll dort einsetzen, wo sie ihre größte Wirkung entfaltet. Bei der bis 1946 sehr geringen Dauerpersonalbesetzung des Instituts (2 Assistenten, 1 Mechaniker, 1 Sekretärin) hat er Hilfskräfte angelernt und aus eigenen oder aus Forschungsmitteln bezahlt. Eine Kinderschwester, die er im Haus nicht mehr brauchte und deren Interesse für systematische Ordnung er kannte, hat über 20 Jahre die statistische Dokumentation seiner Hirnreizexperimente durchgeführt. Wenn Hess beim Experiment einen Reizerfolg sah, fragte er sie, wann und mit welcher Lokalisation der Effekt schon früher vorkam. Wenn Frl. Jaussi (Abb. 3b, links) es nicht auswendig wußte, blätterte sie in der Kartei, und in wenigen Minuten kam die Antwort. So konnte Hess Reizpunktkarten der verschiedenen vegetativen und motorischen Symptome aufstellen und dann mit anatomischen Strukturen in Verbindung bringen [31, 36].

Charakter und Arbeitsweise. Hess verstand es, systematische Theorie und methodisches Experimentieren in seiner Forschungsarbeit zu vereinen. Weiter Interessenkreis, gedankliche Klarheit und hartnäckiger Fleiß, geführt durch gezielte Theorienbildung und gezügelt durch methodische Beschränkung, das waren die Charakteristika seiner Arbeitsweise und die Grundlagen seiner Erfolge. Daraus entwickelte Hess seine Synthese der Physiologie, die mit seinen Forschungsleistungen dargestellt werden muß.

Hess war eine glückliche Mischung eidgenössischer Zähigkeit und Strenge (die er von seinem Schweizer Vater herleitete) mit lebhaftem Forschungsgeist und wachem Interesse für alles Neue (das er als Erbteil seiner sächsischen Mutter ansah [40]). Diese beiden Seiten seines Charakters bestimmten Hess' Forschungsimpetus: Die Aufgeschlossenheit für neue Probleme förderte die Vielseitigkeit seiner Physiologie. Zähigkeit und Exaktheit führten ihn zur systematischen Planung und Ausarbeitung der Ergebnisse über viele Jahre.

Hess war auch ein Meister experimenteller Methodik und ihrer kritischen Begrenzung. Obwohl er sich in Gespräch und Diskussion für neue Ideen und Ergebnisse begeistern konnte, begrenzte er seine praktische Forschung auf erreichbare Ziele, die er mit systematischem Denken und kritisch-methodischer Strenge verfolgte. Als ich ihm 1937 in jugendlichem Optimismus vorschlug, seine cerebralen Reiz- und Ausschaltungsexperimente durch die Ableitung von Hirnpotentialen zu erweitern, winkte Hess ab. Obwohl er mit O. Wyss einen sehr guten Elektrophysiologen als Oberassistenten hatte, meinte er, hirnelektrische Untersuchungen sollten andere Institute machen: er hätte mit der Auswertung der Reizversuche und ihrer ihm immer etwas fremden hirnanatomischen Korrelation schon zu viele ungelöste Aufgaben für das nächste Jahrzehnt.

Hess beschränkte die Hirnstammexperimente auf das Versuchstier Katze und begründete dies mit Verhaltensbeobachtungen, der geringen Variation von Hirn- und Körpergröße und der guten Sehorientierung von Katzen im Vergleich zum Hund. Experimente an Affen hielt er bei Hirnstammfunktionen für unnötig kostspielig. Für seine Schlafuntersuchungen war die Katze das ideale Versuchstier, da sie gern und leicht am Tage schläft. Dies erleichterte seine frühen Experimente der Schlafauslösung durch Zwischenhirnreizung [8-11, 28].

Hess erfüllte die doppelte Aufgabe als Lehrer und Forscher, indem er den Unterricht zeitlich genau einteilte und sich für die Experimente bestimmte Stunden und Tage frei hielt. Seine Forschung plante Hess kontinuierlich und systematisch. Kein Tag blieb ohne Arbeit. Im Hause am Zürichberg saß er in der Veranda und schrieb, oft mit seinem Papagei auf der Schulter (Abb. 2c). Wenn er aus dem Institut nach Hause kam, und in den Ferien, wenn er seinen Garten in Ascona bestellt hatte, entwarf er seine vielfach umkorrigierten Arbeiten. Im Institut halfen ihm treu ergebene Mitarbeiter, in der Wohnung auf dem Zürichberg und im Sommerhaus in Ascona seine Frau bei der Vollendung seiner wissenschaftlichen Schriften, die alle seine persönliche Prägung trugen. Obwohl ihm, wie er oft sagte, das Schreiben schwer fiel, entstand zwischen 1903 und 1970 in kontinuierlichem Fluß seine erstaunliche Publikationsreihe von 294 Einzelarbeiten und 10 Monographien.

### Hess' Konzeptionen und Forschungsleistungen

Forschungsergebnisse. Die wichtigsten physiologischen Leistungen von *Hess* seien im Folgenden mit ihrem Zeitgang kurz charakterisiert:

- 1) Systemkonzeptionen des Blutkreislaufs und der Atmung als Kombination hämodynamischer Faktoren mit peripheren und zentralen Regulationen (1913–1931 [4, 5, 12]).
- 2) Funktionsordnung des vegetativen Systems mit zentraler Organisation von zwei reziprok koordinierten Untersystemen, die er ergotrop und trophotrop nannte (1924–1938 [7, 16, 30]).
- 3) Methodische Entwicklung der subkortikalen Hirnreizung beim freibeweglichen Tier (1927–1931 [8, 14]).
- 4) Erforschung der Zwischenhirnfunktionen für vegetative und Verhaltensregulationen (1929–1949 [20, 22, 31, 37]).
- 5) Begründung der experimentellen *Schlafforschung* mit Entdeckung der Schlafauslösung durch elektrische Thalamusreizung (1929–1944 [9, 16, 28]).
- 6) Entdeckung richtungsbestimmter motorischer Regulationszentren im Zwischenhirn und Mittelhirn (1941–1943 [22–24]).
- 7) Koordination der Zielbewegung und Stützmotorik beim Menschen (1941–1965 [24, 27, 42]).
- 8) Synthese physiologischer Verhaltenskoordinationen mit psychischen und vegetativen Funktionen (1943–1962 [26, 39]).

Integrative Physiologie. Hess interessierte sich immer für die ganze Physiologie, und seine Arbeiten reichten in einem breiten Spektrum von den Kreislauf- und Atmungsfunktionen bis zur Hirnphysiologie. Reines Spezialistentum war ihm fremd. Er war, wie Verworn, ein synthetischer Denker. Physiologische Mechanismen sah er immer im Zusammenhang mit ihrem biologischen Zweck innerhalb des Organismus und mit dem Ziel des Tieres im triebgesteuerten Verhalten. Kreislauf, Atmung, vegetative Funktionen und Hirntätigkeit studierte er experimentell als Teilfunktionen des ganzen Organismus. Hess wurde nicht müde, seine Schüler mit vielfach variierten Beispielen dynamisch-integrierter Körperregulationen zum funktionellphysiologischen Denken zu erziehen, das er dem statischen "anatomischen" Denken entgegensetzte. Seine Physiologie war eine Synthese von Theorie und Experiment zur Erforschung integrativer zielgesteuerter Leistungen. die eine lebendige Ordnung im Organismus erkennen läßt. Einen ähnlichen Gegensatz sah er zwischen seiner integrativen System-Physiologie und der mehr auf "facts" und Einzelanalyse eingestellten Haltung angelsächsicher Physiologen, denen oft die Tatsache alles und die Theorie nichts galt. Für Hess dagegen brachten Einzeltatsachen nur Hinweise für Funktionsgesetze. Nur ihre gesetzmäßige Wiederholung oder Zusammenhänge mit anderen Tatsachen waren als physiologische Funktionen zu werten, in die er auch psychische Leistungen einschloß [7]. Seine Betonung von Ziel und Funktionsordnung ist oft als "teleologisch" mißverstanden worden, war aber

immer *biologisch* gemeint. Daher suchte er auch Beziehungen zur Verhaltensforschung von *Lorenz* und seiner Schule und zu v. *Holsts* Regelungsprinzipien. Für *Hess* war das Gehirn eine Struktur- und Funktionsordnung, die als "dynamisches Kräftespiel" nie zur Ruhe kommt, nur bei relativem Gleichgewicht Ruhe vortäuscht und in einem dauernden Wechsel von Erregung und Hemmung verschiedener Systeme biologisch-zielstrebig arbeitet [39].

Kreislaufphysiologie. Die Funktionen des Blutkreislaufes haben Hess seit seiner Gefäßarbeit [1] als Student besonders interessiert. Erste selbständige Leistungen waren ein Apparat zur Messung der Blutviskosität 1907 [2] und die Habilitationsschrift über das Ökonomieprinzip der Hämodynamik 1914 [4]. Daraus entwickelte sich im Laufe von zwei Jahrzehnten eine systematische Untersuchung der Kreislaufregelung, die 1930 monographisch dargestellt wurde [12]. 20 Jahre bevor die Regelprinzipien mit dem Stichwort "Kybernetik" in der Neurophysiologie aktuell wurden, hat Hess alle Regulierungen mit dem später so benannten negativen und positiven "Feedback" an den Leistungen des Blutkreislaufs klar beschrieben. Als einfachste Regelung der Gewebsdurchblutung betrachtete er die Nutritionsreflexe, die er auch "Eigenreflexe des Kreislaufs" nannte und mit der Propriozeptivität der Muskulatur in Parallele setzte: Die Gefäßerweiterung durch CO<sub>2</sub> und lokale Stoffwechselprodukte einschließlich der Axonreflexe sorgt für adäquate Blutversorgung entsprechend der Tätigkeit der Organe. Antagonistisch zum Nutritionsreflex wirkt der Entlastungsreflex, der in Parallele zur Vaguswirkung am Herzen, den Depressor- und Carotis-Sinus-Reflexen, eine Schutzfunktion für das Gewebe hat. Beide reziprok arbeitenden Reflexe zügeln auch mit gleichzeitiger Aktivierung die Durchblutung und den Gewebsstoffwechsel. Auch der meist wenig verstandenen Gefäßerweiterung durch antidrome sensible Impulse gab Hess einen funktionellen Sinn zur Regelung der Gewebefunktion: Die Schmerzfasern unterstützen eine schmerzunterschwellige Gewebstrophik. Nur bei stärkeren Reizen entstehen Schmerz oder Entzündung, bei schwächeren nur physiologische Durchblutungssteigerungen. Man erkennt in den antagonistischen Konzeptionen der Durchblutungsregelung unschwer den Ursprung von Hess' Schema des trophotropen und ergotropen vegetativen Systems [16, 30].

Wie der Untertitel seiner Kreislauf-Monographie [12] zeigt und Hess in der Einleitung betonte, war die Kreislauffunktion für ihn "eine Gelegenheit, in die integrativen Leistungen des vegetativen Nervensystems Einblick zu nehmen". Die Durchblutungsregelung arbeitet ebenso wie die vegetative Innervation als Anpassung an die Leistung des Organs und hat eine "Dynamik höherer Ordnung". Neben der Herztätigkeit und den für das Gewebe entscheidenden Kapillarfunktionen betonte Hess auch die Tätigkeit der Arterien und Venen, die er auf ihre organspezifischen Reaktionen untersuchte. Das Herz wird als Hilfsapparat der eigentlichen kapillaren Austauschvorgänge angesehen, und die Entlastungsreflexe werden im Antagonismus zu den Nutritionsreflexen ähnlich der kardialen Vagus- und Sympathicuswirkung als Erholungs- und Leistungsfunktionen gedeutet. Auf die cerebralen Kreislaufregulationszentren ging *Hess* in dieser Monographie noch nicht genauer ein, tat dies aber später nach systematischer Auswertung seiner zentralen Reiz- und Ausschaltungsversuche im Zwischenhirn 1938 [20].

Ergotrope und trophotrope vegetative Leistungen. Sein Interesse für das vegetative System führte Hess zu einer theoretischen Systemkonzeption, die er in einer ersten weitgehend spekulativen Arbeit über "Wechselbeziehungen psychischer und vegetativer Funktionen" 1925 als Programm für experimentelle Untersuchungen veröffentlichte [7]. Im Gegensatz zur anatomischen Zweiteilung sympathischer und parasympathischer Nerven benützte Hess das Leistungskriterium für seine Einteilung in zwei funktionelle Untersysteme: 1) das ergotrope System, das die Leistungsbereitschaft animaler Funktionen fördert; 2) das trophotrope System, das für Erhaltung und Erholung der Gewebsleistungen sorgt. Im ersten Entwurf [7] hatte er das zweite auch "histotropes" und später "endophylaktisches" System [30] genannt, und solche terminologischen Eigenwilligkeiten erschwerten manchmal das Verständnis für Hess' klare Gedankenwelt in der physiologischen Forschung. Mit Cannon konnte sich Hess in einem Gespräch nicht über eine gemeinsame Konzeption und Terminologie einigen, und die Entdeckung der cholinergen und adrenergen Transmitterstoffe machten auch die alte pharmakologische Zweiteilung sympathischer und parasympathischer Drogenwirkungen problematisch. Anatomisch werden ergotrope Funktionen in der Peripherie nach Hess vorwiegend, aber nicht allein durch sympathische Nerven vermittelt. Beide Systeme gehorchen einer zentralen Steuerung, deren Erforschung sich Hess zur Lebensaufgabe machte. Im Gehirn sind beide Funktionssysteme anatomisch nicht klar unterscheidbar und offenbar noch enger antagonistisch verkoppelt. Nachdem Hess 1925 auch den Schlaf mit seiner Erholungswirkung als trophotrope Funktion bezeichnete [7], hat er die lange vernachlässigte physiologische Schlafforschung entscheidend angeregt. 1933 hat er ein übersichtliches Schema über die Funktionsverknüpfungen beider Systeme in einer Programmschrift über den Schlaf [16] gegeben. Diese Theorien konnten erst experimentell geprüft werden, nachdem seit 1934 hirnelektrische Registrierungen aus dem schlafenden Gehirn gemacht wurden und Bergers EEG auch beim Menschen die objektive Erfassung der Schlafveränderungen ermöglichte. Hirnelektirsche Schlafuntersuchungen hat Hess nur in wenigen Experimenten mit seinem Sohn an der schlafenden Katze 1950 begonnen.

Dann überließ er dieses Feld anderen. Daß im letzten Jahrzehnt die Katecholaminforschung neue Aspekte der neurochemischen Schlafregulationen eröffnete, hat *Hess* zwar interessiert, aber die biochemische Forschung lag ihm fern.

Seine hirnphysiologischen Experimente konzentrierte Hess auf das Zwischenhirn, in dem er die Integrationszentren der vegetativen Regulationen vermutete, nachdem seit 1909 nur wenige Reiz- und Ausschaltungsexperimente, vor allem der Wiener Physiologen Karplus und Kreidl auf die Rolle des Hypothalamus für das Vegetativum hingewiesen hatten.

Die Hirnstammreizung an der wachen Katze. Für die Methodik der seit 1925 durchgeführten Zwischenhirnexperimente entwickelte *Hess* eine spezielle elektrische Reizung und Elektrodenlokalisation im Gehirn [14]. Die *Berliner-Blau-Reaktion* nach Eisen-Elektrophorese von den Stahlelektrodenspitzen diente als Makrolokalisation und erste Orientierung auf den horizontal zerlegten Hinrscheiben. Zur histologischen Lokalisation verwendete er auf Rat Oskar *Vogts* abwechselnde Nissl- und Markscheidenfärbungen der horizontalen Schnittserien. Von diesen ließ er 1926 und 1937 einen fotografischen Atlas der Stammganglien und des Zwischenhirns herstellen [14].

Die von Hess entwickelte Hirnreizmethode verwendete unterbrochenen Gleichstrom, um auch markarme und marklose vegetative Fasern zu reizen, die durch die üblichen kurzen faradischen Stromstöße nicht erregt werden. Er konstruierte einen kleinen, auf der Schädeldecke verschraubten Standardhalter für sechs Elektroden, so daß die wache Katze mit ihren Zwischenhirnelektroden auf dem Versuchstisch wie in Abb. 3 frei laufen und ohne Fixation und Narkose in ihrem Verhalten untersucht und gefilmt werden konnte [14]. Dagegen erfaßte die etwa gleichzeitig von W. Ransom in Chicago systematisierte subkortikale stereotaktische Reizung narkotisierter Katzen mit der alten Horsley-Clarke-Methode lediglich grobe motorische Effekte, aber konnte über Verhaltenskorrelationen wenig oder nichts aussagen. Daher beurteilte Hess die Stereotaxie in Narkose sehr zurückhaltend. Ransom wiederum kritisierte Hess' Schlafexperimente mit ihrer langdauernden Reizung und vermutete elektrolytische Läsionen der Ausschaltung statt Reizung. So kam es nicht zu der wünschenswerten Koordination der beiden Forschergruppen von Ransom und Magoun in Chicago und von Hess in Zürich.

Zur Anatomie hatte Hess ein zwiespältiges Verhältnis. Einerseits brauchte er lokalisatorische Kontrollen der Hirnreizungen, andererseits mißtraute er dem statisch-anatomischen Denken. Obwohl seine erste Arbeit 1903 in der Anatomie begann, hat Hess schon damals anatomische Strukturen mit ihrer Funktion korreliert. Reine Strukturbeschreibung hielt er für ein be-





Abb. 3a, b. Reizexperiment an der freien Katze in *Hess'* Laboratorium 1946. a *Hess* gewöhnt die Katze durch Füttern an den Versuchstisch und die experimentelle Situation. b Kopf- und Augendeviation der Katze nach oben bei mesencephaler Reizung. – Der Versuchstisch mit Uhr, Datum und Experiment-Nummer für die Filmaufnahme war seit 1928 eine Standardordnung für jedes Experiment. Die Katze trug eine Zuleitung feiner Drähte zum Elektrodenhalter auf dem Schädel, da es noch keine

drahtlose Reizung gab. Der Institutstechniker *Jenny* sitzt rechts an der Reizapparatur, zwei Assistenten beobachten und filmen die Katze, und die Sekretärin links protokolliert. Vor 1939 machte *Hess* selbst die Filmaufnahmen der Reizeffekte

schränktes anatomisches Konzept. Er mißachtete Anatomen, die sich aus Freude am morphologischen Detail nur auf Formstudien beschränkten.

Hess sah sich die Elektrodenspitzenpositionen zunächst makroskopisch im Berliner-Blau-Bild an und ließ dann Schnittserien anfertigen [14]. Doch überließ er die eigentliche histologische Auswertung am Zell- und Faserbild anderen. Er pflegte zu sagen, es käme im Gehirn weniger auf die Kerne an als auf ihre Verbindungen, in denen sich die physiologische Ordnung zeige. Darum begrüßte er es sehr, als Weisschedel und ich 1936 im Zürcher Institut eine modifizierte Marchi-Methode zur Darstellung degenerierender Fasern von den nach der Reizung durch Koagulation zerstörten Hirnteilen einführten. Die Präparate zeigten nun die efferenten Faserverbindungen der gereizten und koagulierten Strukturen. Allerdings wurden die Grenzen der Marchi-Technik bald deutlich, da sie vorwiegend dicke markhaltige Nervenfasern darstellte, aber die im vegetativen System häufig markarmen, dünnen Fasern nur schlecht erkennbar waren. Die Silbermethoden zur Markierung synaptischer Projektionen wurden damals erst entwickelt und die modernen Enzymdarstellungen anatomischer Projektionen waren unbekannt.

Die Schlafexperimente. Hess' Demonstrationen, daß Hirnreizungen Schlaf auslösen, erregten zunächst Erstaunen, entsprachen aber genau seinem Postulat der Erholungsfunktion des trophotropen Systems von 1924/25 [7]. So wurden seine breiter angelegten Reizversuche an der freibeweglichen Katze seit 1929 vor allem durch Schlafexperimente bekannt, obwohl diese auch mit Skepsis aufgenommen wurden: Die Einwände amerikanischer Kritiker, die mit Ransom bei seinen langen Reizserien durch pulsierenden Gleichstrom elektrolytische Läsionen annahmen, konnte Hess jedoch widerlegen. Seine Experimente brachten 1927–1929 einen entscheidenden Anstoß für die von der Physiologie bis dahin vernachlässigte Schlafforschung. Hirnelektrische Korrelate des Schlafes wurden dann wenig später durch Bergers "Elektrenkephalogramm" (EEG) nachgewiesen, und seit 1935 machten amerikanische Forscher systematisch EEG-Ableitungen beim schlafenden Menschen.

Schwierigkeiten für die hirnphysiologische Deutung der Schlafexperimente von Hess lagen vor allem in der weit verstreuten unspezifischen Lokalisation der Reizpunkte, die vom medialen Thalamus bis zum Striatum reichten, und in der langen und sehr wechselnden Latenz des Schlafeffektes. Dennoch haben Hess' Untersuchungen eine neue Forschungsrichtung begründet, die mit der wenig später beginnenden Verhaltensforschung von Lorenz und v. Holst zum Experimentieren am wachen Versuchstier führte. Mit dem Schlafverhalten konnte Hess zum ersten Mal durch Zwischenhirnreizung eine geordnet ablaufende Instinkthandlung auslösen, die dem normalen Triebvorgang entspricht, der nach Ermüdung zum Ausruhen führt [8, 37]. 1943 konnte er einen umgekehrten Aktivierungseffekt, die "affektive Abwehrreaktion" nach Hypothalamusreizung [26], hinzufügen. Erst viel später wurde das Konzept der sog. unspezifischen Aktivierung der Hirnrinde im EEG nach Reticularis-Reizung im Mittelhirn durch Moruzzi und Magoun 1949 entwickelt. Da Hess sich zunächst auf Reizexperimente oberhalb des Mittelhirns beschränkte und die späteren Reticularis-Reizungen anderer Autoren meist am narkotisierten Tier erfolgten, dauerte es ein bis zwei Jahrzehnte, bis die Befunde von *Hess* mit den hirnelektrischen Ergebnissen und dem Konzept eines unspezifischen aufsteigenden Aktivierungssystems koordiniert werden konnten.

Seit 1960 gelang es dann Jouvet, Moruzzi und ihren Mitarbeitern, auch die Beteiligung tieferer rhombencephaler Hirnteile an der Schlafregulation nachzuweisen. Diese Reiz- und Ausschaltungsexperimente in Pons und Oblongata und die Entdeckung der vom Hirnstamm zum Cortex aufsteigenden Systeme mit verschiedenen chemischen Überträgerstoffen zeigten, daß die Schlafregulationen viel komplizierter sind, als man in den 30er Jahren annahm. Im Prinzip hat jedoch Hess' Theorie, wonach der Schlaf eine trophotrope Ruhigstellung des Großhirns durch vom Hirnstamm aufsteigende dämpfende Impulse ist, auch heute noch ihre Gültigkeit.

Die Motorik. Hess' Interesse für die Bewegungsphysiologie begann schon in seiner ophthalmologischen Zeit, als er ein Schema zur Erfassung der Augenmuskellähmungen entwarf [3]. Die Augenmotorik war für ihn ein Idealfall, der die Bewegungsordnung in den drei Raumebenen mit drei Paaren antagonistischer Muskeln exakt darstellen ließ. Dreißig Jahre später kam er mit einer präkybernetischen Arbeit über die Motorik als Organisationsproblem [24] zur sensomotorischen Koordination zurück. Er konstruierte Blockschaltungen als Modelle der okulomotorischen Beherrschung des Blickfeldes mit einfacher und doppelter Mannigfaltigkeit, die für jede Ausgangsstellung eine Bewegungsfreiheit nach allen Richtungen entsprechend der Gesichtsfeldlokalisation ermöglichten. Diese erweiterte er für die Körpermotorik mit Überwindung der Gravitation durch propriozeptive Regulation. Nach seinen Worten sollten bei der Bewegung Rezeptorsysteme zur Störausschaltung und Muskelinnervation als "zielorientiert gruppierte Kräfte fehlerfrei zum Ziel führen". Diese Bewegungsregulation muß bei jeder neuen Stellung die Muskelzugwirkung verändern. Er postulierte im Gehirn eine Parallelschaltung der Raumwahrnehmung mit der Bewegungskoordination. Seine präkybernetischen Modelle verwendeten Vektorwirkungen für die richtungsmäßige Impulssummation. Gleichzeitig demonstrierte er mit seinen Hirnstammreizungen die Existenz richtungsspezifischer Substrate im Zwischen- und Mittelhirn der Katze [22, 35]. So konnten seine experimentellen Befunde die Realität der von vielen als spekulativ empfundenen Bewegungsorganisation nachweisen.

Seine Bewegungstheorien hat *Hess* zwei Jahre später für die *menschliche* Ziel- und Stützmotorik durch ein lebendes Modell für die praktische Bewegungsleistung ergänzt [27]. Die Wechselwirkung von Haltung und Bewegung wurde durch drei Personen dargestellt. Der Träger repräsentiert Skelet und Schwerkraft-kompensierende Muskulatur, der Stützer die Halteinnervation rumpfnaher Muskeln und der Springer die dynamisch-ballistische Muskelaktion. Bei der aufrechten Haltung des Menschen wird die Stützmotorik vor allem zur Kompensation von Schwerkraftwirkungen eingesetzt. Damit die Zielbewegung gelingt, muß die Stützfunktion frühzeitig aktiviert werden. Diese Antizipation bedeutet ein Primat der Haltung für die Bewegungsbereitschaft. Träger und Stützer müssen den Moment des Sprunges wissen, Gewicht und Absprungkraft fühlen und sich an die Haltungsveränderung anpassen. Die Zielbewegung mißlingt, wenn der ungestützte Träger vom Absprung nicht orientiert wird. Prinzip der Stütz- und Zielmotorik sind demnach Bereitschaftshaltung mit regelnder und steuernder Rückmeldung entsprechend der in den folgenden Jahren entwickelten Biokybernetik [42, 43].

*Hess*' ungewöhnliche Terminologie von 1943, "teleokinetische Motilität" für die Zielmotorik und "ereismatische Motilität" für die Stützmotorik [27], erschwerten zunächst das Verständnis dieses Modellversuchs, so daß *Hess* ihn 22 Jahre später noch einmal mit Filmbildern zusammenfassend dargestellt hat [42, 43].

Die weitere Ausarbeitung überließ Hess anderen. Als seine Schüler Wyss und Koella die Labyrinthreflexe und ich die visuell-vestibuläre Koordination mit Registrierung der Augenbewegungen untersuchten, zog er sich zurück, als die Ergebnisse zu kompliziert für Modellbilder wurden. Doch verfolgte Hess die Reafferenz-Konzepte v. Holsts und andere Regelmodelle, die Blickbewegung und Wahrnehmung durch entsprechende biokybernetische Schaltbilder erklären sollten, mit wachem Interesse.

Die Psychophysiologie. Sein letztes Buch [39], das Hess nach seiner Emeritierung schrieb, war über Jahrzehnte geplant. Es sollte zunächst den Titel "Psychophysiologie" erhalten, den er 1962 in Psychologie in biologischer Sicht änderte. Seine Korrelation von psychischen und neuralen Funktionen hatte ihren frühen Ursprung in der Schrift von 1925 über psychische und vegetative Funktionen [7]. Das Psychologiebuch ist oft mißverstanden worden, besonders die Ausführungen über unbekannte "psychische Kräfte".

Im Gespräch erläuterte Hess die seelische Kraft und ihre unbekannte Natur viel klarer als in dem Buch: Er zog eine Parallele zur Schwerkraft in der Physik, deren Wesen auch niemand kennt, aber deren Wirkungen man beobachten und exakt messen kann. So meinte Hess, sollten auch psychische Kräfte in ihren Auswirkungen zu erkennen und meßbar sein, obwohl ihre Natur völlig unklar bleibt. Ein Rezensent glaubte, das Buch enthalte unwissenschaftliche Vorstellungen von psychischen Kräften ohne Korrelation mit Hirnfunktionen, obwohl Hess mehrfach betont hat, daß psychische Funktionen nur auf der Grundlage neuronaler Hirnprozesse möglich sind. Eine persönliche Diskussion sollte diese Mißverständnisse vor der 2. Auflage beseitigen. Um die Verbindung mit der modernen Forschung zu erhalten, organisierte *Hess* ein kleines Symposion, bei dem Freunde und Schüler neuere Befunde mit informationstheoretischen Fragen besprachen. Diese Diskussion erschien in der 2. Auflage [39].

Über allgemeine Fragen von Hirnfunktion und Bewußtsein und seine Konzeption der psychischen Kräfte korrespondierte *Hess* zuletzt mit dem Pharmakologen H. *Fischer*. Teile dieses Briefwechsels erschienen in englischer Übersetzung 1973 kurz nach seinem Tode. Darin wird eine Lösung dieser Fragen auf empirischer Basis abgelehnt und philosophische und religiöse Gedanken werden nur als mögliche Annäherungen diskutiert. Zu den Grenzen unseres Wissens sagt *Hess:* "Wir müssen uns zufrieden geben, daß noch so vieles in der Welt existiert und vor sich geht, das unserem Verständnis entzogen ist. Darüber hinaus halte ich ein bescheidenes Schweigen als die beste Einstellung zu den aufgeworfenen Fragen."

Nebenwege der Forschung. Die rhythmischen Spontankontraktionen der Arterien verschiedener Organe und ihre humorale Beeinflussung durch vegetativ aktive Stoffe haben *Hess* über viele Jahre interessiert. Die mehr oder weniger spezifischen Veränderungen durch Adrenalin, Acetylcholin und andere Stoffe wurden von zahlreichen Doktoranden untersucht.

Große Verdienste hatte Hess auch für die meteorologisch-physiologische Forschung. Er gründete die Höhenstation Jungfraujoch und verwendete viel Zeit für die Organisation und Einrichtung, bis er sie an von Muralt, Bern, abgab, der die von Hess ehrenamtlich durchgeführte Leitung dann in eine amtliche Direktorenstelle verwandelte. Nach frühen Anregungen für die Wetterforschung durch seinen Vater hat Hess manche Arbeit auf diesem Gebiet veranlaßt, aber war selbst nicht auf dem meteorologisch-biologischen Gebiet tätig, das auch heute noch ungeklärt ist.

### Die Wirkungen seines Forscherlebens

Der akademische Lehrer. In den ersten Jahren seiner Institutsleitung 1917–1924 widmete *Hess* viel Zeit dem Ausbau des physiologischen Unterrichts, bevor er 1924 seine Hirnforschungsarbeit begann. Er verbesserte den physiologischen Kurs mit experimentellen Demonstrationen und führte den Schmalfilm für Unterricht und Forschung ein. Die *Filmdemonstration* blieb für *Hess* seitdem wichtigstes Instrument der Demonstration und Dokumentation. Die Hauptvorlesung für alle Gebiete der Physiologie hielt er jedes Semester, auch in späteren Jahren, in denen die experimentelle Arbeit seine ganze Kraft benötigte. Seine Reizexperimente filmte er

viele Jahre selbst und erst nach 1945, als er das Personal des Institutes vergrößern konnte, übernahmen Mitarbeiter die Filmarbeit, wie Abb. 3b zeigt. Die Prinzipien der Motorik hat *Hess* vorwiegend durch Filmanalyse und Modellexperimente gefördert. Auch in den 30er Jahren, als seine Methode der Hirnreizung bei der freilaufenden Katze seine ganze Arbeitskraft in Anspruch nahm, hat er sich nicht vom studentischen Unterricht zurückgezogen, sondern grundlegende Experimente oft selbst vorgeführt wie in Abb. 2b.

Der Sprecher und Autor. Hess war ein guter Redner, und seine Sprache als akademischer Lehrer war klar und einprägsam. Auch in der Diskussion traf er mit wenigen, richtigen Worten den Kern der Sache. Als er auf der deutschen Physiologentagung 1960 wieder einmal die mangelnde physiologische Hirnforschung in den deutschen Instituten monierte, sagte er: "Wenn Sie es nicht machen, werden die Pharmakologen die Hirnphysiologie übernehmen, aber die Neuropharmakologie wird funktionell wichtige Zusammenhänge übersehen". Wenn Hess dann die gleichen Gedanken in schriftliche Form brachte, wurden die einfach erfaßten Prinzipien zwar mit konkreten Beispielen illustriert, aber sein systematisches, sich mit allen Nebenwirkungen und Widersprüchlichkeiten auseinandersetzendes Denken verführten ihn zu langen Erklärungen und komplizierten Formulierungen. Seine Terminologie war oft eigenwillig und erschwerte die Annahme seiner klaren Konzeptionen durch vermeintliche Ausdrucksverbesserung: z.B. die Umbenennung des anfangs "histotropen" [7] dann "trophotropen" vegetativen Nervensystems [16] in ein "endophylaktisches" System [30], das als Antagonist des ergotropen Systems funktioniert. Auch die Bezeichnung der Zielmotorik als "teleokinetisch" und der Stützmotorik als "ereismatisch" war der Anerkennung dieser wichtigen Leistungsanalyse der Bewegung und Haltung nicht günstig.

Bei Abfassung seiner Manuskripte gab Hess sich große Mühe und schrieb sie immer wieder um. Nach der Emeritierung war er ohne Sekretärin, und in Ascona half ihm seine Frau bei allen späteren Arbeiten unermüdlich an der Schreibmaschine. Zur Begründung mangelnder sprachlicher Eleganz sagte Hess, für jemand, der in der Umgangssprache Schwyzerdütsch rede, sei das "Schriftdeutsch eine Fremdsprache". Aber dies hat mich nie überzeugt, weil Hess auch im hochdeutschen Vortrag spontan klar und präzise sprach. Es war seine Gründlichkeit und Gewissenhaftigkeit, die den einfachen Satz komplizierte. So habe ich bei allen strittigen Fragen lieber persönlich mit ihm gesprochen, als korrespondiert oder alte Publikationen nachgelesen. 1958 wollten Hassler und ich, als seine Schüler, Hess' Konzeption der Ziel- und Stützmotorik im amerikanischen Handbuch der Physiologie darstellen: Bei unserem Besuch in Zürich gab uns Hess an einem Nachmittag mündlich mit einem alten Film ein klares Exposé, das aus der Lektüre seiner zehn vorangehenden Arbeiten nur mühsam zu abstrahieren war. Der Film ließ sich in einer Schemazeichnung zusammenfassen, die *Hess* dann korrigierte und bestätigte: Haltung, Stütze und Zielbewegung war in einem Drei-Personen-Experiment anschaulicher dargestellt als es eine lange Beschreibung vermochte (vgl. S. 14).

Jede Begegnung mit *Hess* war noch bis in die letzten Altersjahre interessant und anregend, denn er blieb noch als 80jähriger produktiv mit neuen Ideen und diskutierte gern auch über neue Gebiete der Verhaltensforschung und Biokybernetik. So habe ich die früheren Zürcher Besuche, zu denen er nach kurzer Anfrage einlud, und die selteneren späteren in Ascona in schönster Erinnerung.

Die Hess-Schule und persönliche Erinnerungen. Während der vier Jahrzehnte seiner Tätigkeit im Zürcher Physiologischen Institut hatte Hess zahlreiche Mitarbeiter, die auf verschiedenen Gebieten der Physiologie arbeiteten und vorwiegend Schweizer Lehrstühle besetzten. Von den älteren Mitarbeitern nenne ich F. Verzar in Basel, A. Fleisch in Lausanne und O.A.M. Wyss, der sein Nachfolger in Zürich wurde und vorwiegend Themen der Neurophysiologie und Atmungsphysiologie bearbeitete, ferner E. Rothlin und W.P. Koella, die neurophysiologische und neuropharmakologische Forschungen in der Schweizer Pharmaindustrie förderten. Spätere neurophysiologische Schüler waren Marcel Monnier, der aus der Neurologie kam und nach einer kurzen Tätigkeit in Genf den Basler Lehrstuhl als Nachfolger Verzars erhielt und die Physiologie und Neurochemie des Schlafs bearbeitete, und R.W. Hunsperger, der die cerebralen Reizversuche über die affektive Abwehrreaktion fortsetzte und die Verbindungen der hypothalamischen Strukturen mit dem Rhinencephalon und dem Mandelkern klärte, Sandro Bürgi, der die anatomische Auswertung von Hess' Experimenten Ende der 30er Jahre übernahm, wurde Neurologe, aber hielt die Verbindung mit dem Institut aufrecht. Der Sohn Rudolf Hess jun. wechselte von der Physiologie zur klinischen Elektroenzephalographie. Konrad Akert war der letzte und in der Forschung aktivste Schüler, mit dem Hess zahlreiche Arbeiten 1945-1952 schrieb und der sein Werk mit dem Aufbau des Zürcher Hirnforschungsinstitutes fortsetzte. Nach mehrjährigen Arbeiten in verschiedenen physiologischen und anatomischen Instituten der USA kam Akert nach Zürich zurück, als Hess, enttäuscht von dem Fortgang der Hirnforschung im Physiologischen Institut die Gründung eines Hirnforschungsinstitutes erreicht hatte, das auch die hirnanatomische Abteilung v. Monakows fortsetzen sollte. Dieses selbständige Institut übernahm Akert, richtete Abteilungen für Neurophysiologie, Neuroanatomie und Neurochemie ein und konnte so die von Hess inaugurierte Forschungsrichtung fortsetzen und erweitern.

Von ausländischen Mitarbeitern erwähne ich den Holländer W.O.C. *Magnus*, Sohn des Utrechter Pharmakologen, den Schweden B. *Andersson*, den Italiener C. *Bartorelli* und E. *Weisschedel*, R. *Hassler* und mich selbst aus Deutschland.

Ich habe Hess über fast 40 Jahre gekannt, und seit meiner ersten Tätigkeit in seinem Institut 1936/37 trafen wir uns regelmäßig. Der große Altersunterschied von drei Jahrzehnten – Hess war damals 55 und ich 25 Jahre alt – bedeutete keine Trennung. Außer dem Lehrer-Schüler-Verhältnis entwickelte sich in den nächsten Jahrzehnten durch meine Zuwendung zur Elektrophysiologie, die Hess selbst nicht betrieb, ein Verhältnis gegenseitiger Anregung für die hirnphysiologische Forschung. Als Hess 1938 mein Laboratorium in Freiburg besuchte und ich ihm die großen, bilateral-synchronisierten EEG-Wellen beim kleinen epileptischen Anfall zeigte, rief er spontan: "Das ist die vegetativ-trophotrope Beeinflussung des Cortex aus dem Zwischenhirn". Eine ähnliche subcorticale Beeinflussung des Cortex war bei den langsamen Schlafpotentialen anzunehmen.

Selbst die Kriegsjahre brachten keine vollständige Unterbrechung unserer wissenschaftlichen Diskussion. *Hess* kam 1941 nach Frankfurt, um seine neuen Motorikuntersuchungen in den drei Raumrichtungen nach Hirnstammreizung bei *Kleist* zu demonstrieren, in dessen Lazarett ich damals arbeitete, und diskutierte klinisch-neurologische Korrelate der Wendeund Drehbewegung seiner Katzen mit uns. In den schwierigen Nachkriegsjahren, als wir in der Besatzungszeit noch ohne wissenschaftlichen Kontakt mit dem Ausland blieben, lud *Hess* hirnanatomisch interessierte deutsche Mitarbeiter wie *Weisschedel* und *Hassler* ein, an der Auswertung seiner Ausschaltungsversuche teilzunehmen [32, 35].

Unsere Reiz- und Ausschaltungsexperimente an der Substantia nigra, die wir auf Wunsch von Spatz bei Hess 1936/37 in Zürich machten, brachten nicht die erhofften Ergebnisse, da die Marchi-Methode für die Degeneration der feinen Fasern nicht ausreichte und die modernen Silberdegenerationsmethoden und die Dopamintransmitter noch nicht bekannt waren. So waren wir bei der Deutung des flachen EEG im Isocortex und der Thetawellen im Allocortex nach Sinnesreizen und Hirnstammreizungen, die ich mit Weisschedel und Kornmüller 1937 in Berlin untersuchte, anatomisch zu skeptisch gegen die von Hess postulierten ascendierenden Cortexprojektionen. Damit versäumten wir, das thalamo-retikuläre System zu erkennen, auf das erst 1949 Moruzzi und Magoun aufmerksam machten, und Hirnstamm-Cortex-Beeinflussungen, die jetzt für die Katecholamintransmitter anerkannt sind.

Jedes physiologische Gespräch mit Hess brachte neue Anregungen, Kritik und Aufmunterung. Anfangs begegnete er unseren Mikroelektrodenstudien mit Zurückhaltung, dann begrüßte er sie, riet aber mehr zu einer Analyse der Motorik, die ihn seit 1940 vor allem interessierte. Zu dieser Arbeit kam ich erst nach seinem Tode, habe aber in den letzten Jahren seines Lebens noch einiges mit ihm diskutieren können. Nur eine Enttäuschung mußte ich *Hess* bereiten: Seinem Wunsch, den Lehrstuhl für Neurologie in Zürich zu übernehmen, zu dem mich die Fakultät 1955 berief, konnte ich nicht folgen, nachdem einige zunächst gegebene Zusagen nicht erfüllt wurden.

Das letzte Jahrzehnt. 15 Jahre nach seiner Emeritierung, als fast alle Zürcher Freunde gestorben waren und *Hess* wegen seiner zunehmenden Schwerhörigkeit nicht mehr zu wissenschaftlichen Tagungen reisen konnte, überließ er seine Bibliothek dem Institut und zog 1967 ohne Bücher und Sonderdrucke von Zürich in sein kleines Sommerhaus nach Ascona. Dort lebte er mit seiner Frau in Ruhe für die letzten sechs Jahre. Beziehungen zur Umwelt behielt er durch lebhafte Gespräche mit besuchenden Freunden und seine dauernde Freude an der Pflege seines Gartens.

Im letzten Jahrzehnt verhinderte seine Altersschwerhörigkeit alle Tagungsdiskussionen, aber in der Einzelunterhaltung blieb Hess aufnahmefähig und anregend wie in jungen Jahren. Die letzte Tagung, an der er noch aktiv teilnahm, war das von seinem Schüler Akert 1964 organisierte Schlafsymposion in Zürich, in dem er einen kurzen Überblick über seine Konzeption des Schlafes als einer aktiven Leistung des gesamten Organismus gab. In Ascona habe ich ihn seit 1967 fast jedes Frühjahr gesehen und konnte ihm 1971 zum 90. Geburtstag die Glückwünsche der deutschen Physiologen überbringen, von deren Gesellschaft er dann zum Münchener Internationalen Physiologenkongreß die Johannes-Müller-Medaille erhielt. Damals zeigte uns Hess den Friedhof auf dem benachbarten Berg, auf dem er begraben sein wollte. Während er sich im August 1973 an der sommerlichen Fülle seines geliebten Gartens freute, kam ein friedlicher Tod durch Herzversagen und beendete sein rastloses Forscherleben.

### Literatur

Das Literaturverzeichnis von W.R. *Hess* umfaßt über 304 Arbeiten, die im Nachruf der Wiener Akademie [45] vollständig verzeichnet sind. Ich bringe daher aus der von 1903–1973 reichenden Publikationsliste nur einige wichtige Arbeiten und die zusammenfassenden Monographien in chronologischer Ordnung (A.) und eine Liste der Nachrufe für *Hess* (B.). Die Arbeiten seines Institutes für die Hirnstammphysiologie sind im Dokumentationsatlas "Hypothalamus und Thalamus" [37] zusammengestellt. Auch die "Psychologie in biologischer Sicht" [39] enthält einige thematisch geordnete Institutsarbeiten.

A. Wesentliche Arbeiten und Monographien von W.R. Hess 1903-1966

- 1) 1903 Eine mechanisch bedingte Gesetzmäßigkeit im Bau des Blutgefäßsystems. Wilhelm Roux' Arch Entwickl. Mech 16:632-641
- 2) 1907 Die Bestimmung der Viskosität des Blutes. Münch Med Wochenschr 54/2: 2225-2229
- 3) 1908 Eine neue Untersuchungsmethode bei Doppelbildern. Arch Augenheilkd 62:233-238
- 4) 1914 Das Prinzip des kleinsten Kraftverbrauches im Dienste hämodynamischer Forschung. Habilitationsschrift 1913. Arch Anat Physiol. Physiol Abt (Lpz) 1914:1-62
- 5) 1923 Die Regulierung des peripheren Blutkreislaufes. Ergeb Inn Med Kinderheilkd 23:1-65
- 6) 1924 Die reflektorische Ruhigstellung schmerzender Körperteile. Pfluegers Arch Ges Physiol 203:539-541
- 7) 1924/25 Über die Wechselbeziehungen zwischen psychischen und vegetativen Funktionen. Schweiz Arch Neurol Neurochir Psychiatr 15:260-277; 16: 36-55 und 285-306; und als Monographie Orell Füssli Zürich
- 8) 1927 Stammganglien-Reizversuche. Ber Ges Physiol 42:554–555
- 9) 1929 Hirnreizversuche über den Mechanismus des Schlafes (Juni 1928). Arch Psychiatr Nervenkr 86:287–292
- 10) The mechanism of sleep. Am J Physiol 90:386–387
- 11) Lokalisatorische Ergebnisse der Hirnreizversuche mit Schlafeffekt (Juni 1929). Arch Psychiat Nervenkr 88:813-816
- 12) 1930 Die Regulierung des Blutkreislaufes, gleichzeitig ein Beitrag zur Physiologie des vegetativen Nervensystem. Thieme, Leipzig
- 13) 1931 Die Regulierung der Atmung. Thieme, Leipzig
- 14) 1932 Die Methodik der lokalisierten Reizung und Ausschaltung subkortikaler Hirnabschnitte. Thieme, Leipzig
- 15) The autonomic nervous system. Lancet 223:1199–1201, 1259–1261
- 16) 1933 Der Schlaf. Klin Wochenschr 12:129–134
- 17) 1935 Das physiologische Zusammenspiel von Kreislauf und Atmung. Verh Dtsch Ges Kreislaufforsch 8:9–31
- 18) 1936 Die Rolle des Vagus in der Selbststeuerung der Atmung. Pfluegers Arch Ges Physiol 237:24-39
- 19) Mit Wyss, O.A.M.: Die Analyse der physikalischen Atmungsregulierung an Hand der Aktionsstrombilder des Phrenicus. Pfluegers Arch Ges Physiol 237:761-770
- 20) 1938 Das Zwischenhirn und die Regulation von Kreislauf und Atmung. Thieme, Leipzig
- 21) 1940 Untersuchungen über das Ursprungsgebiet des primären Atmungsrhythmus. Pfluegers Arch Ges Physiol 243:259–282
- 22) Zwischenhirn und Motorik. Pfluegers Arch Ges Physiol 243:634-650

Walter R. Hess (1881–1973)

- 23) 1941 Mit Wyss O.W.M.: Charakter der im Zwischenhirn ausgelösten Bewegungseffekte. Pfluegers Arch Ges Physiol 244:767–786
- 24) Die Motorik als Organisationsproblem. Biol Zbl 61:545–572
- 25) 1942 Biomotorik als Organisationsproblem (I u. II). Naturwissenschaften 30: 441-448, 537-541
- 26) 1943 Mit Brügger, M.: Das subkortikale Zentrum der affektiven Abwehrreaktion. Helv Physiol Pharmacol Acta 1:33-52
- 27) Teleokinetisches und ereismatisches Kräftesystem in der Biomotorik. Helv Physiol Pharmacol Acta 1:C62–C63
- 28) 1944 Das Schlafsyndrom als Folge diencephaler Reizung. Helv Physiol Pharmacol Acta 2:305-344
- 29) 1946 Mit Bürgi, S., Bucher, V.: Motorische Funktion des Tektal- und Tegmentalgebietes. Monatsschr Psychiatr Neurol 112:1–52
- 30) 1948 Die funktionelle Organisation des vegetativen Nervensystems. Schwabe, Basel
- 31) 1949 Das Zwischenhirn. Syndrome, Lokalisationen, Funktionen. Schwabe, Basel.
   (2. erw Aufl 1954)
- 32) Mit Weisschedel, E.: Die höheren Reflexzentren der regulierten Körperhaltung. Helv Physiol Pharmacol Acta 7:451–469
- 33) 1950 Motorik und Zwischenhirn. Arch Psychiatr Nervenkr 184:267-270
- 34) 1952 Mit Akert, K., McDonald, D.A.: The functions of the orbital gyri in cats. Brain 75:244-258
- 35) 1954 Mit Hassler, R.: Experimentelle und anatomische Befunde über die Drehbewegungen und ihre nervösen Apparate. Arch Psychiatr Nervenkr 192: 488-526
- 36) Diencephalon, autonomic and extrapyramidal functions. Grune & Stratton, New York
- 37) 1956 Hypothalamus und Thalamus. Experimentaldokumente (Monographie-Atlas). Thieme, Stuttgart (erw Aufl 1968)
- 38) 1957 Die Formatio reticularis des Hirnstammes im verhaltensphysiologischen Aspekt. Arch Psychiat Nervenkr 196:329-336
- 39) 1962 Psychologie in biologischer Sicht. Thieme, Stuttgart (2. erw Aufl 1968)
- 40) 1963 From medical practice to theoretical medicine: an autobiographic sketch. Perspect Biol Med 6:400-423
- 41) 1964 The biology of mind. Univ Chicago Press, Chicago
- 42) 1965 Cerebrale Organisation somatomotorischer Leistungen. I. Physikalische Vorbemerkungen und Analyse konkreter Beispiele. Arch Psychiatr Nervenkr 207:33-44
- 43) 1966 Cerebrale Organisation somatomotorischer Leistungen. II. Experimentelle Untersuchungen über die zentrale Koordination der Motorik. Arch Psychiatr Nervenkr 208:209–233

### B. Nachrufe

- 44) Akert K (1974) Walter Rudolf Hess (1881–1973). Obituary. Brain Res 68:5–8
- 45) Auerswald W (1975) Walter Rudolf Hess. Nachruf (mit Schriftenverzeichnis). Almanach Österr Akad Wissenschaft 124:414–433
- 46) Caspers H, Hess R, Kugler J, Petsche O, Struppler A (1974) Nachruf auf W.R. Hess. EEG EMG 5:139-141
- 47) Wyss OAM (1973) Walter Rudolf Hess 17.3.1881-12.8.1973. Die Selbständigkeit der Physiologie. Schweiz Med Wochenschr 103:1758-1759
- 48) Wyss OAM (1973) Zum Hinschied von Walter Rudolf Hess. Neue Zürcher Zeitung 382:20. August 1973

## Neural Organisation and Control of the Baroreceptor Reflex

### K. MICHAEL SPYER \*

### Contents

Abbreviations					
1	Introduction.				
2	2 The Projection of Sinus and Aortic Baroreceptors				
	2.1.1 2.1.2 2.1.2.1 2.2 2.2.1 2.2.2 2.2.2.1 2.2.2.2 2.2.3 2.2.3.1 2.2.3.2 2.2.3.3	SynapseNeuroanatomical StudiesElectrophysiological StudiesAntidromic ActivationBaroreceptor Reflex PathwayNeuroanatomical StudiesElectrophysiological StudiesEvoked PotentialsSingle Unit StudiesBaroreceptor StimulationPulse-synchronous Activity"Cardiovascular" NeuronesSpecific Baroreceptor Inputs to the Brainstem and Hypothalamus	25 27 28 28 31 32 34 35 38 41 41 43 44		
3	Barorece 3.1 3.2 3.3 3.3.1 3.3.2 3.3.3 3.3.3 3.3.3 3.4 3.5 3.6 3.6.1 3.6.1.1 3.6.1.2 3.6.1.3 3.6.2 3.6.3	eptor Control of Preganglionic Autonomic Activity	53 53 55 57 58 61 62 62 65 66 71 73 74 75 75 77 77 77 80 83		

<sup>\*</sup> Department of Physiology, Royal Free Hospital School of Medicine, Pond Street, London NW3 2QG, Great Britain

4	Modula	ation of the Baroreceptor Reflex	83
	4.1	Interactions Between Carotid Sinus and Aortic Arch Baroreceptor	
		Inputs	84
	4.2	Cardiac Component of the Baroreceptor Reflex	86
	4.2.1	Facilitation	86
	4.2.2	Inhibition	88
	4.3	Respiratory Influences on the Baroreceptor Vagal Reflex	92
	4.4	Total Central Inhibition of the Baroreceptor Reflex	97
	4.4.1	The Defence Reaction	97
	4.4.2	The Inferior Olive	101
5	Conclue	ding Remarks	102
R	eferences	§ <i> </i>	104

### Abbreviations

AN	Aortic nerve	5HT	5-hydroxytryptamine
BVM	Preganglionic bronchomotor	IC	Intercostal
	neurone	IML	Intermediolateral cell column
С	Cervical spinal cord	IMM	Intermediomedial cell column
CVM	Preganglionic cardiac vagal	LRN	Lateral reticular nucleus
	motoneurone	NA	Nucleus ambiguus
DLF	Dorsolateral funiculus	NPR	Paramedian reticular nucleus
DLH	Di-homocysteic acid	NTS	Nucleus of the tractus solitarius
DNV	Dorsal motor nucleus of the	SN	Sinus nerve
	vagus	Т	Thoracic spinal cord
ECG	Electrocardiogram	TS	Tractus solitarius

### 1 Introduction

The baroreceptors of the aortic arch and the carotid sinuses participate in the homeostatic control of the cardiovascular system through a regulation of arterial blood pressure. The basic properties of the baroreceptors and the input-output characteristics of the reflex have been reviewed (*Heymans* and *Neil* 1958; *Kezdi* 1967; *Kirchheim* 1976). The years that span these three major reviews have seen a tremendous advance in our understanding of the central nervous control of the circulation, yet this work appears to have made very little impact on the analysis of neural control of the reflex presented in the latest of these reviews. This is likely to reflect the dominance of the concept of a medullary "vasomotor" centre through which central and peripheral inputs exert their influence on the cardiovascular system (*Bayliss* 1923; *Alexander* 1946). Accordingly, since the baroreceptor input has an important role in cardiovascular control, its role has been restricted to a control of this hypothetical "centre". Since the foundations on which both the original concept and the anatomical location of

the centre are based have been questioned (*Peiss* 1965; *Hilton* 1966, 1975; *Downman* 1972; *Smith* 1974), it becomes important to draw attention to the basic neuronal circuitry that mediates the baroreceptor reflex.

This article will endeavour to review the neurophysiological and neuroanatomical studies that have attempted to investigate the pathway within the central nervous system taken by the baroreceptor reflex. The last decade in particular has seen a rapid development in the range and specificity of neurophysiological and neurohistological techniques so that a general framework has emerged concerning the nature and extent of this pathway (Sect. 2). Secondly, an attempt will be made to summarise the available material on the baroreceptor inputs to and control of preganglionic autonomic neurones, since by analogy with the somatic nervous system they may well be a site of integration (Sect. 3). It will be necessary at this time to review the control of sympathetic and vagal preganglionic activity from other brainstem sites and the relationship of baroreceptor inputs to these controls because this may well supplement and, it is hoped, complement, the data reviewed in Sect. 2. Finally, an attempt will be made to evaluate the modifications of the reflex which may be evoked centrally or peripherally (Sect. 4). This has already received extensive review (see for example Korner 1971) since it has often been suspected that a central resetting of the reflex might play a role in the development of hypertension (Kezdi 1967). The present account will concern itself only with those modifications which have been investigated neurophysiologically. This restricted selection may, however, prove adequate, as an analysis of such a limited brief indicates that certain modifications result from a limited range of neuronal mechanisms.

Throughout, only a limited effort will be directed to neuropharmacological studies on the performance of the baroreceptor reflex, since there are few examples of direct investigation, and the expansive literature on the pharmacology of cardiovascular control has received extensive review elsewhere (for example *Day* and *Roach* 1974; *Haeusler* 1976; *Prichard* 1978).

### 2 The Projection of Sinus and Aortic Baroreceptors

# 2.1 Termination of Sinus and Aortic Nerves: Location of the First Synapse

In establishing the central pathway taken by the baroreceptor reflex, the location of the first synapse is of particular importance. Standard neuroanatomical and neurophysiological techniques are available to investigate the projections of primary afferent sensory fibres, but although these have been applied to the problem of the baroreceptor input to the brain, until recently controversy had remained regarding the localisation of even the first relay station.

There are many reasons for this. Firstly, the afferent fibres originating from the baroreceptors do not form a homogeneous group; they range from unmyelinated to medium-sized myelinated fibres (de Castro 1928; Douglas and Ritchie 1956; Douglas and Schaumann 1956; Douglas et al. 1956; Fidone and Sato 1969). Furthermore, in the case of those innervating the carotid sinus, they are carried in the sinus nerve, mixed with afferent fibres innervating the chemoreceptors of the carotid body which have diameters within the same spectrum as baroreceptor afferents (see de Castro 1928; Heymans and Neil 1958; Fidone and Sato 1969). This means that it is impossible to activate preferentially either baroreceptor or chemoreceptor afferent fibres on electrical stimulation of the sinus nerve, although this has sometimes been claimed (see for example de Groat and Lalley 1974). Furthermore, the sinus nerve is a branch of the IXth cranial nerve, which contains afferents from a wide range of areas such as the pharynx, tonsil and posterior part of the tongue, in addition to efferent fibres. The sinus nerve itself contains two groups of efferent fibres: postganglionic sympathetic fibres and a group of nonsympathetic and predominantly unmyelinated fibres which relay from an unknown site in the central nervous system, i.e. true sinus nerve efferents (see Fidone and Sato 1969).

In the case of aortic baroreceptor afferents similar problems arise. In the rabbit, there are no chemoreceptors in the aortic arch (*Heymans* and *Neil* 1958), and the aortic nerve (depressor nerve) is usually independent of the cervical vagus as far as the nodose ganglion. This fortunate anatomical arrangement is not repeated in other species, and in all species the intracranial vagal rootlets contain a heterogeneous mixture of afferents which innervate a wide range of thoracic receptors, both cardiovascular and respiratory, in addition to aortic arch baroreceptors and chemoreceptors and of course efferent axons (for references see *Paintal* 1972, 1973). The cervical vagus may also contain afferents innervating a group of baroreceptors in the carotid artery but independent of the carotid sinus (*Green* 1965).

Neuroanatomical studies on the projection of the IXth and Xth cranial nerves, which have traced degenerating axons and terminals following transection, whilst not providing data concerning the projection of baroreceptors in isolation, do provide a broad estimate of the limits within which these afferents must distribute themselves (see *McAllen* et al. 1979). With the recent application of neurophysiological techniques and specifically using antidromic mapping, it has been possible to obtain a reliable picture of the termination of sinus and aortic nerve fibres. To this it has been necessary to add unit recording coupled with natural afferent stimulation in order to identify specific baroreceptor projections.

### 2.1.1 Neuroanatomical Studies

The projections of aortic and carotid sinus baroreceptors have been inferred from tracing degenerating axons and terminals after sectioning the cranial rootlets of the Xth and IXth nerves respectively. An early study using the Marchi technique, which traces only degeneration in myelinated fibres, showed that following vagal rhizotomy in the cat, degenerating fibres were restricted to the area around the ipsilateral tractus solitarius (TS) of the medulla (Foley and Dubois 1934). They described that the vagal fibres descended through the tractus giving off collaterals to the main nuclear regions – the medial, circumferential and commissural nuclei – in much the same way as described for IXth and Xth afferents by Cajal (1909). Ingram and Dawkins (1945) also described a similar distribution in the cat, although they also showed that some fibres decussated at obex level to terminate within the contralateral commisural nucleus and a small number relayed to the ipsilateral spinal trigeminal nucleus, probably those fibres which arose from the auricular branch of the vagus. The distribution of IXth nerve afferents was broadly similar, with the absence, however, of any projection to the contralateral side. The advent of the Nauta technique, which traces degeneration in unmyelinated fibres as well as myelinated fibres (Glees and Nauta 1955), led to the description of both myelinated and unmyelinated terminal degeneration in the nucleus tractus solitarius (NTS) of many species after section of either vagal or glossopharyngeal rootlets (Torvik 1956; Cottle 1964; Culberson and Kimmel 1972; Cottle and Calaresu 1975).

In the rat, *Torvik* (1956) found terminal degeneration of IXth and Xth nerve afferents throughout the NTS, the majority of degenerating terminals being located caudal to the rostral pole of the hypoglossal nucleus. At this level the heaviest degeneration was found in the lateral division of the NTS, although a large number of finer fibres passed medially to form a plexus in the medial nucleus, especially in its dorsolateral quadrant.

The most authorative study so far is that of *Cottle* (1964) who cut rootlets of both IXth and Xth nerves and traced the resulting degeneration in the medulla of cats. She showed that the rostral third of the NTS received an input from the IXth alone; the caudal-most portion of the NTS receiving an innervation from the Xth. The greatest signs of degeneration were observed in the intermediate zone of the nucleus, which extends from obex level 2.5 mm rostrally, and this zone received degenerating fibres from both IXth and Xth nerves. This region she considered to represent the area receiving the baroreceptor afferent input, since vagal fibres terminating in this area had been shown in an earlier study to originate from vagal rootlets which contained fibres innervating cardiovascular receptors (*von Baumgarten* and *Arranda Coddou* 1959; *Bonvallet* and *Sigg* 1958). Other studies have indicated a marked projection of IXth afferents to this area in different species (mouse, *Aström* 1952; cat, *Kerr* 1962; cat, rat, and guinea pig, *Kimmel* and *Kimmel* 1964; monkey, *Rhoton* et al. 1966; opossum, *Culberson* and *Kimmel* 1972).

With the use of the anterograde transport of horseradish peroxidase, *Katz* and *Karten* (1979) describe the projection of aortic nerve afferent fibres to the subnucleus dorsalis of the NTS in the pigeon. The terminals appear to be restricted to a region which in this species contains catecholaminergic neurones. Using a highly sensitive technique to visualise HRP, *Berger* (1979) described the transganglionic transport of this material from the SN to the NTS in the cat. The ipsilateral medial and lateral portions of the nucleus were densely innervated. A significant contralateral projection to both the commissural and medial subnuclei and to the lateral nucleus at levels rostral to the obex was seen, although this has yet to be confirmed neurophysiologically (see 2.1.2).

The location of vagal and glossopharyngeal terminals is summarised in Fig. 1a, modified from *McAllen* et al. (1979).

### 2.1.2 Electrophysiological Studies

### 2.1.2.1 Antidromic Activation

The anatomical limits for the termination of vagal and glossopharyngeal afferents have been provided by neuroanatomical studies (see Fig. 1a), and within the broad areas thus delineated it would be expected to find the terminals of the baroreceptor afferents originating from both aortic and sinus nerves. In order to identify specifically the projection of these nerves, the technique of antidromic activation of their intracranial projections has been used, recording activity evoked in SN and AN peripherally. This technique has, however, the disadvantages of activating both fibres *en passant* as well as terminals.

In a few studies, an attempt has been made to identify whether evoked responses have resulted from stimulation within terminal fields (*Lipski* et al. 1975; *Jordan* 1977; *Jordan* and *Spyer* 1977a, 1978b). The interpretation of data based on whole nerve evoked activity remains controversial, but the threshold-depth contours obtained in these studies are broadly analogous to those of *Jankowska* and *Roberts* (1972) on single neurones in the spinal cord. More recent studies have confirmed these inferences for single, including aortic, vagal neurones (*Garcia* et al. 1979a, b). It, however, remains a possibility that responses evoked in the sinus nerve on



Fig. 1a-c. Diagrammatic representation of: a projection of IXth and Xth cranial nerves from degeneration studies; b projection of SN and AN afferent fibres revealed by antidromic activation; c location of neurones excited by the carotid sinus baroreceptors and pulse-rhythmically discharging neurones in the NTS. Cross-hatching shows projection of the IXth cranial nerve, sinus nerve afferents and carotid sinus baroreceptors. Stippled area illustrates projection of the Xth cranial nerve, aortic nerve afferents and the location of pulse synchronous neurones. (In this figure, the TS and NTS are projected onto the dorsal surface of the medulla)

stimulating within the medulla can reflect the activation of the efferent nonsympathetic neurones in the sinus nerve. These fibres are thought to be mainly small and predominantly unmyelinated (*Biscoe* and *Sampson* 1968; *Fidone* and *Sato* 1969). *Jordan* and *Spyer* (1977a) drew attention to their inability to activate two efferent nonsympathetic SN neurones on stimulating within the dorsomedial medulla [the NTS, dorsal vagus nucleus (DNV) and the area postrema].

Where this technique has been applied in studying the projections of SN and AN afferents in the cat, antidromic volleys in the SN and AN have always been evoked from the NTS and its near vicinity (*Crill* and *Reis* 1968; *Gabriel* and *Seller* 1970; *de* Groat and Lalley 1974; Lipski et al. 1975; Jordan and Spyer 1977a, 1978b; Donoghue et al. 1977), that is, within the limits provided by the neuroanatomical studies (see McAllen et al. 1979 for review). Crill and Reis (1968), however, have argued for a much wider distribution of the terminals of both SN and AN myelinated afferents. They claimed to have activated them on stimulating within the medial reticular formation in the paramedian reticular nucleus (NPR), an area corresponding to the classical medullary "depressor centre" (Alexander 1946). Lipski et al. (1975) reinvestigated the central projection of the myelinated SN afferents, but were unable to activate these fibres when stimulating anywhere but within the NTS, and even then only from a restricted portion of the NTS, primarily its ventrolateral aspect at levels

rostral to the obex. This restricted localisation of the terminals of largediameter afferents (conduction velocity > 12.5 m/s) has since been confirmed for both the cat and the rabbit (Jordan and Spyer 1977a). It is worthwhile noting at this point that Lipski et al. (1975) were able to reproduce the aberrant results of Crill and Reis (1968) when recording unipolarly from the SN as in Crill and Reis' study. Conventional bipolar recordings never gave equivalent results. For the AN, neither Donoghue et al. (1977) nor Jordan and Spyer (1978b) could evoke antidromic responses except from the close vicinity of the NTS (see below).

Lipski et al. (1975) and Jordan and Spyer (1977) consider that this ventrolateral region of the NTS is the region containing the central afferent terminals of large myelinated SN fibres. This conclusion was reached from studying the relationship between stimulus current and depth (or position of electrode) during a penetration. In this region, the threshold for evoking a response fell abruptly to a low value, this value then being maintained over considerable depth (0.3-0.9 mm) before abruptly increasing again. This contrasts with the effect of stimulating in the region of a discrete nerve tract; at the level where the glossopharyngeal nerve fibres enter the TS, 'point' depth-threshold contours were seen (see McAllen et al. 1979). The presence of terminal arborisations in the NTS conforms to Cajal's description of the distribution of IXth and Xth fibres once they enter and descend through the TS (see Cajal 1909).

Gabriel and Seller (1970) describe a restricted termination of both SN and AN afferents in the dorsomedial portion of the NTS just rostral to the obex in the cat. These must represent the terminals of small myelinated afferents (the latency of these responses being around 10 ms). Jordan and Spyer (1977) have mapped the projections of large myelinated fibres (conduction velocity > 12.5 m/s), small myelinated fibres (conduction velocity 2.5-12.5 m/s), and unmyelinated fibres (conduction velocity < 2.5 m/s) of the SN in both cats and rabbits. The restricted projection and termination of large myelinated fibres have already been described, the small myelinated and unmyelinated afferents being distributed over much wider areas of the NTS, extending beyond its borders especially towards the dorsal vagal nucleus (DNV in the cat, its homologue the n. alaris in the rabbit). In the cat, stimulation at a few points in the area postrema evoked responses in SN afferents of all dimensions. The significance of this projection remains uncertain (for discussion see Jordan and Spyer 1977).

In summary, SN afferents would seem to terminate only in close proximity to the NTS at levels from 0 to 3 mm rostral to obex. Only the largest myelinated afferents (conduction velocity in the range 12.5-32 m/s) have a restricted distribution within the nucleus, smaller myelinated and unmyelinated fibres projecting to much wider areas of the medulla although always in the immediate vicinity of the NTS (see Fig. 1b).

Much less attention has been paid to the projections of the AN, and most information then has come from stimulating the nerve and recording the orthodromically evoked volley in the medulla (see Kumada and Naka*jima* 1972). Crill and Reis (1968), however, provided the first description of the possible localisation of the projection of these afferents using the antidromic technique. Although they claimed a widespread distribution of the afferents within the medulla, complementing their description of the projection of the SN, again their observations do not conform to the anatomical description of the projection of vagal afferent fibres. Although they describe a marked representation of AN afferents, within the NTS, their figures show an abundant distribution in the rostral portions of the NTS (Crill and Reis 1968), which contradicts the restricted nature of degenerating vagal fibres described by *Cottle* (1964) amongst others, within this nucleus. Donoghue et al. (1977), Jordan (1977), and Jordan and Spyer (1978b) have provided evidence that AN fibres projecting to the NTS are found primarily in an area close to the DNV (Fig. 1b). Jordan (1977) and Jordan and Spyer (1978b) have shown that the area of the NTS from which antidromic responses in the AN can be evoked is restricted in both the cat and the rabbit to within 1 mm of the obex. More recent studies by Garcia et al. (1979a, b) have shown that single aortic baroreceptor afferents, whose activity was recorded in the nodose ganglion, project to the vicinity of the NTS. They were shown to send profuse terminal arborisations laterally in the NTS by antidromic activation. In the cat, stimulation within the commissural nucleus of the NTS, both ipsilateral and contralateral to the recording site, evoked responses in myelinated fibres (Crill and Reis 1968; Donoghue et al. 1977; Jordan 1977; Donoghue 1978; Jordan and Spyer 1978b). In the rabbit, aortic nerve afferents appear to project and terminate over a somewhat wider area than in the cat, relatively few terminating in the NTS and most extending from the borders of this nucleus into the nucleus alaris and nucleus intercalatus, but no evidence of a contralateral projection has yet been obtained (Jordan 1977; Jordan and Spyer 1978b).

### 2.2 Baroreceptor Reflex Pathway

From the preceding sections it would seem safe to conclude that the primary relay station in the baroreceptor reflex is located within the confines of the NTS, although its precise localisation and the characteristics of the second-order neurones remain in question. As a consequence of this and the unresolved complexity of connections within the NTS, the efferent connections from the nucleus concerned with cardiovascular control remain ill-defined. Both neuroanatomical and neurophysiological studies, have, however, revealed something of the profusion of probable pathways, and from a combination of these approaches it is not too optimistic to expect the development of new and detailed information on the baroreceptor reflex pathway.

### 2.1.1 Neuroanatomical Studies

The initial neuroanatomical studies involved placing lesions in areas of the NTS and searching the central nervous system for signs of anterograde degeneration (Kosaka and Yagita 1905; Hirose 1916; Morest 1967; Norgren and Leonard 1973; Cottle and Calaresu 1975; Palkovits and Zaborsky 1977) or else placing lesions in presumed target areas of the nervous system before analysing the NTS for signs of chromalysis through retrograde degeneration (Torvik 1957).

Although degeneration studies have been valuable when conducted by skilled investigators, the techniques have sometimes proved capricious, and the interpretation of results is often suspect in the absence of electron microscopy. In recent years there has been a tendency to replace this type of investigation by the use of radioactive-labelled amino acids to reveal efferent connections through anterograde transport (Loewy and Burton 1978; Ricardo and Koh 1978) and by the use of horseradish peroxidase (HRP) which may be transported in both an anterograde and retrograde direction (Kalia 1977; Amendt et al. 1978; Loewy and Burton 1978; Mesulam 1978). Autoradiographic studies have proved particularly useful in determining whether connections between regions were significant or merely represented fibres of passage. Even though these new and apparently reproducible anterograde and retrograde methods represent a distinct advance over previous techniques, the interpretation of the resulting data requires caution, and it is desirable to confirm conclusions using both methods. Moreover, it is important to recognise that in the absence of physiological data it is not possible to determine unequivocally the relationship of specific anatomical pathways to cardiovascular control.

*Morest* (1967), using degeneration techniques, provided the first detailed modern description of the efferent projections of the NTS in the cat, although the study was restricted to the caudal half of the nucleus and then only its medial aspects. He emphasised the problems associated with any encroachment of lesions into the surrounding reticular formation but showed that lesions of the medial NTS at the level of the area postrema resulted in the appearance of degenerating terminals in the lateral nucleus of the NTS, the dorsolateral reticular formation of the medulla, the nucleus ambiguus (NA) and retrofacialis and the lateral reticular nucleus. He traced degeneration also to the n. intercalatus (see also *Cottle* and *Calaresu* 1975, who, however, have been challenged by *Loewy* and *Burton* 1978) and medial reticular formation, but observed no degenerative changes in the spinal cord, or thalamus; indeed they could find no convincing evidence at all for an ascending projection to the diencephalon. *Morest* (1967) also showed that the subnucleus gelatinosa of the NTS sent efferent connections only *within* the NTS. *Cottle* and *Calaresu* (1975) described a broadly similar pattern of degeneration resulting from similar experiments, observing considerable degeneration within the DNV.

The absence of evidence for a solitariospinal projection in the study of *Morest* (1967) is countered by observations of *Torvik* (1957) in the cat. In fact, there is now overwhelming evidence in favour of direct connections from the NTS to the spinal cord, but these appear to emanate mainly from the ventrolateral nucleus (*Torvik* 1957; *Norgren* 1978; *Loewy* and *Burton* 1978). *Torvik* (1957) describes chromalytic changes in NTS neurones after spinal lesions in the rat, and Norgren (1978) has shown anterograde transport of labelled amino acids to the spinal cord from the ventrolateral nucleus. This descending pathway is mainly contralateral (*Norgren* 1978; *Loewy* and *Burton* 1978), but there is evidence of some ipsilateral transport of labelled amino acids (*Loewy* and *Burton* 1978). The pathway appears to be concerned primarily with medullary control of respiratory motoneurones (*Euler* et al. 1973a, b). There is, however, evidence of terminal labelling within both the intermediolateral and intermediomedial columns (*Loewy* and *Burton* 1978).

More detailed information on solitariospinal connections has been provided by the use of HRP. Neurones in the ventrolateral portion of the nucleus have been labelled by HRP injections into the contralateral thoracic and cervical spinal cord (*Loewy* and *Burton* 1978). Interestingly, the intermediate and commissural nuclei have been shown to receive bilateral labelling from injections made at cervical, thoracic, lumbar, and sacral cord levels, although the medial nucleus remains unlabelled (*Loewy* and *Burton* 1978). This latter observation has been questioned by *Seller* and his colleagues, who describe an extensive labelling in the dorsomedial part of the nucleus after HRP injections into the thoracic intermediolateral cell column (*Amendt* et al. 1978). *Loewy* and *Burton* (1978) would, however, argue that this labelling, apparently localised in the caudal part of the dorsomedial NTS, may be confused with labelling in the dorsal column nuclei.

The differences in the patterns of efferent connection of the constituent subnuclei of the NTS that are beginning to the revealed may have profound physiological significance. The ventrolateral nucleus contains the dorsal group of inspiratory neurones (*Baumgarten* and *Kanzow* 1958, and subsequently others) and nonrespiratory neurones there receive an excitatory input from the SN and carotid sinus baroreceptors (*Lipski* et al. 1975), whilst the medial nucleus receives AN and SN inputs (see Sect. 2.1.2) but there is as yet little convincing evidence of a direct sinus baroreceptor input (*Lipski* et al. 1975). The commissural and intermediate nuclei may receive an AN input (*Jordan* and *Spyer* 1978b; *Garcia* et al. 1979a, b).

The rostral projections from the NTS are likely to be particularly important in both cardiovascular and respiratory control. In the case of respiratory control the connections with the pontine reticular nuclei, the nucleus parabrachialis and Kolliker-Fuse nuclei, shown using HRP (Kalia 1977; Loewy and Burton 1978) and by injections of amino acids in the NTS (Loewy and Burton 1978), are particularly significant especially as reciprocal connections may exist (see Kalia 1977). In a particularly extensive study, Loewy and Burton (1978) have shown not only dense projections from the NTS to these pontine nuclei but also projections ascending as far rostrally as the periaqueductal grey, the inferior colliculus and even to the bed nucleus of the stria terminalis. From other studies using HRP it is clear that the NTS projects to the contralateral locus coerulus with a small ipsilateral projection (Cedarbaum and Aghajanian 1975; Sakai et al. 1977); this structure is itself known to project to the diencephalon (Loizou 1969).

Accordingly, these observations both complement and extend the study of Morest (1967) whilst beginning to question the absence of a diencephalic projection (see also *Ricardo* and *Koh* 1978). They also confirm and illustrate the profusion of connections within the medulla: there are projections to the medial reticular formation, including the NPR and to the raphe complex and striking projections to the NA and lateral reticular nucleus (LRN). Loewy and Burton (1978) and Ricardo and Koh (1978) draw special attention to the connections between the NTS and the Al group of noradrenergic neurones in the LRN (Dahlström and Fuxe 1965). In addition they illustrate a pathway from the lateral NTS to the medial accessory olivary nucleus Loewy and Burton 1978). In the context of ascending projections, the autoradiographical and HRP studies of Ricardo and Koh (1978) provide indications in the rat, at least, that the NTS sends efferent projections to the lateral subthalamus, the parabrachial area and to the paraventricular, dorsomedial and arcuate nuclei of the hypothalamus. In addition the medial preoptic area, periventricular nucleus of the thalamus, bed nucleus of the stria terminalis and central nucleus of the amygdala seem to receive an input in this species. Neurophysiological studies are now required to assess the significance of these pathways in the baroreceptor reflex and cardiovascular control.

### 2.2.2 Electrophysiological Studies

The limits for the projection of primary afferent fibres of the SN and AN have been inferred from neuroanatomical studies and confirmed by the
antidromic mapping experiments described in a previous section. The more traditional neurophysiological technique of recording evoked responses from a population of neurones on stimulating a peripheral nerve has also been used to obtain information on the projection of SN and AN afferents, and provides a potential advantage in evoking discharges at several levels within the reflex pathway. In this way it is possible to use the generalised field response as a marker for single unit studies. This is of particular importance in the case of the projection of SN and AN afferents, as in the former case it is possible then to use physiological stimulation of the afferent receptors, baroreceptor and chemoreceptor, so as to identify the role of neurones in specific reflex pathways.

# 2.2.2.1 Evoked Potentials

Lam and Tyler (1952), in the most important early study of this type, were able to record an evoked potential of complex waveform in the ipsilateral NTS on stimulating the cervical vagus nerve of the rabbit, primarily in its commissural and "dorsosensory" areas (using the nomenclature of Cajal 1909), i.e. dorsomedial portion of the nucleus. Anderson and Berry (1956), in a subsequent and more detailed study in the cat, described similar evoked responses in the NTS but showed a more widespread distribution of points within the medulla from which they could record evoked potentials in response to vagal stimulation. Indeed, they drew particular attention to the fact that they could record within the intramedullary course of vagal afferent fibres (see also Lam and Tyler 1952), a factor not always accepted as complicating many later studies. Furthermore, they observed what they considered postsynaptic activity in the spinal nucleus of the Vth cranial nerve, the gracilis nucleus, and more often in the DNV, NA, n. intercalatus and parasolitary nucleus, as well as in the lateral reticular formation. They also noted the antidromic activation at short latency of neurones in the DNV and NA on stimulating the cervical vagus, a point later reiterated by Porter (1963) in a similar study. Clearly, observing evoked potentials on stimulating an afferent input cannot prove a response to have been generated postsynaptically; indeed, any conclusion regarding the complexity of connection between the afferent input and the neuronal site under investigation must be viewed critically if it rests solely on the evidence of evoked potentials.

The Aortic Nerve. The earliest work concerning specifically the effects evoked on stimulating the AN was that of Anderson and Berry (1956) in the cat. They showed that electrical stimulation of the central end of the cut AN evoked activity in both the intracranial and medullary course of vagal fibres and the NTS. The negative field potential in the NTS consisted

of two major waves, which were considered to represent activity generated by the different fibre types found in the AN; Paintal (1972) refers to a bimodal distribution of fibre size in this nerve. Similar negative field potentials have been described by others in the NTS on stimulating the AN (Gahery and Ancri 1967; Biscoe and Sampson 1970a), although often the evoked responses were recorded outside the limits of the nucleus within the nucleus parvocellularis (see Biscoe and Sampson 1970a). Gabriel and Seller (1970) describe a response restricted to the dorsomedial portion of the NTS at a level approximately 0.5 mm rostral to the obex. This response, due in the authors' opinion to activation of only the lowest threshold afferents in the AN, had a latency of 10-11 ms, and this study never revealed responses in the latency range 2-6 ms commonly reported by other workers (Gahery and Ancri 1967; Biscoe and Sampson 1970a; Kumada and Nakajima 1972). The most detailed study so far is that of Kumada and Nakajima (1972) in the rabbit. They recorded evoked responses of widely ranging latency in the NTS and sparsely within the nucleus alaris (or DNV). They assumed that short-latency responses (< 40 ms) reflected activation of large myelinated afferent fibres and that these were likely to be generated monosynaptically. Long-latency responses (> 40 ms), seen in the NTS and elsewhere, were considered a consequence of the activation of small diameter fibres. As responses of short latency were also seen in the NA and lateral reticular formation, this distinction is hardly tenable. Such a conclusion would have a stronger basis if the afferent volley had been monitored as in the study of Lam and Tyler (1952). Even then it is impossible, on the basis of the evidence presented, to deny that both short and long latency responses might be generated over polysynaptic pathways.

In a recent study *Gahery* and *Ancri* (1967) reported an evoked response to AN stimulation in the NTS, in a region medial to TS where they had previously recorded neuronal activity with a cardiac rhythm of sinus baroreceptor origin. The latency of this evoked response fell into two latency ranges (2-3 ms and 4-6 ms).

From the Sinus Nerve. In the late 1960s, the effects of stimulating the SN on the activity of medullary neurones were investigated. Humphrey (1967) reported a maximal evoked response in the intermediate region of the ipsilateral NTS (0.5–2.0 mm rostral to the obex). This response (of negative polarity) had a minimum latency of 3–4 ms; since then similar responses have been observed in the NTS by numerous workers (Miura and Reis 1969a; Biscoe and Sampson 1970a; Spyer and Wolstencroft 1971; McAllen 1973; de Groat and Lalley 1974; Hildebrandt 1974; Lipski and Trzebski 1975). Humphrey (1967) noted that this response was not restricted to the NTS but was also seen in the subadjacent reticular forma-

tion, an observation extended by *Biscoe* and *Sampson* (1970a), who found that the most prominent effect of SN stimulation was an evoked response in the nucleus parvocellularis. *Humphrey* (1967) also noted that at the level of the obex, SN stimulation evoked responses bilaterally in the NTS, the contralateral response having a latency 3-4 ms in excess of the ipsilateral response.

Biscoe and Sampson (1970a) drew attention to positive-going potentials evoked in the medial reticular formation (n. gigantocellularis), which were associated with a decrease in neuronal excitability. The possibility of a major monosynaptic input of SN afferents to neurones of the medial reticular formation, particularly those of the NPR, has been a matter of controversy for a number of years. The question of a primary afferent projection has been approached using the antidromic activation of SN afferents on stimulating within the medulla (see above), and the consensus appears to be that this is unlikely to represent a significant projection. Miura and Reis (1969b), however, claimed the occurrence of a monosynaptic evoked potential within the NPR on stimulating low-threshold SN afferents. The particularly short latency (0.7-1.4 ms), together with the smooth contour and enormous magnitude of the response, seemed surprising in view of the nature of the responses evoked by others in the NTS. Spyer and Wolstencroft (1971) were unable to obtain a similar response on stimulating the SN except in cases where they had sufficient current spread to activate the neighbouring hypoglossal nerve. Only then could they record a similar potential in the region of the NPR, which is ventral to the hypoglossal nucleus. Presumably the "smooth-contoured" response reflects the antidromic potential evoked in the hypoglossal nucleus, viewed at a distance by the microelectrode.

In addition to the "early" response in the NPR, *Miura* and *Reis* (1969a) drew attention to a similar response in the intermediate portion of the NTS. They also observed longer latency response in the NTS and NPR, "late" responses (> 5 ms) being much more widely distributed through the medulla, and long latency responses (6–150 ms latency) were observed in the pons. Responses with a latency of up to 80 ms were observed in the rostral medulla (*Miura* and *Reis* 1969a), and more recently *Adair* and *Manning* (1975) have claimed to evoke activity in the perifornical region of the hypothalamus with a latency in the range from 20 to 120 ms.

Returning to the NTS, Seller and Illert (1969) and Gabriel and Seller (1970) have reported a response restricted to the dorsomedial part of the nucleus, the part where Gabriel and Seller (1970) have also described a restricted input from the AN. They attributed the response to an effect of activating the lowest threshold SN afferents but surprisingly the latency was in the range 7-11 ms, much in excess of the "early" response of Miura and Reis (1969a) or the short latency responses of others (Humphrey

1967; Biscoe and Sampson 1970a; Spyer and Wolstencroft 1971; Lipski et al. 1975). There is no obvious explanation for this discrepancy, nor indeed for the wide variations between response recorded in different laboratories. Presumably the anatomical arrangement of the SN in close proximity to other nerves relaying to the region of the NTS makes effects due to current spread a particular problem, and the wide variety of fibre size in the SN may well militate against consistent observations from one laboratory to another.

## 2.2.2.2 Single Unit Studies

Aortic Nerve Stimulation. The literature contains relatively few reports of the effects of AN stimulation on individual brainstem neurones. Gahery and Ancri (1967), who observed evoked potentials within the NTS on stimulating the AN, did so in areas where they had observed pulse-synchronous activity, which they considered to be of SN origin, as the animals had previously had their AN sectioned. In more recent studies, Donoghue (1978) and Donoghue et al. (1978) report that AN stimulation in cats evokes activity in medullary units with either short (< 20 ms) or long (> 20 ms) latencies. These responses are considered to be generated by activation of myelinated and unmyelinated afferents respectively. Short latency responses were observed in several regions of the medulla in addition to the medial portion of the NTS, the dorsomedial and lateral reticular formation and the NA being areas particularly affected. Those units activated at longer latencies were only observed in the dorsomedial medulla and were concentrated in the medial portion of the NTS. No responsive cells were located in the medial reticular formation. An indication that there was a convergence of input from the myelinated and unmyelinated components of the AN was obtained from observations that some neurones, located in the vicinity of NTS and NA, were activated at both short and long latency. Biscoe and Sampson (1970b) report activating a few single neurones of the NTS on stimulating the AN. Many neurones outside the limits of the NTS in the n. parvocellularis also responded to SN stimulation (as well as showing excitatory responses to other IXth and Xth nerve inputs) and showed excitatory responses to AN stimulation. McAllen 1973) similarly reported a convergence of excitatory inputs from SN and AN onto neurones outside the NTS (around the DVN, in the "parahypoglossal" region, and close to the NA), but within the NTS neurones were excited by either the AN or SN but never both. Schwaber and Schneiderman (1975) also observed synaptic activation at short latency of NTS neurones on stimulating the AN (latency 4–50 ms) in the rabbit. Similar synaptic effects were also noted in this study in the region of the n. alaris (or DNV). Brickman et al. (1977) have also observed effects of AN stimulation on hypothalamic neurones. Stimulating the AN produced excitatory responses in approximately 5% of neurones recorded in the anterior hypothalamus-preoptic region. Considering the conduction distance and the inevitable involvement of a polysynaptic pathway, the latencies of these responses were often remarkably short (14-176 ms). This area would seem equivalent to the area described by *Hilton* and *Spyer* (1971) in the cat as the hypothalamic "depressor" area, whose involvement in the integration of the carotid sinus baroreceptor reflex has been demonstrated (see Sect. 2.2.3.3).

Sinus Nerve Stimulation. The picture from studies on the effects of SN stimulation on single neurones in the brainstem broadly complements the data from recordings of evoked potentials. All authors are agreed that there is a major monosynaptic input to neurones of the NTS (Humphrey 1967; Miura and Reis 1968, 1969a; Biscoe and Sampson 1970b; Spyer and Wolstencroft 1971; McAllen and Spyer 1972; Weiss and Kastella 1972; Miura 1975; Lipski et al. 1975; Lipski and Trzebski 1975; Hildebrandt 1974; Lipski et al. 1976; Miura and Kitamura 1979), although the extent and properties of this input vary widely in the reports from different groups. Whilst the results of Lipski et al. (1975) agree closely with the anatomical data and the results of their antidromic studies, i.e. the major effects of myelinated SN afferents being observed in the ventrolateral portion of the nucleus at levels rostral to the obex, Biscoe and Sampson (1970b) reported only a small proportion of activated neurones in this area, the majority being located outside the limits of the nucleus in the n. parvocellularis. Miura and Reis (1969a) reported that the majority of responses they obtained (unitary activity superimposed on evoked potentials) were in the intermediate part of the NTS, 1 mm ahead and behind the obex.

Humphrey (1967) and Lipski et al. (1975) drew particular attention to the paucity of neurones activated with a latency compatible with a monosynaptic input from large myelinated fibres (i.e. within 8 ms) on stimulation of the SN. Lipski et al. (1975) found that only 17% of neurones in the NTS were activated within 8 ms; Humphrey (1967) and McAllen (1973) consider that it is likely that many neurones are activated over both monosynaptic and polysynaptic pathways, the faster and more direct pathway sometimes requiring considerable temporal and spatial summation to make its effect noticeable (see McAllen 1973; McAllen et al. 1979). Such a complex interneuronal arrangement within the NTS confirms the impression from histological studies of Morest (1967) and Cottle and Calaresu (1975), as well as others discussed earlier.

The latency of responses in the NTS was, however, usually shorter than noted elsewhere in the medulla, although considerable overlap was noted.

The "parahypoglossal region" was shown to receive a major input from the SN over a relatively direct pathway, if latency can provide such evidence (Lipski et al. 1975; McAllen and Spyer 1972), but there is no evidence for a monosynaptic input of SN afferents of any fibre diameter to this region (Jordan and Spyer 1977). Apart from an input to this area, which lies close to the hypoglossal nucleus, Lipski et al. (1975) could find no excitatory (or inhibitory) input to neurones of the medial reticular formation, an area classically termed the "drepressor" area (see Alexander 1946) and containing the NPR, where Miura and Reis (1969a) and Homma et al. (1970) have claimed a significant monosynaptic input. Humphrey (1967) considered the presence of a polysynaptic input to this region and to the midline raphe nuclei as significant, but its influence was never seen by Lipski et al. (1975). In contrast, Biscoe and Sampson (1970a) describe a positive field potential in midline regions associated with a reduction in the excitability of neurones. It is worth noting at this point that Scheibel et al. (1955) were unable to activate neurones in this medial reticular area on stimulating the cervical vagus, and Spyer and Wolstencroft (1971), analysing the response of NPR neurones relaying to cerebellum or activated synaptically by stimulation of the cerebellar peduncles, could find no evidence for a convergent input, either excitatory or inhibitory, from the SN. There is, however, anatomical evidence for connections from the NTS to these areas (see above). Miura and Reis (1972a) present conflicting results arguing for a convergence of fastigiofugal fibres and SN afferents at the level of the NPR. This will be discussed later (Sect. 4.2.2).

There is much greater agreement on the lateralisation of SN effects in the medulla. Most investigators report a significant proportion of neurones in the lateral regions of the medullary reticular formation responsive to SN stimulation (Humphrey 1967; Biscoe and Sampson 1970b; McAllen and Spyer, 1972; McAllen 1973; Davies and Edwards 1973, 1975; Hildebrandt 1974; Lipski et al. 1975). In particular, responsive neurones have been reported in the vicinity of the NA (Davies and Edwards 1973, 1975; Hildebrandt 1974; Lipski et al. 1975; Spyer 1975), although there is some disagreement as to the nature of the input. Lipski et al. (1975) have shown that it is mediated by a polysynaptic pathway, the latency of responses being significantly longer than usually observed within the NTS or "parahypoglossal" region, whilst Davies and Edwards (1975) claim a monosynaptic input, i.e. a latency comparable with that also observed in the NTS. As there is no histological evidence for such a direct connection of either SN or AN afferents and as the antidromic mapping study of Jordan and Spyer (1977) failed to reveal a primary afferent projection of the SN to this area, it would seem best to ignore this conclusion. The significance of an influence of SN inputs on neurones of the NA will be discussed later (Sect. 3.6.2).

## 2.2.3 Baroreceptor Stimulation

The limits for the areas involved in the baroreceptor reflex at medullary levels have been provided by the data summarised above, but as the SN in all species contains both baroreceptor and chemoreceptor afferents, these observations are of limited value in ascribing an integrative role to these areas in baroreceptor responses. Two major approaches have been used in attempts to determine which neurones specifically mediate the baroreceptor reflex. The first arose from the premise that as this afferent input is normally pulse modulated, central neurones should have their activity sculptured by such a phasic input. Accordingly, attempts have been made to locate neurones with activity modulated by the pulse in the medulla. The second approach has involved eliciting a change in the stimulus to the baroreceptors and searching for neurones whose activity fluctuates in relationship to the experimental change in afferent barrage. At its simplest this has involved labelling neurones as "cardiovascular" in function if they either increase or decrease their discharge in response to a rise or fall in arterial blood pressure, usually induced pharmacologically. At its most elaborate it has involved controlled stimulation of the baroreceptor endings of the carotid sinus employing a "blind-sac" preparation, sometimes in association with an initial sampling of unit activity by stimulating the intact SN.

# 2.2.3.1 Pulse-synchronous Activity

Although there have been several attempts to identify pulse-synchronous activity in the brainstem, there are surprisingly few units adequately identified and convincingly free of artifactical modulation (see *Hellner* and *Baumgarten* 1961; *Salmoiraghi* 1962; *Smith* and *Pearce* 1961; *Werz* et al. 1974; *Langhorst* and *Werz* 1974; *Humphrey* 1967; *Stroh-Werz* et al. 1977a, b). However, there remains an isolated claim of a profusion of such neurones in the medulla (*Middleton* et al. 1973). The observation that even at the level of the NTS pulse-synchronous activity is rare in single units (*Salmoiraghi* 1962; *Humphrey* 1967; *Stroh-Werz* et al. 1977a, b) may be taken as an indication that the complexity of connections within this nucleus, referred to above, may well *smooth* what is clearly a powerful pulse-modulated input from the peripheral baroreceptors.

A detailed analysis of the firing pattern of NTS neurones, particularly those in the dorsomedial portion of the nucleus, showed that only 28 neurones of several hundred recorded in 17 dogs and 80 cats had activity clearly related to the cardiac cycle (*Stroh-Werz* et al. 1977a), but even then a single and direct baroreceptor modulation was not convincingly demonstrated. The possibility remained that the cardiac rhythm arose from cardiovascular mechanoreceptors other than arterial baroreceptors, and indeed the major result was a convergence of vagal inputs onto neurones that received an input from extravagal fibres (presumably sinus nerve afferents although this was not directly tested). *Miura* and *Reis* (1972b), however, have recorded from 12 neurones which displayed a pattern of discharge which "often bore a striking similarity to the discharge pattern of baroreceptor afferent fibres", this activity being obliterated on bilateral carotid occlusion. Seven of these were in the dorsomedial portion of the NTS within 0.5 mm of the obex, three lateral to the TS, one in the hypoglossal nerve tract, and one just medial to it (presumably the "parahypoglossal" region, see *Lipski* et al. 1975). By constructing post-R-wave histograms of NTS unit activity, *McCall* et al. (1977) have demonstrated cardiac-related rhythmicity in several neurones. The activity of the majority of these neurones (8 out of 14) was not modulated in a simple manner, exhibiting a polymodal distribution of discharge in relation to the R wave of ECG.

The consensus from all these studies would be that distinct pulse-related activity, when present, is mainly restricted to the dorsomedial portion of the NTS (*Salmoiraghi* 1962; *Smith* and *Pearce* 1961; *Humphrey* 1967; *Miura* and *Reis* 1972b; *Middleton* et al. 1973; *Schwaber* and *Schneiderman* 1975; *Stroh-Werz* et al. 1977a, b; see Fig. 1c). Even so, it becomes clear that in only a few cases can this pattern of activity be attributed unequivocally to activation of the carotid sinus baroreceptors, although *Seller* and *Illert* (1969) recorded mass responses (and some single units) with a cardiac rhythm in this area in cats after vagotomy (see also *Gahery* and *Ancri* 1967). Indeed, many authors draw attention to background cardiac rhythmicity (see *Stroh-Werz* et al. 1977a), although the occurrence of convincing single unit responses is so rare.

The observation of a marked convergence of afferent inputs onto single neurones in the NTS (*Stroh-Werz* et al. 1977b) has led to other studies in which the "afferent spectra" of medullary neurones has been investigated. In these studies, a computer analysis of a unit's activity often reveals a cardiac rhythm when post-R-wave histograms or autocorrelogram functions are derived (see *Langhorst* and *Werz* 1974; *Langhorst* et al. 1975; *Stroh-Werz* et al. 1976; *McCall* et al. 1977), but even where this rhythm is abolished by vagal cooling or SN section or both (*Koepchen* et al. 1967, reviewed by *Langhorst* and *Werz* 1974) it is clear that there is no a priori reason for defining the unit as "cardiovascular", as other rhythms can often be distinguished simultaneously.

### 2.2.3.2 "Cardiovascular" Neurones

The question of the identification of "cardiovascular" neurones in the brainstem has been mooted for a number of years. On the basis that both the afferent input from the cardiovascular mechanoreceptors and the efferent outputs from the central nervous system controlling the cardiovascular system are modulated in phase with the cardiac cycle, the belief has grown that the interneurones in the baroreceptor reflex pathway are likely to have a cardiac rhythm. Indeed, it has been assumed that if a central neurone can be shown to have a rhythm in common with the sympathetic efferent output, there is an a priori connection between that neurone and the sympathetic outflow [i.e. it is part of the sympathetic-activity-generating system; see *Gootman* et al. (1975) for a detailed statement of this hypothesis]. This simplistic model rests on the hypothesis that sympathetic activity is generated solely by a tonically active medullary centre (i.e. the "vasomotor" centre), which is reflexly regulated by the baroreceptor input. The paucity of clear pulse modulation at the level of the NTS immediately produces a call for caution in using a "cardiac" rhythm as the sign of such a correlation. Indeed, the earliest claims for identifying "cardiovascular" units comes from studies in which the relative absence of pulse-rhythmic unit activity was stressed (Salmoiraghi 1962; Pryzybyla and Wang 1967). In the first study, "cardiovascular" units were identified as cells whose activity changed in a reciprocal manner to drug induced changes in arterial blood pressure (Salmoiraghi 1962); in the second study, a "cardiovascular" unit was considered to be one that decreased its activity by at least 30% for a 30 mmHg increase in arterial pressure (Pryzybyla and Wang 1967). This latter definition is a very convenient extrapolation, as these authors considered they were recording within the medullary "pressor" area (dorsolateral reticular formation), which they considered to be the sympathetic tone generating centre (cf. Alexander 1946).

Langhorst and his co-workers (Langhorst and Werz 1974; Langhorst et al. 1975), have made the most detailed analysis of the behaviour of brainstem neurones in terms of their relatonships to afferent inputs and possible efferent connections. They have demonstrated the presence of a cardiac rhythm in the activity of many neurones of the "unspecific" reticular formation (i.e. units located outside the limits of the clearly defined reticular nuclei) in the dog, using post-event time histograms and spectral analysis. They have shown that this forms only one of many afferent inputs that can modify neurone activity: they are truly reticular, receiving somatic inputs, respiratory influences and have patterns of activity relating to the delta-theta oscillations of the EEG. Often the cardiac-related discharge was obliterated by section of the relevant afferent input, either vagus or sinus nerve (see Koepchen et al. 1967). Furthermore, this rhythmic discharge was not fixed rigidly: their activity could fluctuate from a rhythm state to a tonic nonmodulated discharge pattern. The most significant conclusion of this study is that no reticular neurone with a rigidly organised pattern of activity has yet been identified. Reticular neurones have multisensorial inputs, somatic as well as vegetative, and evidence of the common rhythms can be observed in both somatic and autonomic outflows. This precludes any definition of reticular neurones as "cardiovascular".

To compound this, there are also reports of pulse-related discharges in neurones in the absence of any baroreceptor input. These have been described in the medulla (*Porszasz* and *Porszasz-Gibischer* 1968), the midbrain (*Baust* and *Niemczyk* 1968) and the hypothalamus (*Baust* et al. 1963). The frequency of the delta-theta EEG oscillation may often, at least in the cat and dog, be similar to that of the cardiac rhythm, so without a full spectral analysis their separation is often impossible (see *Langhorst* et al. 1975) and these apparent cardiac rhythms may well be insignificant. None the less, if taken at face value, these observations underline the problems in using an apparent rhythm to ascribe function, in particular the fallibility of using a 3 Hz rhythm as an indication of a cardiac rhythm in establishing the central pathway of the baroreceptor reflex.

# 2.2.3.3 Specific Baroreceptor Inputs to the Brainstem and Hypothalamus

The baroreceptor input to the brainstem has been investigated by observing the changes in unit activity evoked by slow changes in blood pressure in units already identified as receiving an input from the sinus nerve (Humphrey 1967) or by observing the effects on similarly identified neurones of bilateral carotid occlusion (Humphrey 1967; Miura and Reis 1972b; Nosaka 1976). Both these physiological tests are not "pure": slowly changing levels of blood pressure will evoke changes in the activity of numerous cardiovascular receptors, whilse bilateral carotid occlusion will lead to withdrawal of the carotid sinus baroreceptor input but a concomitant excitation of the carotid body chemoreceptors. Natural baroreceptor stimulation using a "blind-sac" preparation of the carotid sinus to test the responses of units previously shown to receive an SN input is by far the most specific test (Biscoe and Sampson 1970b; McAllen and Spyer 1972; Lipski et al. 1975; Lipski and Trzebski 1975; Lipski et al. 1976). In other cases, the effects of baroreceptor stimulation alone have been used (Trzebski et al. 1962; Trzebski and Peterson 1964; Spyer 1972; Richter and Seller 1975; Yamashita 1977).

The Input to the Medulla. Trzebski et al. (1962) and Trzebski and Peterson (1964) described experiments in which natural stimulation of the carotid sinus baroreceptors was shown to evoke changes in the activity of medul-

lary neurones. Unfortunately details of the location of responsive neurones and the pattern of their response were not given. Humphrey (1967) described in detail the nature of the SN input to the medulla (Sect. 2.2.2). In particular, of 89 neurones in the NTS which were excited by SN stimulation, none had an obvious cardiac rhythm and all showed irregular spontaneous activity. Of these, 28 were investigated in detail for evidence of a hidden "cardiac" rhythm, but even after constructing post-R-wave histograms, no cardiac-related discharge was apparent (Humphrey 1967). This might suggest either that through the complexity of interconnections within the NTS the cardiac rhythm so apparent in the SN was "smoothed" or that these units received an input solely from chemoreceptor afferents. In 18 of these neurones, however, changes in activity correlating with slow changes in arterial blood pressure were apparent. Unfortunately none was tested for a response to sinus inflation. Biscoe and Sampson (1970b) reported on the responses of neurones in the n. reticularis parvocellularis, excited by both SN and baroreceptor stimulation: here they found the largest profusion of neurones excited on stimulating the IXth and Xth cranial nerves and their peripheral branches (see Biscoe and Sampson 1970a, b). Nine neurones excited by nerve stimulation were also depressed by sinus inflation.

Miura and Reis (1972b) reported that of a small population of neurones in the dorsomedial medulla the majority (9 of 12), located within the NTS at levels rostral to the obex, had their cardiac rhythm abolished by bilateral carotid occlusion. It would seem possible that these neurones were also excited by stretching the carotid artery. In a more detailed study, McAllen and Spyer (1972) and Lipski et al. (1975) described a baroreceptor input to a large number of medullary neurones, identified originally on the basis of their excitatory response to stimulation of the SN. Those with the shortest latency were found in the NTS and are illustrated in Figs. 1c and 2. All responded to sinus inflation before reflex changes in arterial blood pressure, and most responded abruptly with a discharge related to the sudden elevation in intrasinusal pressure, with an adapting discharge during the phase of elevated intrasinusal pressure (Lipski et al. 1975). A few, however, showed only gradual changes in activity.

Excited neurones were found also beyond the NTS in the n. reticularis parvocellularis (i.e. equivalent to the description of *Biscoe* and *Sampson* 1970b), many in the "parahypoglossal" region, and others in the lateral reticular formation, particularly in and close to the NA (see Fig. 2). This distribution has been confirmed in other similar but less extensive studies (*Lipski* and *Trzebski* 1975; *Lipski* et al. 1976), which stress the preponderance of responsive neurones in the ventrolateral portion of the NTS extending into the neighbouring reticular formation (*McAllen* and *Spyer* 1972; *Lipski* et al. 1975; *Lipski* and *Trzebski* 1975). The significance of



Fig. 2. Distribution of 91 identified baroreceptor-sensitive neurones plotted onto four standard sections of the medulla. A nucleus ambiguus; AP area postrema; C cuneate nucleus; DNV dorsal motor nucleus of the vagus; EC external cuneate nucleus; IO inferior olive; NTS nucleus of the tractus solitarius; TS tractus solitarius; XII hypoglossal nucleus. Lipski, McAllen and Spyer (1975)

the baroreceptor projection to the NA has been stressed (*Spyer* 1975, 1979) and will form an important part of a subsequent section of this review (Sect. 3.6.2), as it appears that this nucleus contains the preganglionic vagal neurones that supply the heart (see *McAllen* and *Spyer* 1976, 1977, 1978a, b). In this context it is notable that no evidence has been found of a baroreceptor input to the DNV in the cat (see *Lipski* et al. 1975).

The other major feature of the study of Lipski et al. (1975) was the absence of any major input to the medial reticular formation. Four neurones excited by the SN and receiving an excitatory input from the baroreceptor were found at the lateral edges of what may be loosely termed the medial reticular formation, but although the area and its more medial core were routinely penetrated no other excitatory response was identified (Sect. 2.2.2). This contrasts with results of Humphrev (1967) and Miura and Reis (1969a), although Humphrey (1967) drew particular attention to the requirement of considerable temporal facilitation to alter the activity of neurones in the medial reticular formation, relatively few responding to a single shock to the SN or NTS. Of those neurones excited or inhibited by SN stimulation, however, only 10 out of 30 gave consistent responses to carotid occlusion; even then the response was often the opposite of that predicted on the basis of the response to electrical stimulation of the SN. Only one neurone in the study of Miura and Reis (1972b) showing a sinus baroreceptor input can be said to lie in this area, and even then it was more strictly in the "parahypoglossal" region.

Lipski and Trzebski (1975) studied the properties of 400 medullary units, mostly located close to the NTS, found that some 89 responded to SN stimulation, 61 had a purely excitatory input and 23 purely inhibitory, with a latency of 3-21 ms. Of the irregularly firing neurones (i.e. non-respiratory neurones), 44 were excited by baroreceptor stimulation and only 3 were inhibited. Furthermore a proportion of these irregularly firing neurones projected to cervical levels of the spinal cord, although definite proof was established in only 12, of which 7 projected contralaterally and 5 ipsilaterally. The location of the spinal projecting axon was no better established, the authors noting that the intensity necessary to evoke an antidromic response was usually six times greater than required to produce synaptic activation from the cord.

The result indicates the possible involvement of a bulbospinal inhibitory pathway originating from the NTS through which the baroreceptors control sympathetic activity (Sec. 3.3). As such this represented the first direct demonstration of putative bulbospinal interneurones in the baroreceptor reflex, although the small sample and the rapid conduction velocity of its axons clearly limit its significance. Other studies reported by Lipski et al. (1976) may add a further question: using equivalent procedures, they showed that, of 25 nonrespiratory neurones excited by SN stimulation in this same area of the medulla, 2 responded to natural baroreceptor stimulation alone, 8 responded to both baroreceptor and chemoreceptor stimulation, and the remaining 15 responded to natural chemoreceptor stimulation alone. None of this sample was tested for spinal projection, but it appeared that many were located in the vicinity of the NTS and from their latency to SN stimulation appeared early in the reflex pathway. Such a marked convergence of baroreceptor and chemoreceptor inputs might seem justifiable considering the excitatory effects of both inputs on vagal efferent activity in the anaesthetised animal, but strange in view of their often antagonistic effect on sympathetic efferent activity. Since in general terms the two reflexes can be considered to act antagonistically, this apparently powerful convergence at such an early stage in the reflex pathways must be viewed with caution. The simplest explanation for this apparently anomalous effect of chemoreceptor stimulation is that it exerts a generalised excitatory influence on brainstem neurones, particularly those in the vicinity of the NTS, as part of its general arousal influence.

The demonstration in the work of *Lipski* et al. (1975) and *Lipski* et al. (1976) that SN stimulation affected respiratory neurones reinforces the effects reported by others from similar studies (*Biscoe* and *Sampson* 1970b; *Lipski* et al. 1975, 1976b, 1977) and also that baroreceptor stimulation inhibits inspiratory neurones (*Trzebski* and *Peterson* 1964; *Richter* and *Seller* 1975). Baroreceptor stimulation, however, may prolong the duration of discharge of expiratory neurones (*Trzebski* and *Peterson* 1964;

*Richter* and *Seller* (1975). From intracellular recordings of the activity of the lateral group of respiratory neurones, *Richter* and *Seller* (1975) have shown that whilst inspiratory neurones are powerfully hyperpolarised by baroreceptor stimulation (AN stimulation), no synaptic potentials or changes in membrane potential were noted in expiratory neurones during such stimulation. This indicates that the baroreceptor effects on these expiratory neurones are secondary to their action on inspiratory neurones, i.e. the effect represents disinhibition. Since the baroreceptors also exert an inhibitory control of the dorsal group of inspiratory neurones, the role of bulbospinal respiratory pathways in the control of sympathetic activity must be taken into account (*Lipski* et al. 1977 and Sect. 4.3).

The Effects of Lesions on the Baroreceptor Reflex. In addition to the neurophysiological and neuroanatomical data considered above, the ablation of areas of the central nervous system has provided some information on the possible role of central structures in the organisation of cardiovascular control. However, negative results are not sufficient to allow the conclusion that an area has no involvement in the integration of the baroreceptor reflex, as any modulating influence may well not be readily demonstrated in an anaesthetised preparation.

As might be expected, the most striking results have come from lesions encroaching on the dorsomedial medulla, and specifically the NTS, whether produced electrolytically or by radio-frequency current. The first clear statement on the effects of lesions of the NTS was made by Oberholzer (1960), who described that circumscribed electrolytic lesions of the NTS at levels rostral to the obex, in both cat and rabbit, abolished the reflex response to stimulation of the ipsilateral carotid sinus; these lesions were usually some 1 mm rostral to the obex. Similar results were later obtained in the cat by Humphrey (1967). Oberholzer (1960) also noted that more caudally placed lesions were successful in abolishing the cardiovascular responses to AN stimulation. These results broadly confirm the data cited already regarding the sites of termination of the SN and AN nerves (see Fig. 1). The effectiveness of lesions of the NTS in abolishing the carotid sinus reflex has since been confirmed by numerous authors in the cat and the rat (Hilton and Spyer 1971; Miura and Reis 1972b; Doba and Reis 1973, 1974b; de Jong et al. 1975a, b, 1977; de Jong and Palkovits 1976; Palkovits and Zaborszky 1977; Reis et al. 1977; Zandberg et al. 1977). Burkhart et al. (1977) have shown that transverse cuts through the NTS of the dog passing rostral to the obex do not abolish carotid sinus reflexes but totally abolish the responses to AN stimulation, emphasising that a lesion just rostral to the obex in the cat is sufficient to abolish carotid sinus reflexes (see Gabriel and Seller 1970; Miura and Reis 1972b; Doba and Reis 1973, 1974). It is interesting to note also that the effects of lesions within the NTS are not merely acute but chronic; in fact the survival of rats can be severely jeopardised by NTS lesions (see *Doba* and *Reis* 1973; *Zandberg* et al. 1977), and although long-term survival is more common in the cat, blood pressure remains labile (*Reis* et al. 1977). In contrast *Barman* and *Gebber* (1978a) claim that although lesions in the NTS may abolish the baroreceptor reflex and raise blood pressure, it soon returns to control levels. It is interesting in this context that *Snyder* et al. (1978) claim that selective destruction of catecholamine terminals in the NTS of the rat results in a chronic lability of blood pressure.

The above data is of limited value in ascribing functional characteristics to the different areas of the NTS. Little is ever revealed of the precise area destroyed; it may extend considerably beyond the obvious site of ablation and may often involve the afferent input rather than the neuronal elements of the nucleus. Indeed, lesions elsewhere in the medulla are often claimed to affect the efficacy of baroreceptor reflexes and the development of hypertension and this may well result from inadvertent damage to the NTS. The claim by Ylitalo et al. (1974) that lesions of the area postrema produced hypertension were contested by *Hilton* et al. (1974), who drew attention to the close proximity of the NTS to the area postrema and to the failure of these workers to demonstrate whether baroreceptor reflexes were affected by their destruction of the area postrema. Zandberg et al. (1977) proved the validity of this questioning experimentally; lesions of the area postrema are ineffective in producing hypertension unless they extend to damage the NTS when the baroreceptor reflex is either abolished or severely attenuated.

Much of the work concerned with a possible role of other areas of the brainstem in the baroreceptor reflex pathway may well have suffered from the possibility that extensive lesions in the lower brainstem may have destroyed or damaged sinus nerve and vagal afferents en passant or the tractus solitarius and the surrounding nucleus in part. In this context it is not surprising that Wang and Chai (1962, 1967) and Katz et al. (1967) claim that only destruction of the dorsolateral medulla, the "pressor" area, is sufficient to abolish the cardiovascular response to bilateral carotid occlusion. Their lesions, as illustrated, invariably damaged or destroyed the rostral NTS and probably also its afferent input. In contrast, Manning (1962, 1965a, b) reported that massive lesions in the dorsolateral reticular formation did not abolish the reflex responses to bilateral carotid occlusion, although a subsequent decerebration did. However, Wang and Chai (1962, 1967) and Chai and Wang (1962, 1968) have claimed that decerebration at intercollicular levels in no way impairs baroreceptor function, although as *Peiss* (1965) has pointed out decerebration invariably produces some fall in arterial pressure (review even the classical studies of Oswjannikow 1871), which may approach as much as 20 mmHg, in which case baroreceptor reflexes can hardly be considered to be functioning normally. Carotid occlusion is a poor test of baroreceptor function as it results in a concomitant activation of the carotid body chemoreceptors, but the distinction between results is striking. In view of the preceding discussion it is likely that Manning's lesions spared the NTS, as chemoreceptor afferents also terminate there (*Miura* and *Reis* 1972b; *Lipski* et al. 1975; *Jordan* and *Spyer* 1977a).

Manning (1962, 1965a, b) proposed that midline structures in the medulla as well as suprabulbar areas, in particular the hypothalamus, were essential for the typical sympatho-excitatory response to carotid occlusion. In view of the definition of the medial reticular formation as the "depressor" centre (Alexander 1946; Bach 1952), this would seem at first sight an attractive possibility. Unfortunately, electrolytic destruction of the medial reticular formation, including the NPR and raphe nuclei, does not abolish baroreceptor reflexes (Löfving 1961; Hilton and Spyer 1971; Miura and Reis 1972a, b; Barman and Gebber 1978a), although some variable effects have been reported by others. Lesions in the ventromedial medulla have been cited as abolishing the depressor (Smith et al. 1966, quoted by Humphrey 1967) or cardiac (Lee et al. 1972) components of the baroreceptor reflex. Miura and Reis (1972b) claim that the cardiac effects are uneffected, but that whilst the depressor response to SN stimulation is abolished by such lesions, the effects of natural baroreceptor stimulation remain uneffected. Barman and Gebber (1978a), however, leave little doubt that medial areas of the medulla are involved in only non-baroreceptor sympatho-inhibitory responses. Large medial lesions evoked an increase in renal sympathetic activity but left the cardiac rhythm in the renal nerves (postganglionic sympathetic) unaffected, and the sympatho-inhibitory response to baroreceptor activation was unaltered (Barman and Gebber 1978). It appears that lesions in the "parahypoglossal" region, which has been shown to receive both SN and baroreceptor input (McAllen and Spver 1972; Lipski et al. 1975), also do not ablolish baroreceptor reflexes (Doba and Reis 1973).

The effects of laterally placed lesions are, however, much clearer. Lee et al. (1972) state that lesions in the NA of the cat abolish the bradycardia which normally follows the intravenous injection of either adrenaline or veratridine, whilst a lesion in the DNV was ineffective. This result, which has been confirmed for other forms of vagal bradycardia (Kerr 1969; Borison and Domjan 1970), argues for the fact that, at least in the cat, the vagal preganglionic supply to the heart arises from neurones in the NA (see Sect. 3.6.1). This evident lateralisation of the reflex may well extend also to its sympatho-inhibitory component. Evidence cited earlier has argued for powerful efferent projections extending from the NTS to the lateral reticular nucleus (Sect. 2.2.1), and particularly to the A1 group of catecholamine-containing neurones (*Dahlström* and *Fuxe* 1965), which *Coote* and *Macleod* (1974a, b) consider to exert sympatho-inhibitory effects. *Hildebrandt* (1974) describes neurones in the lateral reticular formation which are excited by AN and SN stimulation whose axons can be excited by spinal cord stimulation. This will be discussed in greater detail subsequently (Sect. 3.3).

That baroreceptor function is not totally abolished by decerebration, and may even appear relatively normal, cannot be taken to imply that suprabulbar areas are not essential for a normal expression of the reflex. In fact quite the converse would appear to be the case, for evidence has accumulated that suprabulbar regions may well play an important role in the normal reflex [see Reis and Cuenod (1965) for confusing data concerning the effects of various transections of the neuraxis]. Indeed, Hilton and Spyer (1971) and Spyer (1969) have provided strong evidence for a role of the hypothalamus in the baroreceptor reflex. They defined an area in the anterior hypothalamus of the cat which on electrical stimulation evoked a "depressor" response qualitatively identical to the baroreceptor reflex (Spyer 1969; Hilton and Spyer 1971). Furthermore, Spyer (1969, 1972) has shown that neurones within this area of the hypothalamus are affected by carotid sinus baroreceptor stimulation (see Sect. 2.2.3.3). Bilaterally placed lesions restricted to this region of the hypothalamus reduced the efficacy of the baroreceptor reflex, but never abolished it (Hilton and Spyer 1971). Equally, lesions within the medullary "depressor" area never abolished the reflex response to stimulation of the ipsilateral carotid sinus; they usually reduced it, unless the lesion encroached upon the ipsilateral NTS. Where a significant response remained after a unilateral medullary lesion, bilateral destruction of the hypothalamic depressor area abolished the response completely. Arterial blood pressure did not remain elevated for a considerable time after these hypothalamic lesions, nor was the order of lesions important (Hilton and Spyer 1971). In the same year the importance of suprabulbar areas in the performance of the reflex was reiterated by Kent et al. (1971), who showed that, following decerebration. the range over which the baroreceptors can evoke a change in arterial pressure is severely restricted compared to that in an animal with intact neuraxis.

In summary, the results of lesion studies show that baroreceptors terminate in the NTS and indicate a role for suprabulbar areas in the organisation of the baroreceptor reflex (*Hilton* and *Spyer* 1971). Such a longitudinal arrangement would mirror that for sympatho-excitatory control (see *Hilton* 1975 for review) and would then conveniently encompass other data that argue strongly for hypothalamic interactions in cardiovascular control (*Gellhorn* 1957).

The Baroreceptor Input to Suprabulbar Areas. The importance of suprabulbar areas in the performance of the baroreceptor reflex has been summarised above and data illustrating the ascending projections of the NTS described. It now remains to establish the nature and extent of the baroreceptor input to these rostral structures. Recent studies have shown a baroreceptor influence on hypothalamic unit activity (Thomas and Calaresu 1972; Yamashita 1977; Kannan and Yagi 1978). In particular Spyer (1969, 1972) has described that approximately 5% of neurones investigated in the anterior hypothalamic depressor area (Hilton and Spyer 1971) received a baroreceptor input. Of the 21 neuonres studied, 15 were excited and 6 inhibited by raising the pressure of 200 mmHg in a vascularly isolated carotid sinus. In other experiments it was shown that neurones in this same area were excited on stimulating within the NTS at a latency of 20–50 ms (Spyer, unpublished work). In the rabbit, neurones in the equivalent area have been shown to be excited on AN stimulation, the latency often being surprisingly short (Brickman et al. 1977). In another study, neurones in the perifornical region of the medial hypothalamus were shown to be excited by SN stimulation (Thomas and Calaresu 1972). Of the 23 neurones affected, 7 were excited (latency 17-40 ms, mean 29 ± 3.7 ms), 16 inhibited (latency 30-140 ms, mean  $68 \pm 9.0$  ms). Five inhibited by such stimulation were also inhibited during the pressor response to intravenous noradrenaline. These neurones would appear to be located in the region of the hypothalamus believed to integrate the defence reaction, including its cardiovascular components (see Abrahams et al. 1960). It is interesting that the pattern of response of neurones in this area was the converse of that seen in the hypothalamic "depressor" area (compare Thomas and Calaresu 1972 to Spyer 1972). An antagonistic arrangement of these two hypothalamic areas has been suggested (Hilton and Spyer 1971; cf. Gellhorn 1957), a relationship that may well continue throughout the length of the brainstem. These observations could be interpreted as lending indirect support to such a model.

In addition it has been known for many years that the baroreceptor (and chemoreceptor) inputs alter the release of antidiuretic hormone from the posterior pituitary. It was hence no surprise that *Barker* et al. (1971) were able to demonstrate an effect of SN stimulation on neurosecretory neurones in the supra-optic nucleus. The major effect was excitatory, but more recently *Yamashita* (1977) has shown that neurosecretory neurones in the supra-optic nucleus are excited by chemoreceptor stimulation and inhibited by inflation of the carotid sinus. Similarly *Kannan* and *Yagi* (1978) report an inhibitory effect of the carotid baroreceptors on these neurones.

In addition to baroreceptor inputs to the hypothalamus, there are indications of an AN input to the subthalamus in the rabbit. *Kaufman* et al. (1979) have excited neurones in the zona incerta of the rabbit on AN stimulation with latencies as short as 4 ms. Whilst such latencies are dramatically shorter than previously reported for supramedullary effects and even shorter than usually observed in the medulla, they are consistent with the autoradiographic findings of *Ricardo* and *Koh* (1978) in the rat, which describe a direct projection from the NTS to the lateral subthalamic area.

In addition to this direct demonstration of a baroreceptor influence at the level of the hypothalamus it is well established that this input can have marked effects on behaviour: it can suppress the outbursts of sham rage in hypothalamic animals (*Bartorelli* et al. 1960), influence the sleep-wakefulness cycle (*Guazzi* and *Zanchetti* 1965; *Baust* and *Heinemann* 1967) and influence electrocortical activity (*Bonvallet* et al. 1953). Baroreceptor stimulation is also known to affect motor tone and activity (see *Koch* 1932; *Schweitzer* and *Wright* 1937; *Schulte* et al. 1959; *Dell* and *Bonvallet* 1966), and in a recent study *Coleridge* et al. (1976) have demonstrated that pyramidal tract neurones in the motor cortex of cats are either directly inhibited or prevailing excitatory inputs onto them are suppressed by baroreceptor stimulation. The pathway along which this effect is produced is unknown, although it is likely to involve ascending reticular pathways, which may be influenced from the NTS (*Magnes* et al. 1961; *Akimoto* and *Saito* 1966; *Klee* 1966).

# 3 Baroreceptor Control of Preganglionic Autonomic Activity

## 3.1 Sympathetic Efferent Activity

The central nervous control of the cardiovascular system is exerted primarily by means of the postganglionic sympathetic innervation of vascular beds and the heart. Together with the vagal control of the heart, these efferent pathways constitute the final efferent arm of the various cardiovascular reflexes that are considered to exert a homeostatic control on the circulation (see *Heymans* and *Neil* 1958). Hence it is not surprising that sympathetic efferent activity, particularly that of postganglionic nerves, is distinguished by its marked cardiac or pulse rhythmic discharge (*Adrian* et al. 1932; *Bronk* et al. 1940; *Alexander* 1945, 1946; *Hagbarth* and *Vallbo* 1968; *Polosa* 1968; *Cohen* and *Gootman* 1969, 1970; *Gootman* and *Cohen* 1970, 1973; *Koizumi* et al. 1971; *Mannard* and *Polosa* 1973; *Seller* 1973; *Taylor* and *Gebber* 1975; *Gross* and *Jänig* 1976; *Horeyseck* et al. 1976; *Gregor* et al. 1977). This cardiac rhythm is usually superimposed on a respiratory related pattern of activity (*Adrian* et al. 1932; *Hagbarth* and *Vallbo* 1968; *Polosa* 1968; *Cohen* and *Gootman* 1970; *Gootman* and *Vallbo* 1968; *Polosa* 1968; *Cohen* and *Gootman* 1970; *Gootman* and 1970, 1973; Richter et al. 1970; Koizumi et al. 1971; Seller and Richter 1971; Mannard and Polosa 1973; Seller 1973; Gregor et al. 1977; Lipski et al. 1977; Preiss and Polosa 1977) and is considered to reflect the powerful control of sympathetic activity exerted by the baroreceptor inputs. Indeed this rhythm is usually attenuated, if not totally abolished, by baroreceptor denervation (Gootman and Cohen 1970 and others). There is, however, a claim that this apparent cardiac rhythm of postganglionic sympathetic (and of course preganglionic) activity results merely from an entrainment of the baroreceptor afferent input to an intrinsic rhythm of brainstem origin (Taylor and Gebber 1975; Gebber 1976; Gebber and Barman 1977).

It is well established that preganglionic and postganglionic discharge is characterised by a 3 Hz oscillation in the cat (Green and Heffron 1968; Cohen and Gootman 1970; Gootman and Cohen 1970, 1973; Snyder and Gebber 1973; Taylor and Gebber 1975; Camerer et al. 1977; Gebber and Barman 1977) which has usually been considered to represent the baroreceptor-related inhibitory control of sympathetic discharge. More recently a power spectral analysis of postganglionic sympathetic activity has shown that it contains two peaks at around 3 Hz in the cat (Camerer et al. 1977). One is related to the baroreceptor input and is abolished by baroreceptor denervation, the other is related to the delta-theta oscillation of the EEG and is abolished by decerebration (Camerer et al. 1977). Gebber and Barman (in press), however, have indications that a 2-6 c/s rhythm persists after both baroreceptor denervation and decerebration. Whether this implies that the baroreceptor input is entrained to this "central oscillator" or whether they exert their influences along independent pathways remains to be resolved.

Nevertheless, the cardiac rhythm of the sympathetic discharge survives decerebration (Alexander 1946; Mannard and Polosa 1973; Camerer et al. 1977). This cannot be taken to imply that the normal reflex control of sympathetic activity involves only a bulbar pathway. Furthermore, it is probable that not all sympathetic efferent fibres are affected in a similar manner by baroreceptor inputs. Detailed studies of the sympathetic supply to the hindlimb of the cat, investigating the inputs to hairless and hairy skin and to muscle, suggest that functional distinctions can be made on the basis of the pattern of discharge of individual neurones (Grösse and Jänig 1976; Horeyseck et al. 1976; Gregor et al. 1977; Jänig and Kümmel 1977). Indeed, it would seem convincingly demonstrated at this level of the sympathetic pathway that neurones with a vasoconstrictor function can be distinguished by their powerful cardiac rhythm, muscle vasoconstrictors having the most distinctive cardiac rhythm and inspiratory related discharge (Gregor et al. 1977). These neurones are usually spontaneously active, firing at a low rate, and their pattern of response to various physiological stimuli, as well as stimulation of the neuraxis, provides at least a teleological suggestion of their probable function.

This analogy may also apply at preganglionic levels, where studies on individual fibres or recordings from the cell bodies in the spinal cord suggest that not all preganglionic neurones, when active, have either cardiac or respiratory related discharges (Fernandez de Molina et al. 1965; Polosa 1967, 1968; Koizumi et al. 1971; Mannard and Polosa 1973; Seller 1973; Coote and Westbury 1974, 1979a, b; Preiss et al. 1975; Gebber and McCall 1976). This immediately raises the question of identity: can preganglionic neurones be labelled as vasoconstrictor or cardio-accelerator, or is functional specialisation the reserve of postganglionic elements? As yet, a conclusive answer cannot be given; preganglionic neuronal discharges cannot yet be analysed in as much detail as postganglionic fibres, but a degree of heterogeneity has already been demonstrated by considering just the presence or otherwise of two rhythms, cardiac and respiratory (Fernandez de Molina et al. 1965; Polosa 1967, 1968; Koizumi et al. 1971; Mannard and Polosa 1973; Seller 1973; Coote and Westbury 1974, 1979a, b; Preiss et al. 1975; Gebber and McCall 1976).

This question of identity is not esoteric. Its importance resides in its necessity when establishing the central pathways of cardiovascular as distinct from autonomic control. It has long been considered that the sympathetic nervous system acts in a general and undifferentiated manner (cf. *Cannon* 1929, 1930, 1932), but it is becoming apparent that this is not the case. This original contention arose from two main factors: the exaggerated emotional states investigated and the fact that in most studies whole nerve activity has been recorded rather than the properties of the individual functional groups. With these factors in mind it will be necessary to review the organisation of the preganglionic sympathetic neurones, as the final common pathway within the central nervous system, to determine the synaptic complexity that is responsible for their basic firing properties and, it is hoped, to discern the pathways that mediate the powerful baroreceptor control of their activity.

### 3.2 Baroreceptor Control of Sympathetic Activity

The initial studies relating to baroreceptor control of sympathetic activity have involved the effects of such stimulation on on-going activity (Gootman and Cohen 1970; Koizumi et al. 1971; Foreman and Wurster 1973; Seller 1973; Snyder and Gebber 1973; Taylor and Gebber 1973; Coote and Westbury 1973, 1979a, b; Gebber 1976; Gebber and McCall 1976; Gebber and Barman 1977; McCall et al. 1977; Barman and Gebber 1978a; Geis et al. 1978) and somatic or visceral reflexes into these nerves (Coote and Downman 1966, 1969; Coote et al. 1969; Koizumi et al. 1971; Seller 1973; Coote and Macleod 1974b, 1975). In the context of the present review it is irrelevant to discuss in detail the nature and synaptic complexity of somatic and visceral reflexes into sympathetic nerves. These have been dealt with at length in the past (Sato and Schmidt 1973; Coote 1978; Coote and Sato 1978). The influence of the baroreceptor input on the supraspinal and spinal components of these reflexes will be raised, as this may provide considerable data on the basic circuitry of baroreceptor mechanisms.

It has been known for many years that the baroreceptors exert an inhibitory control of sympathetic efferent activity. Their cardiac rhythm is of baroreceptor origin, but the central pathways involved remained unresolved. The *central delay* of a reflex provides some indication of its synaptic complexity, although affected by the length of the interneuronal pathways, and as such the measurements of the central delay of the baroreceptor-sympathetic reflex are of value. Kezdi and Geller (1968) provided the first detailed open-loop study of the baroreceptor-sympathetic reflex in the dog. They showed that the latency to inhibition of postganglionic sympathetic activity, recorded in either renal or splanchnic nerves, varied from 150-360 ms. From preganglionic recordings in the cat during electrical stimulation of the sinus nerves, Richter et al. (1970) and Seller and Richter (1971) obtained a value of  $181 \pm 23$  ms for complete inhibition of on-going sympathetic activity. Green and Heffron (1968), using natural baroreceptor stimulation, obtained values in the range of 150-300 ms, as did Coote and Downman (1969). These observations, however, leave unanswered the site of inhibition, which could be within the brainstem, or at the level of the cord, or a mixture of both.

The fact that somatic and visceral reflexes into sympathetic nerves have both a spinal and supraspinal pathway (for review see Sato and Schmidt 1973; Coote 1978) at least suggested the possibility that investigating the baroreceptor control of their effectiveness might shed light on the site of interaction between the baroreceptor input and sympathetic control. In fact, the controversy remains, partly no doubt owing to the variability of the early spinal reflex into sympathetic nerves, at least as far as those into the white rami are concerned, and partly to its questioned significance regarding cardiovascular control (see Seller 1973). This questioning has a particular relevance in this context since early or spinal reflexes are rarely seen in cardiac and renal sympathetic nerves (Coote and Downman 1966; Coote and Sato 1978), although often elicited in white rami (see above) and the cervical sympathetic nerves (Coote et al. 1969; Schmidt and Schönfuss 1970). It would appear that at the level of postganglionic nerves, the early response in the anaesthetised animals with intact neuraxis is variable (see Coote and Downman 1969) but is inhibited by baroreceptor stimulation along with the late supraspinal reflex. That an early response

can be recorded in the white rami and postganglionic nerves, yet the white rami responses remain unaffected by baroreceptor stimulation (Coote et al. 1969; Koizumi et al. 1971; Seller 1973) seems at first sight confusing since postganglionic effects have to be transmitted through these rami. Koizumi et al. (1971) found it difficult to record significant early (i.e. spinal) reflexes into renal or cardiac nerves; they were usually small and powerfully affected by the on-going cardiac and respiratory-related discharges of these nerves. These authors do, however, illustrate that when white rami early reflexes were present they were unaffected by baroreceptor stimulation. Coote and Macleod (1974b), however, illustrate that baroreceptor inputs powerfully inhibit the late (i.e. supraspinal) reflex in renal and cardiac sympathetic nerves, but the effects on the early spinal reflex were not entirely clear. The early reflex into cardiac nerves appeared unaffected. They were, however, able to show a baroreceptor inhibition of thoracic white rami early reflexes which contrasts to previous reports (Coote et al. 1969; Koizumi et al. 1971; Seller 1973). This observation has been used by *Coote* and *Macleod* (1974b, 1975) as a significant factor in their claim that the baroreceptor reflex acts, at least in part, by an inhibitory action at the level of the spinal cord (see Sect. 3.3.3.3).

In this context, it is pertinent to draw attention to a study of individual preganglionic fibres recorded in the cat; only 2 from thoracic white rami (out of 55 tested) and 24 from lumbar white rami (out of 54) showed segmental early spinal reflexes to somatic stimulation (*Seller* 1973). Furthermore, only one of these fibres showed a cardiac rhythm in its discharge, and none was silenced by the baroreceptor activation in response to an adrenaline injection.

In conclusion, the central delay of the baroreceptor-sympathetic reflex is long, generally above 150 ms, and there is no controversy regarding the ability of this input to abolish the late (supraspinal) component of somatic and visceral reflexes into sympathetic nerves. The significance for cardiovascular control of those pre- and postganglionic sympathetic neurones receiving an early (spinal) reflex remains in question although in line with the fact that *all* on-going sympathetic activity appears susceptible to powerful baroreceptor stimulation, and if these inputs are in any way concerned with the background discharge of sympathetic neurones some effects on early reflex responses appear possible.

3.3 Preganglionic Sympathetic Neurones

Preganglionic sympathetic neurones are located in the intermediolateral horn of the thoracic spinal cord, with a varying and small representation in the lower cervical and a marked representation in upper lumbar segments of the spinal cord. Some preganglionic neurones may extend from this area into the adjacent white matter of the cord and into the intercalated area (see *Petras* and *Cummings* 1972; *Chung* et al. 1975, and detailed analysis in *Wurster* 1977). The preganglionic supply to the heart originates from T1-5 (*Wurster* 1977) with the greatest density of neurones in the intermediolateral column of the cat being in segments T1-2 and L3-4 (*Henry* and *Calaresu* 1972). Their axons are almost all myelinated and leave the spinal cord via the thoracic and lumbar ventral roots, and after passing for a short distance with the spinal nerves they separate to form the white rami. From here they pass to the sympathetic ganglia, from which unmyelinated postganglionic fibres leave to innervate end-organs. A more detailed discussion of this anatomical arrangement with particular reference to cardiac control is given in a recent review (*Wurster* 1977), and a general description is contained in a survey of sympathetic reflexes (*Coote* 1978).

In the present discussion, interest will be directed specifically at the physiological properties of the preganglionic elements but it will be necessary also to refer to the anatomical arrangements considered to represent the spinal interneuronal circuitry controlling their activity.

#### 3.3.1 Firing Patterns

The activity of individual preganglionic neurones has been recorded by teasing single active units not only from preganglionic nerves, usually the cervical sympathetic (*Pitts* et al. 1941; *Pitts* and *Bronk* 1942; *Preiss* et al. 1975; *Preiss* and *Polosa* 1977) but also from thoracic and lumbar white rami (*Seller* 1973). In these cases, selection was dependent on the presence of spontaneous activity, which is not an ubiquitous property of sympathetic neurones. The alternative approach is to record from the cell bodies of these neurones, identifying the neurone from its antidromic response to stimulation of its axon in the white rami or other preganglionic nerves (*Hongo* and *Ryall* 1966; *Polosa* 1967, 1968; *Mannard* and *Polosa* 1973; *Taylor* and *Gebber* 1973; *Wyszogrodski* and *Polosa* 1977; *McCall* et al. 1977). In addition, intracellular recordings are now possible (*Fernandez de Molina* et al. 1965; *Coote* and *Westbury* 1974, 1979b).

Recordings from preganglionic fibres (Adrian et al. 1932; Pitts et al. 1941; Alexander 1946; Preiss et al. 1975; Preiss and Polosa 1977) show that these neurones fire at a low rate, often without an obvious cardiac rhythm, and it would appear that the frequency-limiting step in the sympathetic outflow is not a property of the ganglion alone but is already imposed by or before the preganglionic neurones (Pitts and Bronk 1942).

The "silent period" following a burst of activity first observed by Pitts and Bronk (1941) was defined as post-excitatory depression, i.e. an orthodromic volley producing a period of subexcitability. These authors considered this to reflect the properties of the excitatory pathways impinging on them. Conversely, Polosa (1967, 1968), recording extracellularly from preganglionic cell bodies in the spinal cord, suggests that the "silent period" following intense activity is a property of the neurone itself, as it may be produced by antidromically evoked spikes. He purported to eliminate the necessity for a recurrent inhibitory interneuronal pathway (Renshaw-type cell), because he provides evidence that the effect was only produced by an antidromic spike, a subthreshold stimulus to the cervical sympathetic failing to elicit a "silent period". Furthermore, in 8 out of 10 cases suprathreshold stimuli failed to affect the duration of the "silent period". Indeed in his later study Polosa (1968) showed that an antidromic spike would reset the on-going rhythm of a preganglionic neurone without significantly shortening the expected interspike interval. In certain cases, however, Polosa (1967) found indications of Renshaw-type recurrent effects on cervical sympathetic stimulation (i.e. 4 neurones), but strychnine did not block the apparent inhibition. More recently, Mannard and Polosa (1973) have extended this study and have retained the conclusion that the firing rate is broadly an intrinsic property and not determined solely by presynaptic events. Two recent studies, have, however, revived the possibility that the activity of a proportion of preganglionic sympathetic neurones may be controlled by a recurrent collateral pathway (Barman and Gebber 1978b; Gebber et al. 1978; Lebedev et al. 1980), although the appropriate interneurone has yet to be described (Rethelyi 1972). Gebber and Barman (1979) consider that the inhibitory interaction between preganglionic sympathetic neurones results from a cholinergic action of sympathetic neurones with C-fibre axons on the more numerous sympathetic B-fibre neurones. It is as yet unresolved whether it involves a recurrent pathway or results from dendrodendritic interactions.

Whatever mechanism(s) is responsible, there is no question that in the intact anaesthetised animal, preganglionic neurones when active fire at rates below 10 impulses/s (*Fernandez de Molina* et al. 1965; *Polosa* 1967, 1968; *Mannard* and *Polosa* 1973; *Snyder* and *Gebber* 1973; *Taylor* and *Gebber* 1973; *Coote* and *Westbury* 1974, 1979a, b; *Gebber* and *McCall* 1976, and see above for fibre recordings). Spontaneous activity appears in as few as 20% of antidromically identified neurones (*Polosa* 1968; *Mannard* and *Polosa* 1973) to as many as 60% (*Coote* and *Westbury* 1974, 1979a), most studies suggesting around 30%. Interestingly, *Mannard* and *Polosa* (1973) have shown that although the incidence of spontaneous discharge is reduced as one goes from the intact, to the spinal, and finally

to the segmental preparation, background firing remains in even this final situation (see also *Alexander* 1945).

The discharge of preganglionic neurones, recorded either from fibre or cell body usually appears irregular, but by constructing post-R-wave histograms (*Seller* 1973; *Taylor* and *Gebber* 1973; *Gebber* and *McCall* 1976; *McCall* et al. 1976, and see Fig. 3) or autocorrelogram functions (*Mannard* 



Fig. 3. Phase relationship between averaged arterial pulse wave and post-R wave TIH of antidromically identified preganglionic neurone. Sample run 256 c/s, address bin 4 ms, blood pressure 160/125 mmHg. Adapted from *Gebber* and *McCall* (1976)

and Polosa 1973) a clear cardiac rhythm can be shown. Coote and Westbury (1974, 1979) have constructed pulse-triggered histograms of sympathetic neuronal activity and have shown that the probability of discharge was minimal 150–200 ms after the beginning of systole. Moreover in a group of neurones the effects of inflating a blind sac of one carotid sinus was tested; the mean latency to inhibition was  $148 \pm 14$  ms (SE, n = 15). This value for units recorded at T3 level is in close agreement with previous studies on the reflex time (see above).

At this point it is worth noting that both *Fernandez de Molina* et al. (1965) and *Coote* and *Westbury* (1979) report intracellular recordings from antidromically identified sympathetic neurones. As yet, however, no clear data have resulted on the nature of the baroreceptor control of their activity, as pulse-related changes in membrane potential have yet to be established.

#### 3.3.2 Spinal Interneuronal Control of Preganglionic Neurones

The possibility that preganglionic neurones are under recurrent control implies the involvement of a local inhibitory interneurone (Barman and Gebber 1978b; Gebber et al. 1978; Gebber and Barman 1979; Lebedev et al. 1980). There is no anatomical support for this contention (Rethelyi 1972), but Gebber and McCall (1976) describe non-antidromically driven neurones in close proximity to antidromically driven preganglionic neurones (stimulating the cervical sympathetic nerve and recording at T1-5). These neurones had a cardiac rhythm (post-R-wave peak  $159 \pm 17$  ms in 15 out of 43 studied) as against a post-R-peak of  $177 \pm 10$  ms for preganglionic neurones (i.e. antidromically identified neurones). These 15 nonantidromically driven neurones were also inhibited by baroreceptor stimulation, as were preganglionic neurones, but in general whilst a preganglionic neurone seldom fired more than once in each cardiac cycle, "interneurones" fired in bursts of up to 4 spikes/cardiac cycle. Further, brainstem stimulation at "pressor" sites evoked excitatory responses in "interneurones" as well as preganglionic neurones, but the onset latency was shorter to the former (mean  $31 \pm 3$  ms compared to  $41 \pm 6$  ms for preganglionic neurones). There was no significant difference, however, in either the latency or duration of the effects of "depressor" area stimulation on their discharge. It was concluded that they represent an excitatory interneurone, presumably mediating descending excitatory (and possibly segmental reflex control) of preganglionic neurones.

In more recent studies, *McCall* et al. (1977) have described neurones in the intermediomedial region of the spinal grey with properties suggesting that they might represent inhibitory interneurones controlling preganglionic activity. It is known that neurones in this area project to the intermediolateral horn (*Bok* 1928; *Petras* and *Cummings* 1972). Twenty-nine neurones in this area were spontaneously active but silenced on bilateral carotid occlusion and had cardiac rhythm. They also received a shortlatency excitatory input from the NTS (latency  $8 \pm 1$  ms; conduction velocity  $15 \pm 2$  m/s). These properties provide indirect evidence for their role as inhibitory interneurones but direct evidence is still lacking.

In the context of delineating the spinal organisation of the control of preganglionic activity, this tentative description of two groups of interneurone may be important in discerning the nature and involvement of descending excitatory and inhibitory pathways in mediating the baroreceptor reflex.

#### 3.3.3 Supraspinal Influences on Sympathetic Activity

It is now well established that electrical stimulation at numerous sites within the brainstem can evoke increases in arterial blood pressure, primarily through an increase in sympathetic activity. There are, however, examples of brainstem stimulation evoking changes in sympathetic activity without concomitant alterations in arterial blood pressure (*Kahn* and *Mills* 1967).

The first direct demonstrations of brainstem influences on sympathetic discharges were those studies which showed an influence of the hypothalamus on sympathetic activity made in the 1930s and 1940s (*Bronk* et al. 1940; *Pitts* et al. 1941; *Pitts* and *Bronk* 1942). Furthermore, these studies revealed a marked interaction between these excitatory effects and baroreceptor inputs. From these and subsequent investigations, it became accepted that the activity of sympathetic preganglionic neurones was determined by the algebraic summation of excitatory and baroreceptor inputs at the level of the "vasomotor" centre. Sympathetic neurones have a low dynamic firing range, partly determined by intrinsic properties, but in general considered to result from interactions at the level of the "vasomotor centre", i.e. from its excitatory input. As such, baroreceptor control of these neurones can be considered to result from disfacilitation through a withdrawal of descending excitatory drives.

This model has come under determined experimental attack in recent years, and it is now established that sympathetic control involves far more than this stereotyped mechanism (reviewed by *Peiss* 1965; *Hilton* 1965, 1966, 1975, 1977; *Smith* 1974). It is now apparent that simple summation at the level of the medulla cannot explain all observations. There are circumstances where the baroreceptor input can be totally over-ridden, in such a way that even non-linear summations are unlikely to provide all the answer (see Sect. 4.4). In addition some sympatho-excitatory effects appear insensitive to baroreceptor inhibition (see *Kahn* and *Mills* 1967). It is also becoming clear that inhibitory effects can occur within the spinal cord, on or near to the final common pathway, the preganglionic neurone. It remains controversial whether this spinal inhibitory mechanism is responsible for baroreceptor control of sympathetic activity, but it does imply an integrative action within the cord and presumably an integrative function for preganglionic sympathetic neurones.

#### 3.3.3.1 Brainstem Inputs to the Cord

The fact that stimulating within the brainstem can excite the sympathetic motoneuronal pool has led to the conclusion that there is a relatively stereotyped bulbospinal excitatory system (see above, and *Alexander* 1946). Recent studies indicate that the sympatho-excitatory pathways are not homogeneous. *Gebber* and his colleagues have provided evidence for two

distinct excitatory systems (Gebber et al. 1973; Snyder and Gebber 1973; Taylor and Gebber 1973; Gebber and McCall 1976) divided into the pattern first described by Kahn and Mills (1967) and subsequently Nathan (1972). This conclusion rested on the fact that although electrical stimulation at sites in the medulla (Gebber et al. 1973; Snyder and Gebber 1973; Taylor and Gebber 1973; Gebber and McCall 1976), midbrain and hypothalamus (Gebber et al. 1973; Snyder and Gebber 1973) excite sympathetic activity, two patterns of response can be discerned: one is a short latency and relatively constant response, and the other, which may be evoked concomitantly from the same site, has a longer latency and is variable in appearance. In addition, the "early" response is resistant to baroreceptor stimulation and the "late" is totally suppressed by baroreceptor inputs. This suggests that the inhibition of the late response is mediated within the spinal cord. Other evidence to support this model will be reviewed below (see Sect. 3.4). The resistance of the "early" excitatory input to sympathetic neurones to baroreceptor stimulation, may not be total. *Coote* and *Macleod* (personal communication) have shown that "early" excitatory responses evoked in thoracic white rami by stimulation in descending sympatho-excitatory tracts of the cervical cord can be inhibited during powerful baroreceptor stimulation.

Medullary Pathways. The "pressor" area is considered to lie in the lateral areas of the medulla (see Alexander 1946). Gebber et al. (1973) showed that stimulating within this area, particularly within the n. reticularis ventralis and the LRN, evoked discharges in the postganglionic external carotid nerve over two pathways. Furthermore, Taylor and Gebber (1973) have described the effects of such stimulation on single preganglionic neurones, the early response (latency 21.4 ± 1.0 ms) as against the late response  $(55.4 \pm 3.8 \text{ ms})$  was resistant to both baroreceptor activation and to stimulation in depressor areas of the medulla. In addition, later experiments have shown that stimulating in these areas of the medulla also excited non-antidromically driven sympathetic "interneurones" located close to preganglionic neurones (T1-5) identified by their antidromic response to stimulating the cervical sympathetic (Gebber and McCall 1976, and see above). The division into sympathetic preganglionic neurones and "interneurones" may not be as firm as Gebber and McCall claim; all that the authors can conclude with certainty is that the "interneurones" do not have an axon running in the cervical sympathetic nerve.

These apparently distinct descending excitatory systems are differentially affected by stimulation in "depressor" areas of the medulla (*Taylor* and *Gebber* 1973; *Snyder* and *Gebber* 1973; *Gebber* and *McCall* 1976). The pathway responsible for the early excitatory response (which is resistant to "depressor" area and baroreceptor stimulation) conducts at around 5 m/s, and the pathway whose effects are powerfully suppressed by both baroreceptors and the "depressor" area conducts at up to 2 m/s. *Seller* (1973) describes the effects of stimulation of the brainstem on sympathetic preganglionic neurones with a latency broadly compatible with the range described above, but without discerning functionally distinct effects.

Considerably faster pathways exerting excitatory effects on sympathetic activity have been described (*Alanis* et al. 1966; *Calaresu* and *Henry* 1970; *Henry* and *Calaresu* 1974a-d). In particular, stimulation in the parahypoglossal area (*Alanis* et al. 1966; *Calaresu* and *Henry* 1970) excites sympathetic neurones (of the inferior cardiac nerve) with a latency of 3-10 ms (conduction velocity 45 m/s). It has already been shown that this area of the medulla receives a baroreceptor input (*Lipski* et al. 1975), but since this is an excitatory input, it is difficult to formulate an explanation for the relationship of this excitatory input to this sympatho-excitatory pathway. *Alanis* et al. (1966) argue that it represents a descending pathway (i.e. fibre tract) arising from the hypothalamus, so there may be no significant connection between the baroreceptor input to and the efferent effects of stimulation of the parahypoglossal area.

In alternative studies *Henry* and *Calaresu* (1974a) have described field potentials and unit responses in the NPR and LRN with a latency of around 1.7 ms on stimulating within the intermediolateral cell column of upper thoracic levels at points which evoked cardio-acceleration. They considered that they were activating descending excitatory and inhibitory pathways converging onto sympathetic preganglionic neurones (Henry and Calaresu 1974a-d), but since the calculated conduction velocities of these putative pathways were around 60 m/s it is impossible to reconcile them with the observations of others (see above). Even more serious was their failure to demonstrate that they were not activating fibres en passant, without direct functional connection to sympathetic preganglionic neurones. This is especially likely, as the intermediolateral horn is adjacent to the lateral funiculus. In this context, the conduction velocities quoted are very similar to those of the bulbospinal respiratory neurones, whose axons traverse the lateral funiculus (see Bianchi 1971; Euler et al. 1973a, b; Merrill 1974; Richter et al. 1975, and others).

In an attempt to reconcile their results with those of others, *Henry* and *Calaresu* (1974d) have stimulated at cardio-accelerator and cardio-inhibitory sites in the medulla recording in the intermediolateral horn at T2. They claim two patterns of response, one of short latency (2.4-5.0 ms, mean 3.0 ms) and another of long latency (12-40 ms, mean 20 ms), which they consider to act at spinal interneurone and preganglionic neurone respectively, but they provide no physiological identification of the neurones investigated. Unfortunately their presumed interneuronal response, which is equivalent to their previous data in terms of latency (*Henry* and *Calaresu*)

1974a), is at variance with the data of *Gebber* and *McCall* (1976), where the interneuronal response is within the overall range of that seen in preganglionic neurones although the modal latency is somewhat less  $(31 \pm 3)$  ms compared to  $41 \pm 6$  ms for preganglionic neurones). *Henry* and *Calare*su's observations can only have any significance if there are complex propriospinal pathways (see *Kirchner* et al. 1975a) but this has yet to be convincingly demonstrated.

Hypothalamic Pathways. The first demonstrations of brainstem control of sympathetic activity were those showing that simulation of the hypothalamus could evoke increases in the activity of post- and preganglionic sympathetic fibres (Bronk et al. 1940, and subsequently others). In the main, studies have since been concerned more specifically with the pathways from the hypothalamus affecting arterial blood pressure (see Abrahams et al. 1960; Schramm and Bignall 1971, amongst many others). Most results suggest that the efferent pathways involve connections within the medulla, but anatomical evidence is accumulating that there may be direct connections from the hypothalamus to the spinal cord, and particularly to the intermediolateral horn of the thoracic and lumbar cord (Beattie et al. 1930; Smith 1965; Kuypers and Maisky 1975; Saper et al. 1976).

More recent studies using spinal injections of HRP (*Kuypers* and *Maisky* 1975; *Saper* et al. 1976) are particularly revealing because they suggest a pathway from perifornical regions of the hypothalamus descending directly to the intermediolateral column of the thoracic cord. Moreover, *Saper* et al. (1976) have demonstrated the anterograde transport of labelled amino acids from the hypothalamus to this region of the cord, arguing for true terminal endings in the intermediolateral column. They have also shown that pathways from the hypothalamus terminate within regions of the midbrain and medulla previously implicated in cardiovascular control. The relationship of these areas of the midbrain and hypothalamus to those stimulated in former studies that resulted in sympathetic activation (see *Gebber* et al. 1973) remains unresolved.

# 3.3.3.2 Spinal Sympatho-Excitatory Pathways

A specific locus of a sympatho-excitatory pathway has been identified in the spinal cord of the cat. This pathway descends in the dorsolateral funiculus (*Illert* and *Gabriel* 1972; *Gebber* et al. 1973; *Coote* and *Macleod* 1974a, b; *Foreman* and *Wurster* 1973; *Henry* and *Calaresu* 1974b, c; *Kirchner* et al. 1975; *Barman* et al. 1976; *Szulcyck* 1976; *Achari* et al. 1978; *Geis* et al. 1978). Lesions in this pathway have been shown to reduce blood pressure and to abolish the cardiovascular responses to bilateral carotid occlusion in acute experiments in the cat (*Foreman* and *Wurster* 1973), but chronic experiments in the dog show that blood pressure reverts to normal levels although the response to carotid occlusion remains absent (*Geis* et al. 1978). The interpretation of these data may be complicated by the fact that this part of the dorsolateral funiculus also contains descending axons of at least two sympatho-inhibitory pathways, one of which has also been implicated in mediating the inhibitory control of sympathetic activity during the baroreceptor reflex (see Sect. 3.3.3.3).

In addition to this established pathway, there is evidence that central respiratory drive, and in particular inspiratory drive, can effect sympathetic efferent activity (*Polosa* 1968; *Mannard* and *Polosa* 1973; *Preiss* et al. 1975; *Preiss* and *Polosa* 1977; *Lipski* et al. 1977). Since the excitability of preganglionic, and indeed postganglionic, vasoconstrictor neurones (see above) is so powerfully influenced by inspiratory activity, it is likely that the bulbospinal pathways from medullary respiratory neurones are involved in a control or background drive to sympathetic neurones, mediated no doubt by spinal interneurones (see for example *Lipski* et al. 1977).

### 3.3.3.3 Descending Inhibitory Control

The medulla, and indeed other levels of the neuraxis, have been known to be capable of reducing arterial blood pressure through an inhibition of sympathetic activity. The medullary "depressor" area was considered to encompass the medial reticular formation, extending medially from the intramedullary course of the hypoglossal nerve tract (Alexander 1946). Its action was supposed to be mediated by an inhibitory action on the "pressor" area (see above), and in consequence baroreceptor control of sympathetic activity was mediated by this medullary reciprocal network. In fact data have accumulated showing that it is very unlikely that the medial reticular formation and the midline raphe system are involved in baroreceptor reflexes. Lesions of this area have been shown to leave baroreceptor reflexes unaffected (see Sect. 2.2.3.3). There is, however, little doubt that the medial regions of the medulla, i.e. the classical "depressor" area, can evoke profound changes in the discharge of sympathetic neurones (Gootman and Cohen 1971; Taylor and Gebber 1973; Gebber et al. 1973; Kirchner et al. 1975a, b; Gebber 1976; Gebber and McCall 1976) and can exert a tonic control of somato-sympathetic reflexes (Coote and Sato 1978, and others).

In this section an attempt will be made to evaluate the role of this area in the control of sympathetic activity and to establish whether other areas of the medulla can exert a significant inhibitory control of sympathetic activity. The role of these areas in mediating baroreceptor reflexes will also be discussed.

The Nucleus of the Tractus Solitarius. The connections of the NTS with the spinal cord were described in Sect. 2.2.1. It seems that there may be pathways independent of the bulbospinal respiratory pathway, descending from the NTS to the spinal cord and which may have a role in baroreceptor-mediated inhibition (*Hildebrandt* 1974; *Lipski* and *Trzebski* 1975; *McCall* et al. 1977).

In this context *Lipski* and *Trzebski* (1975) have shown that some baroreceptor-sensitive neurones in the vicinity of the NTS can be activated antidromically on stimulating within the cervical spinal cord. This pathway conducted at  $29 \pm 5.2$  m/s, but as it was activated by stimulation at C4 no details of its termination, and in particular of its relevance to sympathetic control, were obtained. *McCall* et al. (1977) have provided evidence for a pathway from the NTS to the intermediomedial zone of the spinal grey, the pathway having a conduction velocity of  $15 \pm 2$  m/s. They have illustrated that the firing patterns of neurones in the two areas have similar post-R-peaks and are similarly excited by electrical stimulation of sympathetic afferent fibres in the inferior cardiac nerve. These intermediomedial neurones have been considered as candidates for a role as spinal inhibitory interneurone in the baroreceptor reflex pathway (see McCall et al. 1977).

Paramedian Reticular Nucleus; Ventromedial Medulla. There is considerable evidence that stimulating in the vicinity of the NPR and ventromedial medulla produces an inhibition of sympathetic activity (Coote and Downman 1969; Gootman and Cohen 1971; Gebber et al. 1973; Taylor and Gebber 1973; Coote and Macleod 1974b, 1975; Henry and Calaresu 1974d; Kirchner et al. 1975a, b; Gebber 1976). In many studies the latency of the evoked inhibition has been shown to be brief, in fact of an order of magnitude less than the central delay of the baroreceptor reflex (see Coote and Downman 1969; Gootman and Cohen 1971; Coote and Macleod 1974b, 1975; Henry and Calaresu 1974a; Gebber and McCall 1976). It would appear that the descending pathway from this region passes in the ventral funiculus of the spinal cord (Henry and Calaresu 1974b; Coote and Macleod 1975), and that this must be sectioned to permit the increase in the spinal component of somato-sympathetic reflexes, especially in the case of those into the cardiac sympathetic nerves (Coote and Sato 1978). In other words, it would appear to mediate a tonic inhibition of the spinal component of somato-sympathetic reflexes.

It would seem that this pathway conducts at a fairly rapid rate (ca. 12.0 m/s – see Gebber and McCall 1976) and the short latency would make its involvement in the baroreceptor reflex, where the central delay is so long, unlikely (see Sect. 3.3). To this must be added the fact that lesions, which may be extensive, in this area of the medulla (see Löfving 1961; Hilton and Spyer 1971; Barman and Gebber 1978a, and others) do not impair baroreceptor function. These observations are particularly significant because in other circumstances Gebber and his colleagues argue that

the inhibition evoked from this area is analogous to baroreceptor-mediated inhibition of sympathetic activity (see *Snyder* and *Gebber* 1973; *Taylor* and *Gebber* 1973, 1975; *Gebber* and *McCall* 1976; *McCall* et al. 1977).

Raphe-Spinal Neurones. The inhibition that may be evoked from the midline, mainly in the n. raphe pallidus, has a long latency (Scherrer 1966; Coote and Macleod 1974a, b, 1975). This long latency is in complete disagreement with the latencies quoted by Henry and Calaresu (1974a). Coote and Macleod (1974a) have provided evidence that 5-hydroxytryptamine-(5HT)-containing neurones may make up the sympatho-inhibitory pathway descending from the raphe which passes via the dorsolateral funiculus. Subsequent studies have shown that lesions of this part of the spinal cord abolished the inhibitory effects of stimulating in the raphe on IC9 to T10 WR reflex (Coote and Macleod 1975), but Henry and Calaresu (1974b) concluded that the fast-conducting pathway originating from the raphe traversed the ventral funiculus! In view of the many other discrepancies between this latter study and those of all other groups, it is probably advisable to treat this observation with caution. Pharmacological depletion of this pathway using p-chlorophenylalanine or 5-, 6-dihydroxytryptamine affected neither ongoing sympathetic activity nor baroreceptor control of sympathetic discharge (Coote et al. 1978).

Ventrolateral Sympatho-Inhibitory Neurones. There is evidence that stimulating within LRN can produce falls of arterial pressure and a reduction in sympathetic efferent activity (Coote and Macleod 1974a), although there is also evidence of sympatho-excitatory effects from stimulating in this area (Thomas et al. 1977). Coote and Macleod (1974a) have shown that the area producing these inhibitory effects corresponds to the area in which the A1 group of noradrenaline-containing neurones are located (Dahlström and Fuxe 1965). These bulbospinal neurones are small and have mainly unmyelinated axons, which agrees with the long latency of the sympatho-inhibitory effects produced by stimulating in this area (Coote and Macleod 1974a, b). From the use of 6-hydroxydopamine lesions (Coote and Macleod 1977) and surgical transections (Coote and Macleod 1975, 1977), it appears that these neurones relay in the dorsolateral funiculus, in the area where Illert and Gabriel (1972) described a sympatho-inhibitory pathway but where there is also evidence for a sympathoexcitatory pathway (see above and Fig. 4). The inhibitory effects of stimulating in the ventrolateral medulla on the spinal component of somatosympathetic reflexes has been shown to be abolished by both surgical and pharmacological destruction of this pathway (Coote and Macleod 1974a, b, 1975, 1977). There is now convincing neuroanatomical data, from studies using both the retrograde transport of HRP and the anterograde transport

of labelled amino acids, that axons from the LRN terminate in the intermediolateral cell column (*Martin* et al. 1979).



Fig. 4. Diagram of a transverse section of spinal cord (C 3-4) to show distribution of points, obtained in seven experiments, where electrical stimulation caused either an inhibition ( $\bullet$ ) or an excitation ( $\bullet$ ) of renal nerve activity. Unanaesthetised decerebrate cats, spinalised at C1. Figure has been modified to illustrate *a* course of bulbospinal noradrenergic neurones descending from LRN; *b* raphe-spinal neurones; *c* descending pathway from the ventromedial medulla. See text for further details. Adapted from Coote and Macleod (1974a)

The Spinal Course of Inhibitory Pathways. The preceding sections have described the three main medullary areas with bulbospinal inhibitory projections. The anatomical location of these pathways in the spinal cord and their characteristics have been established by several studies (Illert and Seller 1969; Illert and Gabriel 1972; Coote and Macleod 1974a, 1975; Kirchner et al. 1975a, b) and are summarised in Fig. 4.

The implication is that pathways from the medulla can act either directly or via spinal interneurones on the preganglionic sympathetic neurones. It is now widely accepted that somato-sympathetic reflexes, which involve a spinal pathway as well as a supraspinal loop (reviewed by Sato and Schmidt 1973) are under bulbar inhibitory control. For example, recent studies in which the upper cervical cord has been cooled to block descending inhibitory (and for that matter excitatory pathways) have revealed both an increase in the magnitude and shortening of the latency of intercostal to T3 or T4 white rami reflex (*Dembowsky* et al. 1978). Naturally this type of experiment will not reveal which pathway or pathways are involved, but Coote and Sato (1978) have demonstrated that, with respect to somatic (and visceral) reflexes into cardiac sympathetic nerves, sections of the dorsolateral funiculus are ineffective in releasing the spinal component of the reflex. This would implicate the ventromedial medulla and its efferent pathway through the ventral funiculus in the control of these reflexes (see *Coote* and *Macleod* 1975). It would seem that destruction of this ventral pathway is ineffective in altering the baroreceptor reflex control of sympathetic activity (*Illert* and *Seller* 1969; *Coote* and *Macleod* 1975). However, there is little doubt that the other two inhibitory systems can equally well affect somato-sympathetic reflexes, but presumably they do not exert a tonic control.

For baroreceptor control of sympathetic activity, previous considerations must lead to a discussion of the role of the dorsolateral funiculus as the pathway for its action. This could result from the interplay of both descending excitatory and inhibitory pathways (see *Illert* and *Seller* 1969; *Foreman* and *Wurster* 1973; *Coote* and *Macleod* 1974b, 1977; *Geis* et al. 1978).

With respect to the involvement of descending inhibitory pathways, the noradrenergic neurones in the LRN (sending axons in the dorsolateral funiculus) are a strong candidate (*Coote* and *Macleod* 1974b, 1977). At present the conclusion must rest on the fact that the spinal component of IC 10 to T9 WR reflex is partially suppressed by the electrical stimulation of the sinus nerve and that this suppression is reduced by lesions in the dorsolateral funiculus (*Coote* and *Macleod* 1974b) or by pharmacological destruction of noradrenergic neurones (*Coote* and *Macleod* 1977).

However, other data have been used to counter, or at least question, this claim. Electrical stimulation of the sinus nerve evokes concurrent activation of both chemoreceptor and baroreceptor afferents, the former evoking an increase in sympathetic activity. Accordingly, the apparent depression could result from a "silent period" following the excitatory effect of chemoreceptor stimulation. Szulczyk (1976) has shown that the excitatory effects of sinus nerve and hypoglossal nerve stimulation are blocked by DLF section (cf. Foreman and Wurster 1973 for the effects of such lesions on the response to bilateral carotid occlusion). To distinguish which interpretation is correct, it would seem there is no option but to use single unit recordings.

In the one case in which single sympathetic units have been studied during stimulation of the dorsolateral funiculus, the results are complicated by the fact that this region contains both excitatory and inhibitory pathways (*Kirchner* et al. 1975a). Stimulation normally evoked an early excitatory response followed by a long-lasting "silent period", although in three cases a pure inhibition was seen. As the recordings were made extracellularly at T2 and several of the neurones were activated by the iontophoretic application of glutamate, it would seem likely that the "silent period" could result from a direct inhibitory action. These authors also showed that the inhibition evoked from this area of the cord was present in chronic spinal animals, as judged from effects on somatic reflexes into white rami at lumbar levels and the renal nerve (*Kirchner* et al. 1975b).The authors consider that this makes a direct monosynaptic inhibitory action
unlikely to be the only mechanism for spinal inhibition but indicates the involvement of propriospinal pathways (see also *Kirchner* et al. 1975a).

In contrast, Coote and Macleod (1974b, 1977) argue for a direct noradrenergic inhibitory innervation of sympathetic neurones from bulbospinal neurones. This would be a surprising situation as most long-circuited inhibitory effects are mediated by an inhibitory interneurone close to the target neurone. In support of their claim is the fact that the iontophoretic application of noradrenaline to sympathetic preganglionic neurones depresses their activity (Coote and Macleod, unpublished work), although Hongo and Ryall (1966) have provided less clear-cut results on the effects of noradrenaline. There is, however, evidence that the  $\alpha$ -agonist clonidine, given systemically, reduces the size of spinal reflexes onto sympathetic neurones (Dembowsky et al. 1978; Haeusler 1977). As yet the iontophoretic studies would seem to be at an early stage, and the use of specific antagonists to NA on the effectiveness of baroreceptor inputs to the preganglionic neurones would be indicated. Certainly, since baroreceptor stimulation can affect the performance of somato-sympathetic reflexes (see refs. above and Fussey et al. 1973a), an effect that can be produced by activating any of three inhibitory pathways so far identified, there can be little doubt that at least part of the baroreceptor reflex is mediated by an inhibitory action with the cord.

### 3.4 Integrative Control of Sympathetic Activity

There are indications that two general processes are involved in baroreceptor control of sympathetic activity. The first process is not at variance with the generally accepted view that the baroreceptors act by inhibiting descending excitatory pathways. Its extension is that it does not limit this action to a control of medullary excitatory pathways but opens the possibility that there may be parallel channels from the hypothalamus and midbrain which may pass without synaptic connections in the medulla. The second process is the possibility that active inhibition occurs at the level of the spinal cord. From the various studies referred to in the previous sections, a general conclusion can be made that there are pathways, of unknown synaptic complexity, which descend through the spinal cord and can exert a powerful inhibitory control of sympathetic activity. As yet there is no conclusive evidence that the baroreceptor influence is mediated by such a system, although the ability of the baroreceptor input to suppress both medullary and spinally evoked sympatho-excitatory responses is striking. It is probably not too naive to argue that the baroreceptors which have been known to exert such a powerful control of vasoconstrictor and cardio-accelerator "tone" through the sympathetic nervous system are likely to utilise both systems.

Accepting then the premise that baroreceptor control of sympathetic activity may be exerted by both disfacilitation (i.e. control of descending excitatory inputs) and active inhibition within the spinal cord, is it possible to speculate usefully on the connections, at least those acting within the cord, that integrate such a control? It is probably justified for a review to pose such a question, and to conclude this section, two simple models will be proposed as a challenge for subsequent experimental work (see Fig. 5).



Fig. 5a, b. Diagrams of the relationship of bulbospinal spinal pathways in the mediation of the baroreceptor reflex.  $\bullet$  neurones exerting an excitatory function;  $\circ$  neurones exerting an inhibitory function. 1 medulla; 2 spinal cord; NTS nucleus of the tractus solitarius. PSM preganglionic sympathetic motoneurone; IML intermediolateral cell column; IMM intermediomedial cell column. In a the site of convergence is at the level of the PSM, in b an excitatory interneurone forms an important site of integration. Dotted lines refer to pathways of unknown synaptic complexity. Further details in the text

In considering baroreceptor regulation of sympathetic activity we are really only questioning the integrative role of these preganglionic neurones (Fig. 5a). In this model, their output is determined by the interplay of inhibitory and excitatory inputs at the cell body (and presumably its dendrites) of the sympathetic neurone. This does not preclude interactions further back in multisynaptic pathways onto them, but draws an analogy to the situation in the somatic system. Unfortunately there is insufficient data, as yet, to give credence to the presence of excitatory or inhibitory interneurones in the intermediolateral column that impinge on sympathetic preganglionic neurones. The evidence in favour of an excitatory interneurone is the stronger and received indirect support from a study using intracellular recordings from sympathetic neurones (*Coote* and *Westbury* 1974, 1979b). In the latter study the membrane potential was marked by random EPSPs of relatively small amplitude and short duration, with no evidence of large slow potential changes. This observation may be taken as evidence for the fact that their control is exerted by an excitatory interneurone alone (Fig. 5b). In this situation, inhibitory control, either baroreceptor or recurrent, could be exerted by convergence onto this interneurone.

In both models, it is possible to envisage that some descending excitatory control is exerted directly on the preganglionic neurone, in view of the baroreceptor-insensitive excitatory effects that may be elicited from the brainstem (see Sect. 3.3.3.2). It is, however, unlikely that these inputs are totally baroreceptor-insensitive; their apparent insensitivity probably represents only a more synaptically secure input than other descending excitatory systems. Accordingly, its failure to excite sympathetic activity at heightened levels of baroreceptor stimulation would result from the reduced excitatory input, i.e. disfacilation, (see Fig. 5b) or from the power of the induced inhibitory control of sympathetic activity (Fig. 5a).

To complete an already complex arrangement, the neurones of the intermediomedial nucleus may also function as interneurones in the baroreceptor reflex (*McCall* et al. 1977). It is very simple to accommodate their action in either of the schemes presented in Fig. 5. It may be that a combination of these two models will present the most adequate approximation to reality; detailed experimental studies using conventional neurophysiological and neuroanatomical techniques can be expected to resolve this question in due course.

## 3.5 Vagal Control of the Heart

Nervous control of the heart is mediated by the reciprocal discharges of its sympathetic (see above) and vagal innervations. The role of the vagus in exerting a negative chronotropic influence is so well documented that citation is superfluous. In the context of the present review its significance rests on the abundant demonstration that the baroreceptor control of the heart is mediated primarily by the vagal outflow (see *Heymans* and *Neil* 

1958; Schaeffer 1960; Kirchheim 1976; Levy 1977). This basic control mechanism appears to function relatively normally in the decerebrate animal, but until the present the basic circuitry linking the baroreceptor afferent input to the vagal outflow, i.e. the preganglionic vagal neurones, remained in doubt (for discussion see McAllen and Spyer 1978; Spyer 1979). This uncertainty resulted from the fact that although the vagus had been known for at least the last hundred years to have two motor nuclei in the medulla of mammals, controversy had remained concerning which contains the cell bodies of the preganglionic innervation of the heart (Mitchell and Warwick 1955; McAllen and Syper 1976, 1978a; Spyer 1979). Accordingly, most data relating to vagal control has been obtained from the effects of the baroreceptors on heart rate or the activity of preganglionic vagal fibres in the cervical vagus or cardiac branches of the thoracic vagus.

The studies referred to above have been reviewed extensively (*Heymans* and *Neil* 1958; *Kirchheim* 1976; *Levy* 1977), and it is unnecessary to reiterate in detail that in animals with heart rates less than 180 beats/min the vagal innervation exerts a beat-by-beat control of cardiac function (*Levy* 1977). However, since neuroeffector delays are long, in the region of 0.3 ms (*Warner* and *Cox* 1962), this type of study will provide little of value regarding the central integrative responses underlying the observed changes. It is for this reason that the subsequent discussion will concern the activity and control of preganglionic vagal neurones, firstly with respect to fibre recordings and finally with a discussion of recent studies relating to recordings from the cell bodies of these neurones.

## 3.6 Preganglionic Vagal Cardiomotor Neurones

Preganglionic vagal fibres with presumed cardio-inhibitory function have been described as having distinctive firing patterns. They have a variable pulse rhythm, which can be seen as a burst of discharge synchronous with each cardiac cycle, or revealed by constructing histograms of their pulsetriggered or post-R wave-triggered activity (Magruth et al. 1951; Green 1959; Schaeffer 1960; Okada et al. 1961a, b; Weidinger et al. 1962; Jewett 1964; Katona et al. 1970, 1977; Kunze 1972). This cardiac rhythm of baroreceptor origin is usually superimposed on a respiratory-related rhythm; they normally fire in phase with expiration (Rijlant 1936a, b; Weidinger et al. 1962; Iriuchijima and Kumada 1963, 1964; Jewett 1964; Katona et al. 1970, 1977; Neil and Palmer 1975; Davidson et al. 1976). Their overall discharge frequency is low: in the cat vagal "tone" is especially low (see Kunze 1972; McAllen and Spyer 1976, 1978a), whilst in the dog spontaneous activity is more conspicuous. Even so, the overall characteristics of discharge have similarities, and it is rare for vagal efferent fibres to the heart to fire at greater than 20 impulses/s (*Iriuchijima* and *Kumada* 1963, 1964; *Jewett* 1964; *Katona* et al. 1970, 1977; *Kunze* 1972). However, with arterial pressure elevated to around 200 mmHg, activity may reach 30–40 Hz (see *Jewett* 1964; *Katona* et al. 1970, 1977; *Kunze* 1972), although *Okada* et al. (1961b) claim recording discharges with up to 26 impulses/s per cardiac cycle, at a mean frequency of 120 Hz, in the cardiac branches of the cat.

It is pertinent at this point to question the number of cardiac efferent fibres present in the cervical vagus. It is likely that a very few can have a marked effect on heart rate (McAllen and Spyer 1976, 1978a), and it is known that in the cat there are only some 500 B fibres in the cervical vagus which appear to pass to the thoracic vagus or its various branches (Agostini et al. 1957). It is established beyond reasonable doubt that, in mammals, cardio-inhibitory neurones have B fibre axons (Heinbecker 1930; Heinbecker and O'Leary 1933; Brown and Eccles 1934a, b; Grundfest 1939; Middleton et al. 1950). This has been shown by graded electrical stimulation of the cervical vagus, recording the evoked volley and noting the efferent effects on the heart (see also *McAllen* and *Spyer* 1978a). There are, however, equally strong indications that among the population of B fibres so activated are those destined for the lungs, which have a bronchoconstrictor function (Widdicombe 1961, 1966; Jewett 1964) and the oesophagus (Jewett 1964), so that the total number of cardiac efferents is likely to be very much smaller.

Attempts have been made to assess the complexity of connections between the arterial baroreceptors and these vagal efferent fibres by studying either the nature of their response to electrical stimulation of the sinus nerve (*Iriuchijima* and *Kumada* 1963, 1964; *Kunze* 1972) or the properties of their cardiac rhythm (*Jewett* 1964; *Katona* et al. 1970, 1977; *Kunze* 1972). Both these approaches and the data accrued will be discussed later, but as in both cases efferent conduction time is unknown and in the latter afferent conduction time is also unknown, the reflex times obtained can give little indication of the central delay of the reflex and hence of its neuronal complexity. For this purpose it is necessary to have the possibility of recording from the cell bodies of preganglionic vagal neurones.

# 3.6.1 Location of Preganglionic Vagal Neurones

## 3.6.1.1 Anatomical Studies

The existence of two vagal nuclei in the medulla of mammals has been appreciated for at least a hundred years. There had at first been controversy whether one was sensory and the other motor but by the early part of this century it was agreed that both the dorsal (DNV, dorsal motor nucleus of the vagus) and the ventral (NA, n. ambiguus) had motor functions (see

Mitchell and Warwick 1955 for discussion). By the same token, it became accepted that the DNV was a visceral-motor nucleus, the NA having a somatomotor function (Mitchell and Warwick 1955). For this reason, the consensus was that the DNV contained the cardio-inhibitory vagal neurones (Mitchell and Warwick 1955, and most current textbooks of neuroanatomy), although dissenting voices have remained (Kosaka 1909; Szentagothai 1952).

The major evidence for the DNV as the location for these vagal cell bodies was from studies observing the chromalytic changes produced in the vagal nuclei following section of the vagus and its peripheral branches (*Mitchell* and Warwick 1955; Smolen and Truex 1977 for review). The alternative technique of observing peripheral changes after central destruction has provided contrasting results. Szentagothai (1952) showed that lesions within the NA of the cat were followed by signs of Wallerian degeneration in the vagal cardiac branches; similar discrete lesions in the DNV were ineffective in producing such changes (see also Kosaka 1909; Kerr 1967, 1969). The implication is that the cell bodies of cardio-inhibitory neurones are located in the NA. These results are unfortunately not without certain limitations, as the technique of central destruction will destroy fibres en passant as well as cell bodies, and it is known that efferent axons from the DNV pass close to the NA, whilst the axons of the NA may pass dorsomedially towards the DNV (Cajal 1909).

There are as yet few studies on the transport of HRP from the vagal cardiac branches to the medulla (Bennett et al. to be published; Garcia et al. to be published; Nosaka et al. 1979), but several studies have shown the retrograde transport of HRP from the cervical vagus to both vagal nuclei in the medulla (Devito et al. 1974; Miller 1976; Robertson et al. 1976; Geis and Wurster 1978). In general, labelling is more apparent in the DNV than the NA by factor of at least ten (Miller 1976; Robertson et al. 1976). In one study, medullary labelling has been observed after placing HRP on the sinoatrial and atrioventricular nodes of the heart in the cat (Todo et al. 1977) on the basis that this would label preganglionic vagal neurones. Unfortunately this is not without serious inherent problems: HRP is readily transported in the blood and would be expected to enter the central system whenever the blood-brain barrier is ineffective. It is hence not surprising that labelling was seen in the vicinity of the NTS and DNV close to the area postrema, particularly as no evidence was provided that the material was intracellular.

In a similar study, *Geis* and *Wurster* (1978) have shown that epicardially placed HRP is transported to the brainstem. The majority of labelled neurones (78%) were seen in the NA, just 5% in the DNV and the rest in between these nuclei. Similarly, *Bennett* et al. (to be published) achieved equivalent neuronal labelling from identified cardiac branches in both NA

and DNV, whilst *Garcia* et al. (to be published) noted that labelling from cardiac branches was mostly restricted to the NA, although pulmonary branch axons appeared to originate from neurones in both NA and DNV. *Nosaka* et al. (1979) have identified labelled neurones in both the DNV and NA of the rat, after embedding cardiac vagal branches in HRP, and also in the region between these two nuclei. Since it is apparent that cardiac branches are often contaminated with fibres destined for the lungs, it is impossible to conclude very much from these studies in the absence of physiological data. The labelling in the DNV from cardiac branches might represent the efferents with unmyelinated axons described by *McAllen* and *Spyer* (1976) and demonstrated to be profuse by *Bennett* et al. (to be published).

# 3.6.1.2 Destructive Lesions in the Medulla: Cardiac Effects

The effects of electrolytic lesions in the medulla on vagally mediated bradycardia were summarised earlier (Sect. 2.2.3.3). Broadly, destruction of the DNV is ineffective in abrogating such effects (*Kerr* 1967, 1969; *Bori*son and Domjan 1970), whilst lesions in the NA were effective in the cat (see Lee et al. 1972).

# 3.6.1.3 Neurophysiological Studies

*Electrical Stimulation.* Physiological studies concerning the site of origin of preganglionic vagal neurones supplying the heart, based on electrical stimulation of the medulla, have favoured either or both vagal nuclei (*Miller* and *Bowman* 1915; *Chiurugi* and *Mollica* 1954; *Calaresu* and *Pearce* 1965b; *Gunn* et al. 1968; *Thomas* and *Calaresu* 1974a; *Chen* and *Chai* 1976; *Dugin* et al. 1976). It is worth recalling that the efferent axons from each nucleus may pass in close proximity to the other nucleus (*Cajal* 1909) and hence may account for at least part of the confusion. Equally, current spread from the point of stimulation may account for at least some of the observed changes. This is particularly so in the case of the DNV, which is located close to the NTS, where baroreceptor afferents terminate. Accordingly, bradycardia mediated by vagal fibres can result from stimulating any part of the baroreceptor reflex pathway. Stimulation elsewhere in the CNS may well evoke a bradycardia by a similar process.

There may well be a species difference in the location of these vagal cardio-inhibitory neurones, which may also account for some of the differences in the literature. In the cat, vagal bradycardia can be obtained by stimulating the NA (*Gunn* et al. 1968; *Thomas* and *Calaresu* 1974a; *Chen* and *Chai* 1976) but not the DNV (*Calaresu* and *Pearce* 1965b; *Gunn* et al. 1968). In the dog, *Gunn* et al. (1968) report eliciting vagal bradycardia from both sites. In contrast there are claims that vagal bradycardia can be evoked from stimulating the DNV in the cat (*Miller* and *Bowman*).

1915; *Chiurigi* and *Mollica* 1954; *Dugin* et al. 1976). It is worth noting that the bradycardia elicited from the NA is abolished by ipsilateral vagotomoy in the cat whilst that evoked from the NTS and DNV is only partially reduced by ipsilateral vagotomy (*Thomas* and *Calaresu* 1974a).

Recording Experiments. Calaresu and Pearce (1965a) were unable to record activity correlated with the cardiac cycle in the DNV of the cat. This was perhaps not unexpected in view of the data from stimulating and lesion studies (see above). In conclusion, in the cat it appears likely that the NA would contain vagal cardio-inhibitory neurones. This has been confirmed directly in recent studies (*McAllen* and *Spyer* 1975, 1976, 1977, 1978a, b). In the cat, they have been shown that there are neurones in the NA which can be antidromically activated by stimulating cardiac branches at intensities which evoke cardiac slowing (*McAllen* and *Spyer* 1976, 1978a) and that these neurones have axons which are in the range of B fibres on the basis of calculated conduction velocity. In the same study similar neurones were not found in the DNV; furthermore, on studying a large number of neurones in the DNV relaying to the cervical vagus, none with B fibres was found to relay into the cardiac branches (*McAllen* and *Spyer* 1976; Fig. 6). Three neurones with axons in the C fibre range were,



Fig. 6. The positions of 46 cardiac efferent neurones are shown on four standard sections of the medulla taken at obex level, and at 1-mm intervals rostrally. Inserts, 2 mm square, show details of their relation to the structure of the nucleus ambiguus. TS tractus solitarius; DNV dorsal motor nucleus of the vagus; NA nucleus ambiguus. McAllen and Spyer (1976)

however, seen to relay to the cardiac branches, which could explain certain of the observations of *Todo* et al. (1977), *Geis* and *Wurster* (1978) and *Bennett* et al. (to be published).

This demonstration that vagal neurones with B fibre axons originate in the NA does not on its own prove a cardio-inhibitory function. Among the B fibres in the thoracic vagus are bronchoconstrictor vagal neurones (Widdicombe 1961, 1966; Jewett 1964) and oesophageal-motor vagal neurones (Jewett 1964). Physiological studies have added still further support to the original suggestion (McAllen and Spyer 1977, 1978a, b). These have involved analysing the firing patterns of identified cardiac-branch-projecting neurones, firing patterns from both spontaneously active neurones, and those induced to fire by the iontophoretic application of DL-homocysteic acid (DLH) or glutamate. A major group of neurones with an expiratory firing pattern (the majority were usually silent until the expulsion of excitant amino acid) have been observed and were shown to have a cardiac rhythm of baroreceptor origin in their discharge (McAllen and Spyer 1978a, b). These neurones, when activated by amino acid, were capable of slowing the heart (McAllen and Spyer 1977, 1978a), which confirms the impression that they represent cardiac vagal motoneurones.

A second group of neurones with B fibre axons were also identified as projecting to either cardiac branches or lung branches of the thoracic vagus. These neurones were usually spontaneously active, discharging with an inspiratory rhythm. They were not excited by the arterial baroreceptors (*McAllen* and *Spyer* 1978a) but were excited by laryngeal stimulation and would seem equivalent to the vagal efferent fibres described by *Widdicombe* (1961, 1966) as bronchoconstrictor in function. There was also a degree of spatial separation of the two classes of vagal preganglionic neurone within the NA (see Fig. 7). The bronchoconstrictor neurones tended to be more rostrally and dorsally placed in the nucleus, extending into the n. retrofacialis.

As suggested previously there may be species differences regarding the location of cardiac vagal motoneurones. In the pigeon it is clear from neurophysiological studies that they are located in the DNV (Schwaber and Cohen 1978a, b), but it is generally considered that this species has a single vagal motor nucleus, the DNV. In the dog there is evidence that the NA contains cardiac vagal motoneurones, i.e. neurones activated antidromically from cardiac branches of the right vagus (McAllen and Spyer unpublished work), but no data exists regarding the DNV. In the rabbit, however, there is evidence in favour of their localisation in the DNV. Schwaber and Schneiderman (1975) and Kaufman et al. (1979) have described in the DNV of the rabbit neurones whose axons projected in the cervical vagus and which were, on the basis of conduction velocity, B fibres. These they considered to be cardio-inhibitory: their spontaneous and evoked discharge varied inversely, but proportionally, with heart rate and they were located in the area of the DNV which elicited a low-threshold bradycardia on electrical stimulation. Moreover, the discharge of these neurones evoked by the iontophoresis of DLH elicited a bradycardia (*Jordan* et al. 1979). In a recent study, *Jordan* et al. (1979) have described a population of vagal neurones with B-fibre axons which are located in both DNV and NA. These neurones have been shown to have identical properties to CVMs described in the cat (see above).



Fig. 7. a The position of 21 CVMs ( $\bullet$ ) and nine BVMs (X) on five standard sections of the medulla taken at obex level, 1, 2, 3 and 4 mm rostral to the obex. *DMN* dorsal motor nucleus of the vagus; *NA* nucleus ambiguus; *NRF* nucleus retrofascialis. **b** and **c** show histograms of the conduction velocities of cardiac and bronchomotor units, respectively. *McAllen* and *Spyer* (1978a)

### 3.6.2 Baroreceptor Input to Preganglionic Vagal Neurones

The influence of the arterial baroreceptors on vagal preganglionic neurones is well documented (*Marguth* et al. 1951; *Green* 1959; *Schaeffer* 1960; *Okada* et al. 1961a, b; *Weidinger* et al. 1962; *Jewett* 1962, 1964; *Iriuchijima* and *Kumada* 1963, 1964; *Katona* et al. 1970, 1977; *Davidson* 

et al. 1976), but there is considerably less quantitative data available on this subject. Furthermore, even the quantitative information available is in a form which does not allow any conclusions to be drawn regarding the central mechanisms and pathways linking the baroreceptor afferent input to these neurones.

In the main, studies have fallen into two categories. Firstly, the effects of electrical stimulation of the SN on vagal efferent fibres located in the cervical vagus (*Iriuchijima* and *Kumada* 1964) and vagal cardiac branches (*Iriuchijima* and *Kumada* 1963; *Kunze* 1972) have been observed. Secondly, attempts have been made to analyse the pulse-related discharge of similar vagal efferent fibres (*Jewett* 1962, 1964; *Weidinger* et al. 1962; *Katona* et al. 1970, 1977; *Kunze* 1972).

In the cat, *Kunze* (1972) has shown that stimulation of the SN excites efferent fibres in the vagal cardiac branches with a latency in the range of 26-90 ms, and their probability of discharge was maximal at 70–400 ms, in different units, after the rise in systolic pressure. With increases in arterial pressure the discharge of neurones rose to a peak instantaneous frequency of up to 40 Hz. In similar experiments in the dog, *Iriuchijima* and *Kumada* (1963) report a latency of 60-100 ms for evoked responses in cardiac branch fibres and 50-100 ms for fibres in the cervical vagus (*Iriuchijima* and *Kumada* 1964) on stimulating the SN. They also noted that the stimulus was most effective during expiration, responses rarely being observed from stimuli occurring during inspiration (*Iriuchijima* and *Kumada* 1964; for discussion see Sect. 4.3).

There is general agreement that the pulse-related discharge of vagal efferent fibres in the cervical vagus occurs some 50-250 ms after the systolic rise in aortic pressure (Jewett 1964; Katona et al. 1970). Jewett (1964) showed that the first peak in histograms of post-R-wave-triggered vagal activity occurred some 60-240 ms after the systolic rise in pressure, a second peak related to the dichrotic notch following some 180-300 ms after the first peak. Katona et al. (1970) describe the first peak at 50-40 ms (mostly around 80 ms), the second 80-200 ms after the dichrotic rise in pressure. This contrasts with observations of Weidinger et al. (1962), in the cat, which suggest that the peak in vagal activity follows 45-55 ms after the R wave of the ECG. Since the aortic pressure starts to rise some 60 ms after the R wave of the ECG (see Jewett 1964; Katona et al. 1970), it is unlikely that the cardiac rhythm of these multifibre preparations can result from inputs generated from the arterial baroreceptors, except in the case where the peak is related to the preceding arterial pulse, which might be possible at rapid heart rates.

In order to overcome the problems in interpreting the above data in terms of the reflex time of the baroreceptor-vagal reflex, *McAllen* and *Spyer* (1978b) have made accurate timings of the probability of discharge

of CVMs, in relation to the arterial pulse. These were recorded in the NA of the cat, the afferent barrage from the carotid sinus also being monitored. In a proportion of their preparations the aortic baroreceptors had been denervated, and as the cardiac rhythm of CVMs was then obliterated by bilateral carotid occlusion, it was possible to conclude that this rhythm depended on an input from sinus baroreceptors alone. The pulse-related peak followed the increase in SN activity by from 20-110 ms in individual neurones (*McAllen* and *Spyer* 1978b, see Fig. 8). In a proportion of these



Fig. 8a-c. The cardiac rhythm of a CVM (aortic baroreceptors denervated). Pulse-triggered histograms (256 cycles superimposed, 10 ms bin width) of CVM activity (upper trace) and sinus nerve activity (lower trace). Femoral pulse wave form was averaged simultaneously with the same bin width (middle trace). a Before, b during and c after bilateral carotid occlusion. The unit was firing in response to 17 nA DLH. McAllen and Spyer (1978b)

neurones the effect of SN stimulation was also tested. Invariably the latency of the response was significantly shorter than that measured for natural baroreceptor stimulation (*McAllen* and *Spyer* 1978b; *Spyer* 1979). With the reservation that these values of reflex time are not an accurate measure of central delay (see *Spyer* 1979 for discussion), they do provide an indication of the complexity of connection in the reflex pathway. The reflex time at physiological levels of blood pressure is surprisingly long, considering that these values eliminate efferent conduction (see Fig. 8). This makes it unlikely that the baroreceptor-vagal reflex involves a simple fast-conducting disynaptic pathway (*McAllen* and *Spyer* 1978b; *Spyer* 1979). Baroreceptor afferents are known to terminate in the NTS (Sect. 2.1 and 2.2), and the NTS projects to the NA (Sect. 2.2), but the complexity of connections within both the NTS and NA remains unresolved (*Spyer* 1975, 1979; *McAllen* et al. 1979) so it is not as yet possible to eliminate a role for supramedullary connections in the normal reflex pathway.

# 3.6.3 Supramedullary Inputs to Preganglionic Vagal Neurones

The previous sections have outlined the putative ascending projections in the baroreceptor reflex pathway and have indicated a role for suprabulbar and diencephalic areas in the reflex. Some of the pathway between the baroreceptor input and CVMs has been indicated above, and it is sufficient at this point to emphasise that there is both anatomical and physiological evidence for hypothalamic pathways descending to areas of the medulla known to contain preganglionic vagal neurones. From the use of retrograde transport HRP and the anterograde transport of tritiated amino acids, Saper et al. (1976) described pathways emanating from the hypothalamus and ending in the NA, NTS, and close to the DNV. Swanson (1977) has further demonstrated that descending fibres from the paraventricular nucleus of the hypothalamus containing the carrier protein neurophysin project to or through the NA, DNV, NTS, and lateral tegmental field at the nucleus intermedius. The role of these connections remains to be resolved, and the physiological data relating to suprabulbar control of vagal activity will be considered in subsequent sections (see also Fig. 13, p. 100).

## 4 Modulation of the Baroreceptor Reflex

The previous sections have detailed the current position concerning the central pathways involved in mediating the baroreceptor control of sympathetic and vagal efferent activity. The details are, however, as yet incomplete. A further set of experimental studies have provided information on both the organisation and control of the baroreceptor reflex. These have involved studies on the interaction between the different baroreceptor afferent inputs, between baroreceptors and other peripheral afferent inputs, and finally the influence of stimulation within the brainstem and cerebellum on the efficacy of the baroreceptor reflex. These approaches are diverse, yet the observations have revealed some physiological phenomena that may indicate common synaptic processes in the integration of these various inputs.

# 4.1 Interactions Between Carotid Sinus and Aortic Arch Baroreceptor Inputs

From numerous reports it has been accepted that there is a broad degree of similarity in the receptor properties of the baroreceptors of the carotid sinus and aortic arch (reviewed by *Kirchheim* 1976), although there may be differences in their dynamic sensitivity (see *Landgren* 1952; *Angell-James* 1971a,b), and it has been suggested that they may act over different ranges of pressure (*Pelletier* et al. 1972; *Shepherd* 1973). Since this latter suggestion depended on observations of whole nerve activity, its applicability to the normal in vivo situation is questioned; *Samodelov* et al. (1979) consider that the working ranges and sensitivities of individual receptors from the two areas are in fact similar, at least in the cat.

There is, however, good evidence that the quantitative effects of the two groups of baroreceptors on vasomotor activity and the heart may be different. Angell-James and Daly (1970) have shown that at static pressures within the physiological range, vascular resistance is reduced to a greater extent by a change in sinus pressure than an equivalent change in pressure in the aortic arch. Such differences were not seen during perfusion with pulsatile pressures. These quantitative differences may reflect either differences in the distensibility properties of the two areas or a central phenomenon (Angell-James and Daly 1970). The latter could be due to afferent density or differential effects on central neurones, or both. These authors also demonstrated that simultaneous stimulation of the aortic arch and carotid sinus baroreceptors evoked responses of greater magnitude than stimulating either alone, although the apparent summation resulted in an effect that was less than the algebraic sum of the individual effects. This type of summation has been demonstrated by others (Stegemann and Müller-Büton 1966; Kotter et al. 1970; Warzel and Brattström 1972; Katona and Tan 1975).

Such interactions in the control of vascular resistance imply a form of central summation, but *Wang* and *Borison* (1947b) argue for an occlusive interaction between baroreceptor inputs from the two carotid sinuses, although they readily demonstrated facilitation in the baroreceptor-cardiac reflexes mediated by the vagal efferents. Such a facilitation of vagally mediated cardiac effects appears not uncommon, representing either an intrinsic property or extrinsic control of the vagal preganglionic neuronal pool.

In an earlier section (2.2) the termination of AN and SN afferents in the medulla was described, and mention was made of attempts to identify the convergence of these inputs onto second-order neurones in the NTS. Suggestions for convergence have resulted from monitoring evoked potentials concurrently at the same site on stimulating the AN and the SN (Hellner and Baumgarten 1961; Biscoe and Sampson 1970a; Gabriel and Seller 1970), but single unit studies have as yet failed to demonstrate convergence at this level of the brainstem (McAllen 1973). It is, however, often observed on medullary neurones located beyond the confines of the NTS (Biscoe and Sampson 1970b; McAllen 1973), which presumably represent later stages in the reflex pathway.

Direct studies on the interactions of SN and AN inputs are few (Biscoe and Sampson 1970b; Gabriel and Seller 1970; McAllen 1973; Jordan 1977; Jordan and Spyer 1978b), and all are complicated by the fact that electrical stimulation of SN evokes a concomitant activation of chemoreceptor afferents (as also occurs with AN stimulation in the cat). Gabriel and Seller (1970) have shown that the evoked response to electrical stimulation of the SN recorded in the NTS is modified by a conditioning stimulus to the AN, and vice versa. Equally, the response to one input can be modified by a conditioning stimulus to the same nerve, the response to SN stimulation at 10 Hz being only 50% of that observed at 1 Hz (Seller and Illert 1969; Gabriel and Seller 1970). Similarly, contralateral inputs from SN and AN can modify the responses to ipsilateral inputs (Gabriel and Seller 1970). It may be that the first synapse represents a frequency-limiting step in the reflex pathway, since the cardiovascular responses to SN stimulation are maximal at 20–30 Hz, while those from stimulating the NTS are maximal at 120 Hz (Seller and Illert 1969). These apparently occlusive interactions provide an explanation for the observations of Wang and Borison (1947a, b) on the vascular component of the baroreceptor reflex, but are clearly at variance with the simultaneous observations of summation (or facilitation) in the cardiac arm of the reflex. These studies may thus indicate an early decussation of the reflex, which was also implied by the observation of Lipski et al. (1976) on chemoreceptor-baroreceptor interactions in or close to the NTS.

If the data from evoked potentials can be taken to indicate a convergence of these baroreceptor inputs at the first synapse, the interaction is most certainly postsynaptic. *Gabriel* and *Seller* (1970) and *Jordan* and *Spyer* (1978b) have failed to reveal presynaptic interactions of SN and AN afferent fibres. Conditioning stimuli in either AN or SN failed to alter the magnitude of the antidromically evoked potential in the other nerve of the pair elicited by stimulation in the NTS (*Gabriel* and *Seller* 1970; *Jordan* and *Spyer* (1977, 1978a, 1979) have failed to demonstrate presynaptic influences on either SN and AN afferent terminals in the medulla, a result which confirms the impression of *Rudomin* (1968) that although AN inputs could evoke presynaptic influences on other vagal afferent inputs at this level, they were themselves not amenable to presynaptic modulation. Thus, within the limits of the techniques so far applied, it would seem that interactions between AN and SN occur postsynaptically at the first synapse or at subsequent synapses in the reflex pathway.

In resolving the connections within the NTS that may account for convergence, it would appear that there are many complicating factors. For SN inputs alone it appears that some neurones may receive both monosynaptic and polysynaptic input (*Humphrey* 1967; *McAllen* 1973). Furthermore, any interaction between SN and AN inputs will reflect the relative potency of each input onto recipient neurones, which may vary greatly across the population of NTS neurones. Equally, the functional effectiveness of the pattern of convergence will be dependent on the efferent projection of these recipient neurones.

There is anatomical and neurophysiological evidence that the intermediate portion of the NTS contains afferent terminals of both the SN and AN (Sect. 2.1.2), particularly in the case of small myelinated and unmyelinated fibres (see *Jordan* and *Spyer* 1977a, 1978b, among others). It is interesting to note that unmyelinated cardiac vagal afferents appear to excite neurones in this area also (*Fussey* et al. 1973b; *Donoghue* 1978). Since stimulation of these fibres evokes a pattern of response qualitatively the same as baroreceptor stimulation (*Öberg* and *Thoren* 1973a, b; *Little* et al. 1975), some form of convergence may be indicated at this level. In addition the activation of myelinated cardiac afferents has been shown to activate neurones in the NTS (*Baertschi* et al. 1975; *Keith* et al. 1975; *Ward* et al. 1977).

## 4.2 Cardiac Component of the Baroreceptor Reflex

### 4.2.1 Facilitation

In reviewing only the interactions of baroreceptor inputs it appears that there are considerable difference in the integration of the vascular and cardiac components of the response. In the previous section, evidence was cited which showed summation, and even implied facilitation, in the cardiac arm of the reflex. Since baroreceptor control of the heart is exerted primarily by its vagal innervation (*Wang* and *Borison* 1947a, b; *Heymans* and *Neil* 1958; *Levy* and *Zieske* 1969; *Levy* 1977), it is clear that we are concerned with mechanisms related to the control of vagal preganglionic activity. This is reinforced by the demonstration that whatever the level of ongoing sympathetic activity, vagal efferent activity is the major determinant of heart rate (*Levy* and *Zieske* 1969; *Levy* 1977).

The relationship between vagal efferent activity, as inferred from stimulation frequency, and heart rate is nonlinear (see Fig. 9), the slope being especially steep at low frequencies, and it is clear that convergent inputs may well produce enhanced effects, since vagal discharge is usually low. This may well account for the facilitation that can sometimes be observed between any two inputs that evoke a vagal bradycardia. This has been claimed for inputs from the two carotid sinuses (*Wang* and *Borison* 1947b), baroreceptors and sinus arrhythmia (*Schweitzer* 1935), chemoreceptors and nasal receptors (*Angell-James* and *Daly* 1969, 1973, 1975a, 1978; *Daly* et al. 1978, among numerous authors), the "diving" response and the baroreceptor and chemoreceptor reflexes (*Angell-James* and *Daly* 1975b; *Elsner* et al. 1977; *Angell-James* et al. 1978), superior laryngeal nerve stimulation and baroreceptor inputs (*Lopes* and *Palmer* 1976a, 1978; *Angell-James* and *Daly* 1978), and the effects of central ischaemia and medullary stimulation (*Borison* and *Domjan* 1970). There is a respiratory related control of the excitability of preganglionic vagal neurones (*McAllen* and *Spyer* 1978b; *Spyer* and *McAllen* 1979; *Spyer* 1979; see Sect. 4.3), which may contribute to the low firing of these neurones and hence may be partly responsible for the pattern of interaction demonstrated above.

Fig. 9. Change in heart rate as a function of the frequency of vagal and sympathetic stimulation frequency in Hz. The response surface represents the mean data from 10 anaesthetised dogs. Levy and Zieske (1969)

Aside from the facilitation of the baroreceptor-cardiac reflex by other peripheral afferent inputs (see above), it has often been noted that electrical stimulation at sites within the central nervous system also facilitates its effects. In the 1950s Gellhorn published a monograph proposing that the anterior hypothalamus facilitated the baroreceptor reflex (*Gellhorn* 1957). Direct evidence for such an interaction has since come from studies in which the anterior hypothalamus-preoptic region and the septum have been stimulated electrically in the cat (*Klevans* and *Gebber* 1970; *Gebber* 



and *Klevans* 1972; *Lopes* and *Palmer* 1978). Part of this area would appear to correspond to the anterior hypothalamic "depressor" area identified on the basis of the similarity of the response evoked to the baroreceptor response (*Hilton* and *Spyer* 1971). This has been broadly confirmed in the rabbit, where hypothalamic stimulation at sites which evoke a vagal bradycardia either summate with the response to AN stimulation or evoke facilitation (*Evans* 1977; *Gimpl* et al. 1976). Moreover such stimulation has been shown to excite presumed vagal preganglionic neurones in the DNV and interneurones excited by AN stimulation.

Gebber and Klevans (1972) describe a facilitation of the baroreceptorvagal reflex evoked in the cat from the amygdala and septum. In addition it would seem that this component of the reflex can be facilitated by stimulation in the midbrain and hippocampus (Hockman et al. 1969). Kaufman et al. (1979) argue for a role of the lateral subthalamus (zona incerta) in the vagal bradycardia evoked by AN stimulation, because stimulation in this area excites presumed cardio-inhibitory neurones in the DNV (and close to the NTS in the rabbit. Neurones in the zone incerta were also excited by AN stimulation at remarkably short latency (as short as 4 ms) and an anatomical connection between the NTS and this subthalamic area has been demonstrated (Ricardo and Koh 1978), therefore a facilitatory loop in baroreceptor/vagal control appears likely.

Such apparent facilitation of baroreceptor-vagal reflex may again reflect the respiratory effects of central stimulation. *Hilton* and *Spyer* (1971) drew attention to the inhibition of respiration evoked from the hypothalamic "depressor" area, and even baroreceptor stimulation alone is known to powerfully suppress inspiration (see Sect. 2.2.3.3). The anterior hypothalamus can evoke a pattern of response very similar to the baroreceptor reflex and its destruction can attenuate the reflex (*Hilton* and *Spyer* 1971), and this argues powerfully for its involvement in the integration of the baroreceptor reflex. That its stimulation may facilitate the cardiac component of the reflex (*Klevans* and *Gebber* 1970; *Gebber* and *Klevans* 1972; *Lopes* and *Palmer* 1978) or summate with it (*Jordan* et al., unpublished work) provides additional support for this contention.

### 4.2.2 Inhibition

Just as there are well-documented instances where the cardiac component of the baroreceptor reflex can be facilitated by peripheral or by central input, so there is plentiful evidence for its suppression by other inputs. This inhibitory effect could result either from influences on the excitability of the preganglionic vagal neurone pool, i.e. disfacilitation, or from a simple summation of antagonistic drives at their membranes. Alternatively, the baroreceptor reflex input might itself be gated, a possibility that has been tested experimentally (see Sects. 4.3 and 4.4). Equally, these mechanisms may not be mutually exclusive; the apparent suppression of the cardiac arm of the reflex could result from a mixture of disfacilitation, summation, and a true reflex "gating". In this context, lung inflation, with resulting activation of lung stretch afferents, may provide a valuable indication of the mechanisms where by the baroreceptor-cardiac reflex is modified. Lung inflation has been shown to suppress baroreceptor control of heart rate (*Haymet* and *McCloskey* 1975; *Davidson* et al. 1976; *Angell-James* and *Daly* 1978; *Gandevia* et al. 1978) and vagal efferent fibre discharge (*Davidson* et al. 1976), as well as affecting resting heart rate. Since it is more appropriate to consider the influence of respiration, both the central patterning and afferent feedback together, discussion will be considered later (Sect. 4.3).

However, there are plentiful examples of other peripheral inputs, such as sciatic nerve stimulation (*Quest* and *Gebber* 1972; *Kumada* et al. 1975), evoking an increase in arterial blood pressure and a simultaneous suppresson of the cardiac component of the baroreceptor reflex. This effect, which may simply represent an alerting stimulus, would be expected to excite those areas of brainstem involved in the integration of the defence reaction (Sect. 4.4). In very much the same way a powerful stimulation of the baroreceptor reflex through eliciting a defence response (*Marshall* 1977).

In addition to peripheral inputs evoking a suppressive effect on the cardiac arm of the reflex, stimulation at certain sites in the brainstem, amygdala, and hypothalamus also exert a significant inhibitory control of vagally evoked bradycardia.

It is well illustrated that stimulation at many sites in the hypothalamus, particularly in medial and posterior regions, can suppress the vagal arm of the baroreceptor reflex (Hilton 1963; Feigl et al. 1964; Folkow et al. 1964, 1968; Djojosugito et al. 1970; Gebber and Snyder 1970; Kylstra and Lisander 1970; Bagshaw et al. 1971; Humphreys et al. 1971; Wilson et al. 1971; Coote and Perez-Gonzalez 1972; Thomas and Calaresu 1974b; Kumada et al. 1975; Lopes and Palmer 1975; Keith et al. 1976; McAllen 1976; Jordan and Spyer 1977b, 1979; Coote 1978; Coote et al. 1979). while the vascular arm of the reflex either remains unaffected (*Feigl* et al. 1964; Folkow et al. 1964, 1968; Diojosugito et al. 1970; Gebber and Snyder 1970; Kylstra and Lisander 1970; Bagshaw et al. 1971; Humphreys et al. 1971; Wilson et al. 1971; Kidd and Penna 1976) or is even potentiated (Kumada et al. 1975). The vascular effects are usually considered to represent an algebraic stimulation of the baroreceptor response and the centrally evoked sympatho-excitatory response, although stimulation in the defence area may well block both cardiac and vascular components of the baroreceptor reflex (see Sect. 4.4.1). The common features of the centrally evoked response are a rise in arterial pressure, sometimes an increase in heart rate and usually an increase in respiration (see above references). Similar effects are also observed during stimulation in the midbrain (*Kumada* and *Sagawa* 1974) and amygdala (*Schlör* and *Stock* 1978; *Timms* 1977) and are accompanied by a suppression of the vagal arm of the baroreceptor reflex. Stimulation within the medulla may also evoke blocking of the baroreceptor reflex (*Perez-Gonzales* and *Rojas* 1976). In the rabbit, stimulation within the posterior hypothalamus usually suppresses the vagal bradycardia accompanying baroreceptor or AN stimulation (*Brickman* et al. 1977; *Gimpl* et al. 1976), although hypothalamic stimulation itself may not increase either heart rate or blood pressure.

In addition to these effects elicited from the brainstem it is clear that cerebellar stimulation can affect the baroreceptor reflex. As long ago as 1938, Moruzzi suggested that electrical stimulation in the cortex of the anterior lobe of the cerebellum could reduce the respiratory and cardiovascular response to both vagal stimulation and bilateral carotid occlusion (Moruzzi 1938, 1940, 1947, 1950). Although such stimulation might affect respiration, it appeared to have no effect on resting arterial blood pressure (Moruzzi 1940). Since the inhibitory effect of cerebellar stimulation was manifest on the effects of chemoreceptor excitation, it may be that the effect on the response observed during bilateral carotid occlusion represents an effect on this reflex rather than on the baroreceptor reflex. Indeed it could represent a secondary influence evoked through its inhibitory influence on respiration (see Sect. 4.3). Even allowing for these problems of interpretation these studies represented the first controlled investigation of cerebellar influences on the cardiovascular system. Since then interest has focussed more onto the role of the fastigial nucleus.

Zanchetti and Zoccolini (1954) described that stimulation in the fastigial nucleus evoked both a pressor response and bursts of sham rage in the thalamic cat. Subsequently several reports describe a characteristic pressor response elicited from the rostral pole of the fastigial nucleus in anaesthetised cats (Achari and Downman 1969, 1970; Miura and Reis 1969b, 1970, 1972a; Lisander and Martner 1971a, b, 1973). There is also an accompanying suppression of the cardiac component of the baroreceptor reflex, whether evoked through the injection of noradrenaline (Achari et al. 1973) or by vagal or sinus nerve stimulation (Hockman et al. 1970; Gurevitch and Vyshatira 1973) or distension of an isolated carotid sinus (Lisander and Martner 1971b).

There may be important functional implications in these observations: it would appear that fastigial stimulation evokes bursts of sham rage, which is equivalent to a defence reaction (*Zanchetti* and *Zoccolini* 1954; *Moruzzi* 1947), while stimulation in the white matter of the anterior lobe suppressed the autonomic components of the defence reaction (Lisander and Martner 1971a). In view of the profound effects of the defence reaction on the baroreceptor reflex, which will be reviewed later (Sect. 4.4) this implies that either the anterior lobe evokes a generalised inhibitory action on reflexes or else that its inhibitory action is concerned with chemoreceptor reflexes, which may themselves evoke defence reactions, having no influence of importance on the baroreceptor reflex except via its effect on the defence reaction. In essence, cerebellar influences may be considered to represent the antagonistic balance between cerebellar cortex and fastigial nucleus influences on the defence reaction. Lisander and Martner (1973) discount this possibility on the basis that fastigial stimulation might suppress the cholinergic vasodilatation in skeletal muscles evoked in the defence reaction, although vasoconstrictor effects were clearly facilitated (Lisander and Martner 1973; Achari et al. 1973). Behavioural components may similarly fail to show facilitation in conscious animals (Achari et al. 1973), so this apparent discrepancy cannot be taken to result from a suppressive effect of anaesthesia and caution is required in attributing a functional significance to any of the observations made so far.

It has been suggested that the anatomical and physiological connections described between the fastigial nucleus and the paramedian reticular nucleus (see *Jansen* and *Brodal* 1954; *Spyer* and *Wolstencroft* 1971; *Ghelar-ducci* et al. 1974, among many others) may mediate both the cardiovascular responses and this apparent suppression of vagally mediated bradycardia of the baroreceptor reflex (*Miura* and *Reis* 1972a). This may well prove insignificant: the role of this medullary nucleus in mediating the baroreceptor reflex has been shown to be highly questionable, its stimulation usually evoking a suppression of sympathetic activity (Sect. 3.3.3), and the interactions at the neural level appear confusing (*Miura* and *Reis* 1972a).

There is also a report that stimulating in the motor area of the cerebral cortex evokes increases in heart rate and blood pressure as well as a suppression of the bradycardia of the baroreceptor reflex (Achari and Downman 1978). Another study, however, indicates that stimulation in the motor cortex exerts little, if any, direct effect on the cardiovascular system, the changes being secondary to evoked movement (Hilton et al. 1975, 1979). Since, either directly or through evoked movement, respiratory changes may accompany stimulation in the motor cortex, the observed change in the efficacy of the reflex may again manifest only its powerful control by respiratory activity.

The most important observation may prove to be the fact that stimulating in the various brainstem sites, cerebral, cortical and cerebellar loci that profoundly modify or suppress the baroreceptor-cardiac reflex, invariably also evokes an increase in ventilatory drive. It would seem advisable at this juncture to consider the respiratory influences on the baroreceptor reflex, for such consideration may provide at least a partial answer to the nature of the synaptic mechanisms responsible for these apparent suppressive influences and the facilitatory interactions discussed previously.

## 4.3 Respiratory Influences on the Baroreceptor Vagal Reflex

In considering the ability of peripheral and central inputs to suppress or facilitate the cardiac arm of the baroreceptor reflex, indications have been obtained that central respiratory drive might be responsible in part for this apparent gating of the reflex. Lung inflation was also described as a potent stimulus to the suppression of vagal efferent discharge. Indeed, the influence of respiration on the heart, and by implication vagal efferent activity, has been appreciated since 1936, when Anrep et al. (1936a, b) showed that the respiratory related fluctuations in heart rate were effected through two mechanisms. The first had a central origin and was related to the genesis of respiratory activity (Anrep et al. 1936b); the second was the result of an inhibitory input related to lung inflation (Anrep et al. 1936a). These two mechanisms are normally in phase and the result is sinus arrhythmia (see also Katona and Jih 1975). Accordingly, if the excitability of CVMs is the key to their ability to respond to excitatory inputs, a respiratory "gate" at their membrane might explain all the apparent inhibitory influences on the baroreceptor cardiac reflex.

This argument for an effective respiratory gating of the baroreceptor reflex was first made by Koepchen and his colleagues (*Koepchen* et al.



Fig. 10. A scheme of the mechanisms responsible for the respiratory modulation of the sinus baroreceptor control of the heart (for explanation see text). (Taken from Koepchen et al. 1961b)

1961a, b). In their studies of the SN and baroreceptor control of heartperiod, they demonstrated that a stimulus to the SN was effective in prolonging heart-period only if timed to occur within expiration and was ineffective, or considerably less effective, if timed to occur during inspiration. Accordingly, three explanations were tendered to account for the observation (see Fig. 10): it could result from a "gating" of the afferent input to the vagal neurones (Fig. 10a), a respiratory related control of the excitability of CVMs (Fig. 10b), or a combination of both (Fig. 10c). These pioneering observations on heart rate (Anrep et al. 1936a, b) and baroreceptor control of heart rate (Koepchen et al. 1961a, b) have been confirmed by numerous groups. It is now generally accepted that a brief stimulus to the carotid sinus baroreceptors evokes a bradycardia only if timed to occur during expiration, an equivalent stimulus in inspiration being ineffective (Iriuchijima and Kumada 1963; Katona et al. 1970; Haymet and McCloskey 1975; Neil and Palmer 1975; Davidson et al. 1976; Lopes and Palmer 1976; McAllen and Spyer 1975). Consequently the same is true for the baroreceptor input to vagal efferent neurones (Iriuchijima and Kumada 1963; Katona et al. 1970; Davidson et al. 1976; McAllen and Spyer 1978b; see Fig. 11). Furthermore a chemoreceptor stimulus will only evoke a bradycardia and activate vagal efferent fibres if timed to occur during expiration (Davidson et al. 1976).

Considering then the possible explanations for this apparent "gating" of the reflex, we can eliminate a presynaptic control of the central terminals of the baroreceptor afferents within the NTS. *Jordan* and *Spyer* (1978a, b, 1979) have shown that the excitability of SN and AN afferent



Fig. 11. Dog. Chloralose and morphine. Records of carotid sinus blood pressure, respiratory air flow and the activity of single cardiac vagal efferent nerve (CVE) are shown. A burst of firing in the cardiac efferent nerve was evoked by a baroreceptor stimuls timed so as to occur in the expiratory pause. No firing was evoked when a similar stimulus was given during inspiration. *Davidson* et al. (1976)

terminals (without distinguishing them as either baroreceptor and chemoreceptor afferents) does not alter in phase with central respiratory activity. This was demonstrated in both cats and rabbits, in artificially ventilated and paralysed preparations, with central inspiratory activity being recorded from the phrenic nerve. Antidromic potentials evoked in the SN and AN by stimulating within the NTS showed no variation in amplitude (or threshold) in phase with central respiratory drive, nor were changes observed in phase with lung inflation, although this was not tested with large lung inflations.

It remains possible that a "gate" may occur at a postsynaptic site within the NTS, as many neurones there show both respiratory and cardiac rhythm (*Stroh-Werz* et al. 1977a, b). In contrast, many neurones receiving SN and baroreceptor input have no obvious respiratory rhythm (*Lipski* et al. 1975).

If such a "gate" exists, it cannot represent an "all or none" mechanism. The evidence for this conclusion is shown in Fig. 12. In this experiment McAllen and Spyer (1978b) took advantage of the fact that by recording



Fig. 12a-c. Cardiac rhythm of a CVM and its response to stimulation of the sinus nerve. Trace from top downwards: pulse-triggered histograms of CVM activity (120 cycles, 10 ms bins) with femoral pulse wave on same time scale, histograms of response to sinus nerve stimulation (0.1 ms pulse at 2 V, 128 cycles, 5 ms bin width). a Analysed throughout the respiratory cycle; b analysed in expiration; c analysed in inspiration. Unit firing in response to 60 nA DLH. McAllen and Spyer (1978b)

from CVMs directly and altering their excitability by the iontophoresis of DLH they could induce CVMs to fire during inspiration, when they are normally silent and inexcitable by reflex inputs, as well as during expiration. By analysing the firing patterns of CVMs, a pulse rhythm of baroreceptor origin was observed in both inspiration and expiration (see Fig. 12; McAllen and Spyer 1978b). Furthermore, stimulation of the SN was effective in both inspiration and expiration in this situation of heightened excitability. The implication is that CVMs receive an excitatory baroreceptor input throughout the central respiratory cycle but that it is normally subliminal during inspiration (McAllen and Spyer 1978b; Spyer and McAllen 1979; Spyer 1979). Hence the "gate" in the baroreceptor-cardiac reflex is essentially at the level of the CVM. This does not eliminate the possible influence of other respiratory related mechanisms acting on the pathway to CVMs but it does imply that these are relatively ineffective. To accommodate the role of lung inflation inputs, McAllen and Spyer (1978c) studied open-chest artificially ventilated cats, where central respiratory activity was out of phase with the respiratory pump; it is only necessary to suggest that such inputs act via a pathway that impinges on CVMs.

The question of the synaptic mechanisms responsible for the respiratory related control of CVM activity has been raised (*Lopes* and *Palmer* 1976, 1978; *McAllen* and *Spyer* 1976, 1978b; *Spyer* and *McAllen* 1979; *Spyer* 1979) and the simplest explanation would seem to be that it is related to central inspiratory drive. *Lopes* and *Palmer* (1976, 1978), from a range of indirect observations, have suggested that the R $\beta$  inspiratory neurones of the NTS might be the ideal candidate, since they have a central inspiratory rhythm and they are excited by lung inflation (*Baumgarten* and *Kanzow* 1958, and subsequently others). In the cat, they certainly project to the NA, where CVMs are located (*Merrill* 1974).

More directly, *McAllen* and *Spyer* (1978b) and *Spyer* and *McAllen* (1979) have argued for a role of the inspiratory neurones of the NA, among which CVMs are located (*McAllen* and *Spyer* 1976, 1978b). The pattern of discharge of CVMs firing either spontaneously or in response to the iontophoresis of DLH does not resemble the recruiting discharge of central expiratory activity (see *Merrill* 1974; *Sears* 1964), and the firing pattern of CVMs could be best explained by an inhibitory sculpturing of their activity through an inspiratory related inhibitory input (*Spyer* 1979). Indeed, more positive evidence in favour of such a mechanism has arisen from the demonstration of *Garcia* et al (1978) that atropine applied iontophoretically blocks both the inhibitory action of acetylcholine exerted on CVMs by iontophoretic application and the inspiratory related depression of activity of CVMs firing in response to DLH. *Katona* et al. (1977) had previously reported that small doses of atropine, considerably smaller than

those normally used to block vagal efferent effects on the heart, evoked both an increase in vagal efferent activity and a fall in heart rate, but only if the animal had central respiratory activity. This postulated synaptic mechanism has the added attraction that it conveniently explains both the apparent "gating" of the baroreceptor reflex and the phenomenon of sinus arrhythmia. It also leaves the original contention that the major excitatory input to CVMs arises from the arterial baroreceptors, its effectiveness being adjusted by the excitability of the CVM. Hence, any input that promotes inspiratory drive will reduce the excitability of CVMs, and any mechanism which reduces inspiratory drive will increase vagal efferent excitability. This observation does not, however, exclude the participation of other inhibitory controls that are independent of the "inspiratory gate".

On the basis of these observations alone it may be premature to discuss the synaptic mechanisms responsible for the inhibitory control of CVM activity. It could involve a direct cholinergic innervation from inspiratory neurones or be mediated via a neighbouring inhibitory interneurone which also receives an excitatory input from CVM axon collaterals, i.e. in this circumstance ACh would excite the interneurone, which would also have an inspiratory rhythm.

It would be wrong to leave the impression that only the vagal arm of the baroreceptor reflex is influenced by the respiratory cycle. The excitability of sympathetic preganglionic neurones is altered in phase with respiratory activity (Sect. 3.3.1; Lipski et al. 1977). Accordingly it is not surprising that the baroreceptor control of the heart mediated by sympathetic efferents is powerfully affected by the respiratory cycle (Seller et al. 1968; Richter et al. 1970; Seller and Richter 1971; Davis et al. 1977). The changes in heart rate evoked by the baroreceptors in vagotomised dogs are reduced during inspiration, and the effecitveness of the baroreceptor control of sympathetic activity shows quantitative differences during the respiratory cycle (Seller et al. 1968; Seller and Richter 1971; Davis et al. 1977). The duration of the silencing of sympathetic activity evoked by the baroreceptors was shown to be minimal in the middle of the phrenic nerve discharge and maximal shortly after the end of inspiration. Since there is plentiful support for an inspiratory related excitatory input to sympathetic preganglionic neurones, it is possible that the disfacilitation of these neurones at the end of inspiration is sufficient to explain their heightened sensitivity to the baroreceptor inhibitory input at that time. In effect there is no respiratory "gate" in the baroreceptor-sympathetic reflex, merely a changing sensitivity to its inhibitory action and, accordingly, a quantitative reduction in effectiveness. Thus the control of the activity of sympathetic neurones falls into the pattern described by *Pitts* et al. (1941), where summation was considered a suitable explanation of their control.

# 4.4 Total Central Inhibition of the Baroreceptor Reflex

In the previous section, the baroreceptor cardiac reflex was seen to be profoundly modified by both peripheral and central drives. In particular the portion of the reflex mediated by vagal efferent neurones was shown to be particularly sensitive to respiratory related factors, a mechanism that was seen as part of direct control of CVM excitability by respiration. In contrast, the baroreceptor sympathetic reflex showed quantitative variations during the respiratory cycle but there was no evidence of an "all-or-none" gating of this arm of the reflex. The quantitative variations in efficacy of baroreceptor control of sympathetic activity with respect to the respiratory cycle seemed to be explicable on the basis of an algebraic summation of baroreceptor inhibitory inputs and on-going excitatory inputs not dissimilar from the interactions between baroreceptor inputs and the excitatory drive from the hypothalamus first described by *Pitts* et al. (1941) and *Pitts* and *Bronk* (1942).

It is now generally accepted that the cardiac component of the baroreceptor reflex is particularly sensitive to other inputs, but there are two centrally evoked patterns of response that appear to include a total suppression of the baroreceptor reflex. This central inhibition is considered an integral part of the defence reaction (*Hilton* 1963, 1965, 1966) and can also be evoked on stimulating in a restricted portion of the inferior olive (*Smith* and *Nathan* 1966).

# 4.4.1 The Defence Reaction

It is beyond the scope of this review to describe in detail the physiological and anatomical arrangement of limbic and hypothalamic integration of defensive or aggressive behaviour, but it is important to underline that the autonomic component of such responses involves a characterisitc pattern of cardiovascular response easily distinguished from more general pressor responses evoked by central stimulation. This response, which may be elicited on electrical stimulation of the hypothalamus of anaesthetised animals, involves an increase of mean and pulse pressure, tachycardia, and a generalised peripheral vasoconstriction, although the blood flow through skeletal muscle increases dramatically (for refs. see Hilton 1966). In the cat, this hindlimb vasodilatation involves both an activation of sympathetic cholinergic vasodilator fibres and a reduction in vasoconstrictor tone (Eliasson et al. 1951; Abrahams et al. 1960). The concomitant increase in heart rate and arterial pressure suggests a modification, or at least a resetting, of the baroreceptor reflex. Indeed, Hilton (1963) claimed that the reflex was totally suppressed during the hypothalamically induced defence response. This preliminary report triggered off considerable controversy and a spate of reports claiming that although the cardiac component of the baroreceptor reflex was suppressed during the defence reaction, the vascular component was either unaffected (*Feigl* et al. 1964; *Folkow* et al. 1964, 1968; *Djojosugito* et al. 1970; *Kylstra* and *Lisander* 1970; *Bagshaw* et al. 1971; *Humphreys* et al. 1971; *Wilson* et al. 1971; *Kidd* and *Penna* 1976) or even potentiated (*Kumada* et al. 1975) and that the two inputs might act synergistically in producing the characteristic hindlimb vasodilatation of the defence reaction (*Djojosugito* et al. 1970). Behaviourally the effects of the two drives are opposite (see *Hilton* 1975) and may even be mutually inhibitory, so a synergistic relationship between their effects on the cardiovascular system would seem surprising.

The case for a synergistic interrelationship rested on the observations of Folkow et al. (1964) and Bolme et al. (1967) that vasoconstrictor tone has to be more or less totally eliminated before cholinergic vasodilatation can be fully developed. They argued that the powerful vasoconstrictor drive to other vascular areas, with an accompanying increase in cardiac output and hence blood pressure, led to a baroreceptor-mediated suppression of the vasoconstrictor supply to skeletal muscle (Djojosugito et al. 1970). This extrapolation has since been shown to be unnecessary. Coote et al. (1973) have shown in the cat that stimulating in part of the descending pathway for the defence reaction, in a narrow strip close to the dorsal surface of the medulla, evokes an increased hindlimb flow that is independent of both cholinergic vasodilator fibres and baroreceptor inputs but is abolished by guanethidine; hence it is likely to result from an active withdrawal of vasoconstrictor tone specifically in skeletal musculature. Moreover, Horevseck et al. (1976) have shown that stimulation in areas of the hypothalamus which evoke a cholinergic vasodilatation in hindlimb skeletal muscles, evokes a discharge in normally silent postganglionic neurones (putative cholinergic sympathetic neurones) and a complex pattern of response in postganglionic vasoconstrictor neurones. These latter neurones may be silenced briefly, followed by a burst of activity which precedes a second and prolonged suppression, at a time when cholinergic vasodilator fibres are most conspicuously active.

The argument of Folkow et al. (1964) is further handicapped by the fact that their observations would demand a differential effect on vasoconstrictor activity of different vascular beds. It is now well documented that renal sympathetic activity is powerfully excited during defence are stimulation (McAllen 1976; Jordan and Spyer 1977b, 1979; Coote et al. 1979) and that this remains elevated throughout stimulation. Stimulating at sites beyond this integrative area similarly elevates sympathetic activity but the response is not maintained. Furthermore, baroreceptor inputs are ineffective in lowering or silencing sympathetic activity during such defence area stimulation (McAllen 1976; Coote et al. 1979). Since the effects of SN stimulation are also blocked by such stimulation (*Jordan* and *Spyer* 1977, 1979; *Coote* et al. 1979), it is clear that the mechanism involves a central suppression and cannot be explained by a sympathetic mediated resetting of the baroreceptors themselves, as remained a possibility from the observations of the effect of sympathetic activity on baroreceptors (*Koizumi* and *Sato* 1969).

It would thus appear that the central mechanism underlying the interaction between baroreceptors and the defence reaction is more than a simple saturation effect due to the enormous excitatory drive to the sympathetic neuronal pool evoked on hypothalamic stimulation. Increases of sympathetic activity of comparable magnitude evoked from other sites in the hypothalamus, for instance, remain buffered by the baroreceptors (*Coote* et al. 1979).

There has been much speculation on the nature of the mechanisms responsible for this complete suppression of the reflex. The receptors themselves do not appear to be significantly affected, and it has been proposed that the afferent terminals in the medulla might be under a presynaptic control (Weiss and Crill 1969). They claimed that stimulation within the fields of Forel, an area of the hypothalamus which is not a part of the integrative matrix of the defence reaction, evoked a depolarisation of SN afferent endings in the NTS. Jordan and Spyer (1977, 1979) have looked directly at the effect of conditioning stimuli delivered within the hypothalamic defence area on SN and glossopharyngeal afferent terminals in the NTS. They could show no effect on SN terminals, although glossopharyngeal afferents were markedly affected by stimulation at hypothalamic sites which totally suppressed the cardiovascular responses to SN stimulation. There are, however, indications that the excitatory responses of NTS neurones to both baroreceptor and SN stimulation may be blocked by conditioning stimuli delivered to the hypothalamic defence area (McAllen 1976). On the basis of the latency of the response of these neurones to SN stimulation it would seem that they were located within one or two synapses of the afferent input (McAllen 1976). The specificity of this action is in question: Adair and Manning (1975) claim to have blocked the normal excitatory response of NTS neurones to SN stimulation on stimulating in the hypothalamus, although the hypothalamic site of stimulation and its effect on the baroreceptor reflex were never analysed. These observations indicate that at least part of the block of the baroreceptor reflex during the defence reaction may be effected close to the termination of baroreceptor afferents. In this type of interaction we are concerned with the functioning of polysynaptic pathways, and it is likely the synaptic processes mediating these changes may involve actions at several levels of the neuraxis, including the hypothalamus (Gellhorn 1957; Hilton and Spyer 1971; Hilton 1975).

The block of the cardiac component of the reflex during the defence reaction may be elicited through the respiratory "gate" (Sect. 4.3). Defence area stimulation certainly evokes hyperphoea, with an especially powerful inspiration drive. However, Lopes and Palmer (1978) have suggested that hypothalamic inputs might exert an additional tonic inhibitory control of CVM activity. Jordan, Khalid, Schneiderman and Spyer (unpublished work) have since shown that stimulation in the defence area of the hypothalamus of the cat inhibits both on-going CVM activity (i.e. that induced by the iontophoresis of DLH) and AN inputs to CVMs and that this inhibitory action is not blocked by the iontophoresis of atropine although this did block inspiratory related inhibition. Accordingly, hypothalamic descending inhibition is likely to be mediated, at least partly, by a mechanism independent of the inspiratory-gate. Furthermore, it appears that at least a portion of the CVM neuronal pool is amenable to recurrent inhibition (Jordan et al., unpublished work), an influence that is antagonised by iontophoresis of atropine. Although at a preliminary stage, the observations together provide an indication of the nature of the inhibitory mechanisms that control vagal efferent discharge (see Fig. 13).

The defence area influence on the sympathetic and vascular components of the baroreceptor reflex is not necessarily an all-or-none phenome-



Fig. 13. Diagram illustrating the control of the baroreceptor input to cardiac vagal motoneurone (CVM). Inspiratory neurones of the NA (I) exert an inhibitory control of CVM either directly, or via an interneurone which may also receive recurrent collateral inputs from CVMs. This inhibitory mechanism is sensitive to atropine. The hypothalamic defence area may inhibit CVM activity and block their baroreceptor input through this mechanism, but also by an alternative mechanism. This may involve a direct inhibitory control of CVM, or via a modification of transmission through the NTS. See text for further details. Dotted lines represent pathways of unknown synaptic complexity, excitatory pathways are shown by thick lines, inhibitory by thin lines

non. The temporal relationship between these antagonistic stimuli is important. Stock (pers. communication) has shown that if a baroreceptor stimulus is timed early in a defence response elicited by amygdala stimulation, it is totally blocked, but delaying the test stimulus until later during the defence response allows the vascular component to reassert itself although the cardiac component remains suppressed. The fatigue of cholinergic sympathetic dilatation has been documented (*Djojosugito* et al. 1968), and the slowly increasing vasoconstrictor tone in the supply to skeletal muscle may well provide a basis on which to explain this observation (*Horeyseck* et al. 1976). Also, many of the interactions may be reserved for the level of the preganglionic sympathetic neurones, so the temporal and spatial nature of descending inputs may play an important role in the pattern of interaction observed.

# 4.4.2 The Inferior Olive

It has been suggested that a region in the dorsomedial portion of the inferior olive exerts an inhibitory control of the baroreceptor reflex (Smith and Nathan 1966). Stimulation of this region evokes no obvious change in arterial blood pressure or heart rate but completely blocks the effect of carotid sinus inflation. Smith and Nathan (1966) suggested that this area of the medulla might mediate the defence area inhibition of the baroreceptor, and more recent neurohistological studies have indicated a descending pathway from the perifornical region of the hypothalamus to this part of the olivary complex (Saper et al. 1976). Smith and Nathan (1966) implied that the inhibitory control might be mediated via the cerebellum, since Moruzzi (1938, 1940) and subsequently others (Hoffer et al. 1966) had indicated that the anterior lobe of the cerebellum might exert an inhibitory control of autonomic reflexes in general and the baroreceptor reflex in particular. In some unpublished studies, *Coote* et al. have confirmed the original observation of Smith and Nathan (1966) but have shown that the inhibitory effect on the baroreceptor reflex survives cerebellectomy. They also have evidence that bilateral lesions in the olive, destroying the specific region from which this effect can be elicited, in no way impair the suppressive effect of the hypothalamic defence area on the baroreceptor reflex. Interestingly, these restricted lesions alone produce an enormous increase in both mean arterial blood pressure and pulse pressure, which developes slowly and is maintained. Furthermore, against a background of high sympathetic tone produced by bilateral carotid occlusion, stimulation in the olive may evoke a small fall in arterial pressure. It seems remarkable that despite blocking the baroreceptor reflex at normal levels of background blood pressure, olivary stimulation is ineffective in evoking obvious cardiovascular responses and so is apparently leaves ongoing baroreceptor

inputs unaffected. At this stage, the pattern of effect can best be described as interesting: further studies will be required before the significance of the olive in cardiovascular control can be assessed.

## 5 Concluding Remarks

This review has endeavoured to consider the baroreceptor reflex as a mechanism with a distinctive input-output relationship whose central pathway might shed light on basic problems concerning the nervous control of the heart and circulation. As such, it has dealt mainly with establishing the central connections mediating the baroreceptor control of sympathetic and vagal preganglionic neurones, but the survey has been extended to discern the synaptic processes responsible for the modification of the pattern of reflex response that occurs in different physiological states. This latter, relatively restricted discussion may in fact hold the most important clues for understanding the integrative basis of the baroreceptor reflex and consequently the nervous control of arterial blood pressure.

These rather broad generalisations can be supported by some significant conclusions that arise from such a survey of the literature. For example, there is now abundant evidence in favour of an essential role of the NTS in the mediation of the baroreceptor reflex and in cardiovascular and respiratory control. It remains to resolve whether this involves simply an ordered organisation of the diverse afferent inputs which impinge on this nucleus or whether the output from this nucleus in response to, say, baroreceptor stimulation can be modified by other afferent inputs to this area, both those arising from peripheral receptors and from central structures. Such modification would imply an integrative function and could be expected to result in differential effects on the heart and vasculature under different physiological situations. There are techniques currently available to solve this problem which are analogous to those neurophysiological and neuroanatomical approaches being used so successfully in studying the processing of afferent information in the dorsal horn of the spinal cord.

A start has certainly been made in this direction with the recent reappraisals of the anatomical description of this area and its efferent connections, to which has been added neurophysiological data on the inputs to the NTS. In this context probably more is known of the baroreceptor innervation of the NTS than for any other of its afferent inputs, yet its interactions at this level with other afferents have, as yet, barely been approached.

From this first relay in the NTS, there are indications of a complex series of connections that mediate the baroreceptor reflex and extend through divergent areas of the brainstem. This effectively diffuse pathway finally converges to control the activity of the autonomic preganglionic neurones, the sympathetic and vagal neurones. A major advance has been achieved in the last decade with the realisation that these neurones have an integrative function. In this, their baroreceptor input over a polysynaptic pathway forms just one of the many inputs that modify their excitability, but such an organisation can certainly be seen to minimise the role of any distinctive and restricted "vasomotor centre" in the control of the circulation. The many links in the baroreceptor pathway may still remain uncertain, but we have reached a stage where analytical rather than simply qualitative studies can be undertaken to identify the action of the baroreceptors and their interactions with other inputs on the activity of these neurones.

The divergent nature of the baroreceptor input may well have contributed to our failure to identify more precisely the interneuronal matrix of the reflex. In fact connections with little direct relevance to the basic reflex may well have obscured its most important connections, since so many of its influences appear to involve the reticular formation. As a reasonable working hypothesis, it is possible to suggest that the baroreceptor input may have a preferential pathway through the reticular formation related to cardiovascular control and an indirect influence on both this and behaviour through more general effects on the reticular formation. The "preferential" system would seem to offer the most immediate challenge to neurophysiological investigation; certainly a series of possible reticulospinal connections have been revealed which may contribute to the baroreceptor control of sympathetic efferent activity. The latter and more generalised effects may well also alter the excitability, and hence effectiveness, of the preferential pathway, but except in a few instances little has yet been achieved in investigating the more general effects of the baroreceptor input on the reticular formation.

An involvement of the baroreceptors in the regulation of reticular mechanisms, such as sleep-wakefulness, has been described. The marked cardiovascular changes in these states certainly imply interactions, but whether these are at the level of the brainstem, at the final common pathway, the preganglionic neurone, or at both remains to be resolved.

An attempt has been made to assess how certain patterns of physiological behaviour evoked from the central nervous system can influence the effectiveness of baroreceptor inputs. Indeed, studies on the influence of central respiratory activity and the defence reaction on the efficacy of the baroreceptor control of vagal and sympathetic activity have shed considerable light on the basic mechanisms underlying the reflex. Furthermore, since these two patterns of activity are so powerfully affected by peripheral inputs and behavioural state, they may well offer particularly valuable clues to the basic mechanisms underlying not only the integration of the baroreceptor reflex, but also the nervous control of circulation.

Acknowledgements. It is a pleasure to acknowledge the contributions made, often unwittingly, by several colleagues, (Prof. S.M. Hilton, Dr. D. Jordan, Dr. R.M. McAllen and Dr. D.W. Richter) to the author's understanding of the problems outlined in this review. The provision of research facilities by Prof. Hilton, financed through grants from the Medical Research Council, is gratefully acknowledged. Dr. D. Jordan has read and criticised the complete manuscript, and Dr. N. Schneiderman a considerable portion; their help was invaluable. Special thanks are due to Sandra Field for typing the manuscript and to Rita Hore for producing the illustrations.

## References

- Abrahams VC, Hilton SM, Zbrozyna A (1960) Active muscle vasodilatation. Produced by stimulation of the brainstem: its significance in the defence reaction. J Physiol (Lond) 154:491-513
- Achari NK, Downman CBB (1969) Autonomic responses evoked by stimulation of fastigial nuclei in the anaesthetized cat. J Physiol (Lond) 204:130
- Achari NK, Downman CBB (1970) Autonomic effector responses to stimulation of nucleus fastigius. J Physiol (Lond) 210:637-650
- Achari NK, Downman CBB (1978) Inhibition of reflex bradycardia by stimulation of cerebral motor cortex. Brain Res 150:198-200
- Achari NK, Al-Ubaidy SS, Downman CBB (1973) Cardiovascular responses elicited by fastigial and hypothalamic stimulation in conscious cats. Brain Res 60:439-447
- Achari NK, Al-Ubaidy SS, Downman CBB (1978) Spinal sympathoexcitatory pathways activated by stimulating fastigial nuclei, hypothalamus and lower brain stem in cats. Exp Neurol 62:230–240
- Adair JR, Manning JW (1975) Hypothalamic modulation of baroreceptor afferent unit activity. Am J Physiol 229:1357–1364
- Adrian ED, Bronk DW, Phillips G (1932) Discharges in mammalian sympathetic nerves. J Physiol (Lond) 74:115-133
- Agostoni E, Chinnock JE, Daly M de B, Murray JG (1957) Functional and histological studies of the vagus nerve and its branches to the heart, lungs and abdominal viscera in the cat. J Physiol (Lond) 135:182-205
- Akimoto H, Saito Y (1966) Synchronizing and desynchronizing influences and their interactions on cortical and thalamic neurones. Prog Brain Res 21A:323-351
- Alanis J, Mascher D, Miyamoto J (1966) Hypothalamic stimulation and the inhibition of the activity of cardiac sympathetic and parasympathetic nerves. Arch Int Physiol Biochim 74:766-784
- Alexander RS (1945) The effects of blood flow and anoxia on spinal cardiovascular centers. Am J Physiol 143:698-708
- Alexander RS (1946) Tonic and reflex functions of medullary sympathetic centers. J Neurophysiol 9:205-217
- Amendt K, Czachurski J, Dembowsky K, Seller H (1978) Localisation of brainstem neurons projecting to the intermediolateral nucleus in the cat. Pfluegers Arch 373: 76
- Anderson FD, Berry OM (1956) An oscillographic study of the central pathways of the vagus nerve in the cat. J Com Neurol 106:163-181

- Angell-James JE (1971a) The effects of altering mean pressure, pulse pressure and pulse-frequency on the impulse activity in baroreceptor fibres from the aortic arch and right subclavian artery in the rabbit. J Physiol (Lond) 214:65-88
- Angell-James JE (1971b) The responses of aortic arch and right subclavian baroreceptors to changes in non-pulsatile pressure and their modification by hypothermia. J Physiol (Lond) 214:201-223
- Angell-James JE, Daly M de B (1969) Cardiovascular responses in apnoeic asphyxia; role of arterial chemoreceptors and the modification of their effects by a pulmonary inflation reflex. J Physiol (Lond) 201:87-104
- Angell-James JE, Daly M de B (1970) Comparison of the reflex vasomotor responses to separate and combined stimulation of the carotid sinus and aortic arch baroreceptors by pulsatile and non-pulsatile pressures in the dog. J Physiol (Lond) 209: 257-293
- Angell-James JE, Daly M de B (1973) The interaction of reflexes elicited by stimulation of carotid body chemoreceptors and receptors, in the nasal mucosa affecting respiration and pulse interval in the dog. J Physiol (Lond) 229:113-149
- Angell-James JE, Daly M de B (1975a) Some aspects of upper respiratory tract reflexes. Acta Otolaryngol (Stockh) 79:242–251
- Angell-James JE, Daly M de B (1975b) Role of the arterial chemoreceptors in the control of the cardiovascular responses to breath-hold diving. In: Purves MJ (ed) The peripheral arterial chemoreceptors. Cambridge University Press, London, pp 387-405
- Angell-James JE, Daly M de B (1978) The effects of artificial lung inflation on reflexly induced bradycardia associated with apnoea in the dog. J Physiol (Lond) 274: 349-366
- Angell-James JE, Daly M de B, Elsner R (1978) Arterial baroreceptor reflexes in the seal and their modification during experimental dives. Am J Physiol 234:730-739
- Anrep GV, Pascual W, Rössler R (1936a) Respiratory variations of heart rate. I. The reflex mechanism of the respiratory arrhythmia. Proc R Soc Lond (Biol) 119:191-217
- Anrep GV, Pascual W, Rössler R (1936b) Respiratory variations of the heart-rate. II. The central mechanism of respiratory arrhythmia and the inter-relationships between central and reflex mechanisms. Proc R Soc Lond (Biol) 119:218-230
- Åström KE (1952) On the central course of afferent fibres in the trigeminal, glossopharyngeal and vagal nerves and their nuclei in the mouse. Acta Physiol Scand (Suppl 106) 29:209-320
- Bach LMN (1952) Relationships between bulbar respiratory, vasomotor and somatic facilitatory and inhibiting areas. Am J Physiol 171:417-435
- Baertschi AJ, Munzer RF, Ward DG, Johnson RN, Gann DS (1975) Right and left arterial B-fibre input to the medulla of the cat. Brain Res 98:189-193
- Bagshaw RJ, Iizuka M, Peterson LH (1971) Effect of interaction of the hypothalamus and the carotic sinus mechanoreceptors system on the renal haemodynamics in the anaesthetized dog. Circ Res 29:569-585
- Barker JL, Crayton JW, Nicoll RA (1971) Supraoptic neurosecretory cells: autonomic modulation. Science 171:206-207
- Barman SM, Gebber GL (1978a) Tonic sympathoinhibition in the baroreceptor denervated cat. Proc Soc Exp Biol Med 157:648-655
- Barman SM, Gebber GL (1978b) Site of recurrent inhibition of preganglionic sympathetic unit discharges. Fed Proc 37:744
- Barman SM, Wurster RD (1975) Visceromotor organization within descending spinal sympathetic pathways in the dog. Circ Res 37:209-214
- Barman SM, McCaffrey TV, Wurster RD (1976) Cardiovascular and electrophysiological responses to sympathetic pathway stimulation. Am J Physiol 230:1095-1100

- Bartorelli C, Bizzi E, Libretti A, Zanchetti A (1960) Inhibitory control of sinocarotid pressoceptive afferents on hypothalamic autonomic activity and sham-rage behavior. Arch Ital Biol 98:308-326
- Baumgarten R von, Aranda Coddou L (1959) Distribución de las aferencias cardiovasculares y respiratorias en las raices bulbares del nervio vago. Acta Neurol Lat Am 5:267-278
- Baumgarten R von, Kanzow E (1958) The interaction of two types of inspiratory neurones in the region of the tractus solitarious of the cat. Arch Ital Biol 96:361-373
- Baust W, Heinemann H (1967) The role of the baroreceptors and blood pressure in the regulation of sleep and wakefulness. Exp Brain Res 3:12-24
- Baust W, Niemczyk H (1968) The influence of mesencephalic structures on phasic regulation of blood pressure. Pfluegers Arch 301:31-42
- Baust W, Niemczyk H, Schaeffer H, Vieth J (1963) Über ein pressosensibles Areal im hinteren Hypothalamus der Katze. Pfluegers Arch 174:374–384
- Bayliss WM (1923) The vasomotor system. Longman, London
- Beattie J, Brow GR, Long CNH (1930) Physiological and anatomical evidence for the existence of nerve tracts connecting hypothalamus with spinal sympathetic centres. Proc R Soc Lond (Biol) 105:253-275
- Bennett JA, Kidd C, Lafit AB, McWilliams PN (to be published) The brainstem locations of cell bodies of vagal efferent fibres in cardiac and pulmonary branches in the cat. J Physiol (Lond)
- Berger AJ (1979) Distribution of carotid sinus nerve afferent fibers to solitary tract nuclei of the cat using transganglionic transport of horseradish peroxidase. Neurosci Lett 14:153-158
- Bianchi AL (1971) Localisation et étude des neurones respiratoires embaires. Mise en jeu antidromique par stimulation spinale ou vagale. J Physiol (Paris) 63:5-40
- Biscoe JJ, Sampson SR (1968) Rhythmical and non-rhythmical spontaneous activity recorded from the central cut end of the sinus nerve. J Physiol (Lond) 196: 327-338
- Biscoe TJ, Sampson SR (1970a) Field potentials evoked in the brain stem of the cat by stimulation of the carotid sinus, glossopharyngeal, aortic and superior laryngeal nerves. J Physiol (Lond) 209:341-358
- Biscoe TJ, Sampson SR (1970b) Responses of cells in the brainstem of the cat to stimulation of the sinus, glossopharyngeal, aortic and superior laryngeal nerves. J Physiol (Lond) 209:359-373
- Bok T (1928) Das Rückenmark. In: Mollendorf W (ed) Handbuch der mikroskopischen Anatomie des Menschen. Srpinger, Berlin
- Bolme P, Ngai SH, Rosen S (1967) Influence of vasoconstrictor nerve activity on the cholinergic vasodilator response in skeletal muscle in the dog. Acta Physiol Scand 71:323-333
- Bonvallet M, Sigg B (1958) Etude electrophysiologique des afferences vagales au niveau de leur penetration dans le bulbe. J Physiol (Paris) 50:63-74
- Bonvallet M Dell P, Hiebel G (1953) Sinus carotidien et activé électrique cérébrale. C.R. Soc Biol (Paris) 147:1166-1169
- Borison HL, Domjan D (1970) Persistence of the cardioinhibitory response to brainstem ischaemia after destruction of the area postrema and the dorsal vagal nuclei. J Physiol (Lond) 211:263-277
- Brickman AL, Kaufman MP, Petrik GK, Schneiderman N (1977) Responses of anterior hypothalamic neurons to stimulation of aortic nerve and caudate nucleus in the rabbit. Exp Neurol 56:622-627
- Brodal A (1969) Neurological anatomy in relation to clinical medicine, 2nd ed. Oxford University Press, London
- Bronk DW, Tower SS, Solandt DY (1938) The transmission of trains of impulses through sympathetic ganglion and its postganglionic nerves. Am J Physiol 122: 1-15
- Bronk DW, Pitts RF, Larrabee MG (1940) Role of the hypothalamus in cardiovascular regulation. Res Publ Assoc Res Nerv Ment Dis 20:323-341
- Brown GL, Eccles JC (1934a) The action of a single vagal volley on the rhythm of the heart beat. J Physiol (Lond) 81:211-240
- Brown GL, Eccles JC (1934b) Further experiments on vagal inhibition of the heart beat. J Physiol 81:241-257
- Burkhart SM, Ledsome JR (1977) The response to distension of the pulmonary veinleft atrial junctions in anaesthetised dogs after section of the rostral medulla. J Physiol (Lond) 273:57-68
- Burkhart SM, Furnell L, Ledsome JR (1977) Effects of medullary lesions on arterial baroreceptor reflexes and responses to distension of pulmonary vein-left atrial junctions in anaesthetized dogs. J Physiol (Lond) 273:69-82
- Cajal S, Ramon Y (1909) Histologie du systeme nerveux de l'homme et des vertebres. Maloine, Paris
- Calaresu FR, Cottle MK (1965) Origin of cardiomotor fibres in the dorsal nucleus of the vagus in the cat: A histological study. J Physiol (Lond) 176:252-260
- Calaresu FR, Henry JL (1970) The mechanism of cardio-acceleration elicited by electrical stimulation of the parahypoglossal area in the cat. J Physiol (Lond) 210: 107-120
- Calaresu FR, Pearce JW (1965a) Electrical activity of efferent vagal fibres and dorsal nucleus of the vagus during reflex bradycardia in the cat. J Physiol (Lond) 176: 228-240
- Calaresu FR, Pearce JW (1965b) Effects on heart rate of electrical stimulation of medullary vagal structures in the cat. J Physiol (Lond) 176:241-251
- Camerer H, Stroh-Werz M, Krienke B, Langhorst P (1977) Postganglionic sympathetic activity with correlation to heart-rate and central cortical rhythms. Pfluegers Arch 370:221-226
- Cannon WB (1929) Sympathetic division of the autonomic system in relation to homeostasis. Arch Neurol Psychiatry 22:282-294
- Cannon WB (1930) The autonomic nervous system: an interpretation. Lancet I:1109-1115
- Cannon WB (1932) The wisdom of the body. Norton, New York
- Cedarbaum JM, Aghajanian GK (1978) Afferent projections to the rat locus coeruleus as determined by a retrograde tracing technique. J Comp Neurol 178:1-16
- Chai CY, Wang SC (1962) Localisation of central cardiovascular control mechanism in lower brainstem of the cat. Am J Physiol 202:25-30
- Chai CY, Wang SC (1968) Integration of sympathetic cardiovascular mechanisms in medulla oblongata of the cat. Am J Physiol 215:1310-1315
- Chen HI, Cahi CY (1976) Integration of the cardiovagal mechanism in the medulla oblongata of the cat. Am J Physiol 231:454-461
- Chess GF, Tam RMK, Calaresu FR (1975) Influence of cardiac neural inputs on rhythmic variations of heart period in the cat. Am J Physiol 228:775-780
- Chiurugi E, Mollica A (1954) Contributo alla localizzazione del centro vagale cardioinhibitore. Arch Fisiol 54:249-267
- Chung JM, Chung K, Wurster RD (1975) Sympathetic preganglionic neurons of the cat spinal cord. Horseradish peroxidase study. Brain Res 91:126-131
- Ciriello J, Calaresu FR (1977) Do some cardioinhibitory axons originate in the external cuneate nucleus? Neuroscience Abstract 3:19
- Cohen DH, Schnall AM, MacDonald RL, Pitts LH (1970) Medullary cells of origin of vagal cardioinhibitory fibres in the pigeon. I. Anatomical structure of peripheral vagus nerves and the dorsal motor nucleus. J Comp Neurol 140:299-320
- Cohen, MI, Gootman PM (1969) Spontaneous and evoked oscillations in respiratory and sympathetic discharge. Brain Res 16:265-268
- Cohen MI, Gootman PM (1970) Periodicities in efferent discharge of splanchnic nerve of the cat. Am J Physiol 218:1092-1101

- Coleridge HM, Coleridge JCG, Rosenthal F (1976) Prolonged inactivation of cortical pyramidal tract neurones in cats by distension of the carotid sinus. J Physiol (Lond) 256:635-650
- Coote JH (1978) Afferent input as factors in aberrant autonomic, sensory and motor function. In: Korr IM (ed) The neurobiologic mechanisms in manipulative therapy. Plenum, New York, pp 91-127
- Coote JH, Downman CBB (1966) Central pathways of some autonomic reflex discharges. J Physiol (Lond) 183:714-729
- Coote JH, Downman CBB (1969) Supraspinal control of reflex activity in renal nerves. J Physiol (Lond) 202:161–170
- Coote JH, Macleod VH (1974a) The influence of bulbospinal monoaminergic pathways on sympathetic nerve activity. J Physiol (Lond) 241:453-457
- Coote JH, Macleod VH (1974b) Evidence for the involvement in the baroreceptor reflex of a descending inhibitory pathway. J Physiol (Lond) 241:477-496
- Coote JH, Macleod VH (1975) The spinal route of sympatho-inhibitory pathways descending from the medulla oblongata. Pfluegers Arch 359:335-347
- Coote JH, Macleod VH (1977) The effect of intraspinal microinjections of 6-hydroxydopamine on the inhibitory influence exerted on spinal sympathetic activity by the baroreceptors. Pfluegers Arch 371:271-277
- Coote JH, Perez-Gonzalez JF (1972) The baroreceptor reflex during stimulation of the hypothalamic defence region. J Physiol (Lond) 224:74-75
- Coote JH, Sato A (1978) Supraspinal regulation of spinal reflex discharge into cardiac sympathetic nerves. Brain Res 142:425-437
- Coote JH, Westbury DR (1974) The influence of the carotid sinus baroreceptors on activity in single sympathetic preganglionic neurones. J Physiol (Lond) 241:22-23
- Coote JH, Westbury DR (1979a) Functional grouping of sympathetic preganglionic neurones in the third thoracic segment of the spinal cord. Brain Res 179:367-372
- Coote JH, Westbury DR (1979b) Intracellular recordings from sympathetic preganglionic neurones. Neurosci Lett 15:171-175
- Coote JH, Downman CBB, Weber WV (1969) Reflex discharges into thoracic white rami elicited by somatic and visceral afferent excitation. J Physiol (Lond) 202: 147-155
- Coote JH, Hilton SM, Zbrozyna AW (1973) The ponto-medullary area integrating the defence reaction in the cat and its influence on muscle blood flow. J Physiol (Lond) 229:257-274
- Coote JH, Macleod VH, Martin TL (1978) Bulbospinal tryptaminergic neurones. A search for the role of bulbospinal tryptaminergic neurones in the control of sympathetic activity. Pfluegers Arch 377:109-116
- Coote JH, Hilton SM, Perez-Gonzalez JF (1979) Inhibition of the baroreceptor reflex on stimulation in the brainstem defence centre. J Physiol (Lond) 288:549-560
- Cottle MK (1964) Degeneration studies of primary afferents of IXth and Xth cranial nerves in the cat. J Comp Neurol 122:329-343
- Cottle MKW, Calaresu FR (1975) Projections from the nucleus and tractus solitarius in the cat. J Comp Neurol 161:143-158
- Crill WE, Reis DJ (1968) Distribution of carotid sinus and depressor nerves in the cat brain stem. Am J Physiol 214:269-276
- Culberson JL, Kimmel DL (1972) Central distribution of primary afferent fibres of the glossopharyngeal and vagal nerves in the opossum didelphis virginiana. Brain Res 44: 325-335
- Dahlström A, Fuxe K (1965) Evidence for the existence of monoamine neurons in the central nervous system. II. Experimentally induced changes in the intraneuronal amine levels in the bulbospinal neuron systems. Acta Physiol Scand (Suppl 247) 64:7-85
- Daly M de B, Elsner R, Angell-James JE (1977) Cardiorespiratory control by the carotid chemoreceptors during experimental dives in the seal. Am J Physiol 232: H508-516

- Daly M de B, Korner PI, Angell-James JE, Oliver JR (1978) Cardiovascular-respiratory reflex interactions between carotid bodies and upper-airways resistance in the monkey. Am J Physiol 234:H293-299
- Davidson NS, Goldner S, McCloskey DI (1976) Respiratory modulation of baroreceptor and chemoreceptor reflexes affecting heart-rate and cardiac vagal efferent nerve activity. J Physiol (Lond) 259:523-530
- Davies RO, Edwards MW (1973) Distribution of carotid body chemoreceptors afferents in the medulla of the cat. Brain Res 64:451-454
- Davies RO, Edwards MW (1975) Medullary relay neurons in the carotid body chemoreceptor pathways of cats. Respir Physiol 24:69–79
- Davis AL, McCloskey DI, Potter EK (1977) Respiratory modulation of baroreceptor and chemoreceptor reflexes affecting heart rate through the sympathetic nervous system. J Physiol 272:691-703
- Day MD, Roach AG (1974) Central adrenoreceptors and the control of arterial blood pressure. Clin Exp Pharmacol Physiol 1:347–360
- De Castro F (1928) Sur la structure et l'innervation du sinus carotidien de l'homme et des mammiferes. Nouveau faits sur l'innervation et la function du glomus caroticum. Etudes anatomiques et physiologiques. Trab Insv Cajal Invest Biol 25:331-350
- De Groat WC, Lalley PM (1974) Reflex sympathetic firing in response to electrical stimulation of the carotid sinus nerve in the cat. Brain Res 80:17-40
- De Jong W, Nijkamp P, Bohus B (1975a) Role of noradrenaline and serotonin in the central control of blood pressure in normotensive and spontaneously hypertensive rats. Arch Int Pharmacodyn Ther 213:272-284
- De Jong W, Zandberg P, Bohus B (1975b) Central inhibitory noradrenergic cardiovascular control. Brain Res 42:285–298
- De Jong W, Palkovits M (1976) Hypertension after localised transection of brainstem fibres. Life Sci 18:61-64
- De Jong W, Zandberg P, Palkovits M, Bohus B (1977) Acute and chronic hypertension after lesions and transections of the rat brainstem. Prog Brain Res 47:189-198
- Dell P, Bonvallet M (1966) Influénces d'origine cardio-vasculaire et réspiratoire sur l'activité somatique. Acta Neuroveg 28:148–168
- Dembowsky K, Czachurski J, Amendt K, Seller H (1978) Tonic, supraspinal monoaminergic inhibition on spinal somato-sympathetic reflexes. Pfluegers Arch 373:76
- De Vito JL, Clausing KW, Smith OA (1974) Uptake and transport of horseradish peroxidase by cut ends of the vagus nerve. Brain Res 82:269-271
- Djojosugito A, Folkow B, Lisander B, Sparks H (1965) Mechanism of escape of skeletal muscle resistance vessels from the influence of sympathetic cholinergic vasodilator fibre activity. Acta Physiol Scand 72:148-156
- Djojosugito AM, Folkow B, Kylstra PH, Lisander B, Tuttle RS (1970) Differentiated interaction between hypothalamic defence reaction and baroreceptor reflexes. Acta Physiol Scand 78:376-385
- Doba N, Reis DJ (1973) Acute fulminating neurogenic hypertension produced by brainstem lesions in the rat. Circ Res 32:584-593
- Doba N, Reis DJ (1974a) Role of cerebellum and the vestibular apparatus in regulation of orthostatic reflexes in cats. Circ Res 34:9–18
- Doba N, Reis DJ (1974b) Role of central and peripheral adrenergic mechanisms in neurogenic hypertension produced by brainstem lesions in the rat. Circ Res 34: 293-301
- Donoghue S (1978) An electrophysiological study of the connections of aortic and cardiac vagal nerves in the brainstem of the cat. Ph.D. thesis, University of Leeds
- Donoghue S, Fox RE, Kidd C (1977) The distribution of aortic nerve afferent fibres in the brainstem of the cat. J Physiol (Lond) 273:80-81P
- Donoghue S, Kidd C, McWilliam PN (1978) The distribution of neurones in the brain stem of the cat activated by A and C fibres of the aortic nerve. J Physiol (Lond) 285:56-57P

- Douglas WW, Ritchie JM (1956) Cardiovascular reflexes produced by electrical excitation of non-medullated afferents in the vagus, carotid sinus and aortic nerves. J Physiol (Lond) 134:167-178
- Douglas WW, Schaumann W (1956) A study of depressor and pressor components of the cat's carotid sinus and aortic nerves using electrical stimulus of different intensities and frequency. J Physiol (Lond) 132:173-186
- Douglas WW, Ritchie JM, Schaumann W (1956) Depressor reflexes from medullated and non-medullated fibres in the rabbit's aortic nerve. J Physiol (Lond) 132:187-198
- Downman CBB (1972) The vasomotor centre. In: Downman CBB (ed) Modern trends in physiology. Butterworths, London, pp 292–308
- Dugin SF, Zakharow ST, Samonina GE, Udelnov MG (1976) The effect of electrical stimulation of the vagal nuclei in anaesthetized and unanaesthetized cats. Sechenov Physiol J USSR 62:382-386
- Elliason S, Folkow B, Lindgren P, Üvnas B (1951) Activation of sympathetic vasodilator nerves to skeletal muscles in the cat by hypothalamic stimulation. Acta Physiol Scand 23:333-351
- Elsner N, Angell-James JE, Daly M de B (1977) Carotid body chemoreceptor reflexes and their interactions in the seal. Am J Physiol 232:H517-H526
- Euler C von, Hayward JN, Marttila I, Wyman RJ (1973a) Respiratory neurones of the ventrolateral nucleus of the solitary tract of cat. Vagal inputs, spinal connections and morphological identification. Brain Res 61:1-22
- Euler C von, Hayward JN, Marttila I, Wyman R (1973b) The spinal connections of the inspiratory neurones of the ventrolateral nucleus of the cat's solitary tract. Brain Res 61:23-33
- Evans MH (1977) Facilitation of reflex bradycardia by hypothalamic stimulation in the anaesthetised rabbit. J Physiol (Lond) 265:33-34P
- Feigl E, Johansson B, Lofving B (1964) Renal vasoconstriction and the defence reaction. Acta Physiol Scand 62:429-435
- Fernandez de Molina A, Kuno M, Perl E (1965) Antidromically evoked responses from sympathetic preganglionic neurones. J Physiol (Lond) 180:321-335
- Fidone SJ, Sato A (1969) A study of chemoreceptor and baroreceptor A and C fibres in the cat carotid nerve. J Physiol (Lond) 205:527-548
- Foley JO, Dubois FS (1934) An experimental study of the rootlets of the vagus nerve in the cat. J Comp Neurol 60:137-159
- Folkow B, Öberg B, Rubinstein EH (1964) A proposed differentiated neuro-effector organisation in muscle resistance vessels. Angiologica 1:197-208
- Folkow B, Lisander B, Tuttle RS, Wang SC (1968) Changes in cardiac output upon stimulation of the hypothalamic defence area and the medullary depressor area in the cat. Acta Physiol Scand 72:220-233
- Foreman RD, Wurster RD (1973) Localization and functional characteristics of descending sympathetic spinal pathways. Am J Physiol 225:212-217
- Fussey IF, Kidd C, Whitwam JG (1973a) The effects of baroreceptors on the latency of evoked responses in sympathetic nerves during the cardiac cycle. J Physiol (Lond) 229:601-616
- Fussey IF, Kidd C, Whitwam JG (1973b) Activity evoked in the brainstem by stimulation of C-fibres in the cervical vagus nerve in the dog. Brain Res 49:436-440
- Gabriel M, Seller H (1970) Interaction of baroreceptor afferents from carotid sinus and aorta at the nucleus tractus solitarii. Pfluegers Arch 318:7-20
- Gahery Y, Ancri D (1967) Projections vago-aortiques et corticales au niveau de la region due noyeau du faisceau solitaire. J Physiol (Paris) 59:408
- Gandevia SC, McCloskey DI, Potter EK (1978) Inhibition of baroreceptor and chemoreceptor reflexes on heart rate by afferents from the lungs. J Physiol (Lond) 276: 369-381

- Garcia M, Jordan D, Spyer KM (1978) Studies on the properties of cardiac vagal neurones. Neurosci Lett (Suppl) 1:S16
- Garcia M, Jordan D, Spyer KM (1979a) The central projections of single vagal afferent neurones in the rabbit. J Physiol (Lond) 289:42-43
- Garcia M, Jordan D, Spyer KM (1979b) Identification of aortic nerve cell bodies in the nodose ganglion of the rabbit and their central connections. J Physiol 290:23-24
- Garcia M, Jordan D, Schneiderman N, Spyer KM (to be published) Vagal cardiomotor neurones identified by the anterograde transport of horseradish peroxidase
- Gebber GL (1976) Basis for phase relations between baroreceptor and sympathetic nervous discharge. Am J Physiol 230:263-270
- Gebber GL, Barman SM (1977) Brainstem vasomotor circuits involved in genesis and entrainment of sympathetic nervous rhythms. Prog Brain Res 47:61-76
- Gebber GL, Barman SM (1979) Inhibitory interaction between preganglionic sympathetic neurons. In: Meyer P, Schmitt H (eds) Nervous system and hypertension. Willey-Flammarion, Paris, pp 131-145
- Gebber GL, Barman SM (to be published) The cardiac-related rhythm in sympathetic nerve discharge (SND): Baroreceptor, brain stem or cortical in origin. In: Sleight P (ed) Baroreceptor and hypertension. Oxford University Press, Oxford
- Gebber GL, Klevans, LR (1972) Central nervous system modulation of cardiovascular reflexes. Fed Prod 31:1245-1252
- Gebber GL, McCall RS (1976) Identification and discharge patterns of spinal sympathetic interneurons. Am J Physiol 231:722-733
- Gebber GL, Snyder DW (1970) Hypothalamic control of baroreceptor reflexes. Am J Physiol 218:124-131
- Gebber GL, Taylor DG, Weaver LC (1973) Electrophysiological studies on organization of central vasopressor pathways. Am J Physiol 224:470-481
- Gebber GL, Barman SM, McCall RB (1978) Recurrent inhibition of preganglionic sympathetic unit discharges. Fed Proc 37:743
- Geis GS, Wurster RD (1978) Localisation of cardiac vagal preganglionic soma. Neurosciences Abstract 4:20
- Geis GS, Barratt G, Wurster RD (1978) Role of descending pressor pathway in conscious and pentobarbital-anaesthetized dog. Am J Physiol 234:H152-156
- Gellhom E (1957) Autonomic imbalance and the hypothalamus. University of Minnesota Press, Minneapolis
- Gellhom E, Nakao H, Redgate ES (1956) The influence of lesions in the anterior and posterior hypothalamus on tonic and phasic autonomic reactions. J Physiol (Lond) 131:402-423
- Ghelarducci B, Pompeiano O, Spyer KM (1974a) Activity of precerebellar reticular neurones as a function of head position. Arch Ital Biol 112:98-125
- Ghelarducci B, Pompeiano O, Spyer KM (1974b) Distribution of the neuronal responses to static tilts within the cerebellar fastigial nucleus. Arch Ital Biol 112: 126-141
- Gimpl MP, Brickman AL, Kaufman MP, Schneiderman N (1976) Temporal relationships during barosensory attenuation in the conscious rabbit. Am J Physiol 230: 1480-1484
- Glees P, Nauta WJH (1955) A critical review of studies on axonal and terminal degeneration. Monatsschr Psychiatr Neurol 129:74–91
- Gootman PM, Cohen MI (1970) Efferent splanchnic activity and systemic arterial blood pressure. Am J Physiol 219:897–903
- Gootman PM, Cohen MI (1971) Evoked splanchnic potentials produced by electrical stimulation of medullary vasomotor regions. Exp Brain Res 13:1–14
- Gootman PM, Cohen MI (1973) Periodic modulation (cardiac and respiratory) of spontaneous and evoked sympathetic discharge. Acta Physiol Pol 24:97–109
- Gootman PM, Cohen MI, Piercey MP, Wolotsky P (1975) A search for medullary neurones with activity patterns similar to those in sympathetic nerves. Brain Res 87: 395-406

- Green JH (1959) Cardiac vagal efferent activity in the cat. J Physiol (Lond) 149:47-49
- Green JH (1965) Physiology of baroreceptor function. Mechanism of receptor stimulation. In: Kezdi P (ed) Baroreceptors and hypertension. Pergamon, Oxford London, pp3-16
- Green JH, Heffron PF (1968) Studies upon the relationship between baroreceptor and sympathetic activity. Q J Exp Physiol 53:406-418
- Gregor M, Janig W, Wiprich L (1977) Cardiac and respiratory rhythmicities in cutaneous and muscle vasoconstrictor neurones to the cat's hindlimb. Pfluegers Arch 370:299-302
- Grösse M, Janig W (1976) Vasoconstrictor and pilomotor fibres in skin nerves to cat's tail. Pfluegers Arch 361:221-229
- Grundfest H (1939) The properties of mammalian B fibres. Am J Physiol 127:252-262
- Guazzi M, Zanchetti A (1965) Blood pressure and heart rate during natural sleep of the cat and their regulation by carotid sinus and aortic reflexes. Arch Ital Biol 103: 789-817
- Gunn CG, Sevelius G, Puiggari MS, Myers FK (1968) Vagal cardiomotor mechanisms in the hindbrain of the dog and cat. Am J Physiol 214:258-262
- Gurevitch MI, Vyshatina AI (1973) Cardiovascular responses to electrical stimulation of the fastigial nuclei. Sechenov Physiol J USSR 59:1715-1722
- Haeusler G (1976) Central adrenergic neurons in experimental hypertension. In: Onesti G, Fernandez M, Kim KE (eds) Regulation of blood pressure by the central nervous system. The Fourth Hahnemann International Symposium on Hypertension (1975). Grune & Stratton, New York, pp 53-64
- Haeusler G (1977) Neuronal mechanisms influencing transmission in the baroreceptor reflex arc. Prog Brain Res 47:95–109
- Hagbarth RE, Vallbo AB (1968) Pulse and respiratory groupings of sympathetic impulses in human muscle nerves. Acta Physiol Scand 74:96-108
- Hamilton RB, Wallach JH, Petrik GK, Schneiderman N (1978) Putative short-latency barosensory circuit linking the baroreceptors, caudal diencephalon, and dorsal vagal nucleus in the rabbit. Neurosciences Abstracts 4:20
- Haymett BT, McCloskey DI (1975) Baroreceptor and chemoreceptor influences on heart-rate during the respiratory cycle in the dog. J Physiol (Lond) 245:699-712
- Heinbecker P (1930) The potential analysis of the turtle and cat sympathetic and vagus nerve trunks. Am J Physiol 93:284-306
- Heinbecker P, O'Leary J (1933) The mammalian vagus nerve functional and histological study. Am J Physiol 106:623-646
- Hellner K, Baumgarten R von (1961) Über ein Endigungsgebiet afferenter, kardiovaskulärer Fasern des Nervus vagus im Rautenhirn der Katze. Pfluegers Arch 273: 223-234
- Henry JL, Calaresu FR (1972) Topography and numerical distribution of neurons of the thoraco-lumbar intermediolateral nucleus in the cat. J Comp Neurol 144:205-214
- Henry JL, Calaresu FR (1974a) Excitatory and inhibitory inputs from medullary nuclei projecting to spinal cardioacceleratory neurons in the cat. Exp Brain Res 20: 485-504
- Henry JL, Calaresu FR (1974b) Pathways from medullary nuclei to spinal cardioacceleratory neurons in the cat. Exp Brain Res 20:505-514
- Henry JL, Calaresu FR (1974c) Origin and course of crossed medullary pathways to spinal sympathetic neurons in the cat. Exp Brain Res 20:515-526
- Henry JL, Calaresu FR (1974d) Responses of single units in the intermediolateral nucleus to stimulation of cardioregulatory medullary nuclei in the cat. Brain Res 77:314-319
- Heymans C, Neil E (1958) Reflexogenic areas of the cardiovascular system. Churchill, London

- Hildebrandt JR (1974) Central connections of the aortic depressor and carotid sinus nerves. Exp Neurol 45:590-605
- Hilton SM (1963) Inhibition of baroreceptor reflexes on hypothalamic stimulation. J Physiol (Lond) 165:56-67
- Hilton SM (1965) Hypothalamic control of the cardiovascular responses in fear and rage. Sci Basis Med 217–233
- Hilton SM (1966) Hypothalamic regulation of the cardiovascular system. Br Med Bull 22:243-248
- Hilton SM (1974) The role of the hypothalamus in the organisation of patterns of cardiovascular response in recent studies of hypothalamic function. In: Lederis K, Cooper KE (eds) Int Symp Calgary 1973. Karger, Basel, pp 306-314
- Hilton SM (1975) Ways of viewing the central nervous control of circulation -- old and new. Brain Res 87:213-219
- Hilton SM (1977) Supramedullary organization of vasomotor control. Brain Res 47: 77-84
- Hilton SM, Spyer KM (1969) The hypothalamic depressor area and the baroreceptor reflex. J Physiol (Lond) 165:56-57
- Hilton SM, Spyer KM (1971) Participation of the anterior hypothalamus in the baroreceptor reflex. J Physiol (Lond) 218:271-293
- Hilton SM, McAllen RM, Spyer KM (1974) Area postrema and blood pressure. Nature 250:354
- Hilton SM, Spyer KM, Timms RJ (1975) Hindlimb vasodilatation evoked by stimulation of the motor cortex. J Physiol (Lond) 252:22-23
- Hilton SM, Spyer KM, Timms RJ (1979) The origin of the hindlimb vasodilatation evoked by stimulation of the motor cortex in the cat. J Physiol 287:545-558
- Hirose K (1916) Über eine bulbo-spinale Bahn. Folia Neurobiol Biol (LP2) 10:371-382
- Hockman CH, Talesnik J, Livingstone KE (1969) Central nervous system modulation of baroreceptor reflexes. Am J Physiol 217:1681–1689
- Hockman CH, Livingstone KE, Talesnik J (1970) Cerebellar modulation of reflex vagal bradycardia. Brain Res 23:101–104
- Hoffer BJ, Ratcheson R, Snider RS (1966) The effects of stimulation of the cerebellum on the circulatory system. Fed Proc 25:701
- Homma S, Miura M, Reis DJ (1970) Intracellular recordings from paramedian reticular neurons monosynaptically excited by stimulation of the carotid sinus nerve. Brain Res 18:185-188
- Hongo T, Ryall RW (1966) Electrophysiological and micro-electrophoretic studies on sympathetic preganglionic neurones in the spinal cord. Acta Physiol Scand 68:96-104
- Horeyseck G, Janig W, Kirchner F, Thamer V (1976) Activation and inhibition of muscle and cutaneous postganglionic neurones to the hindlimb during hypothalamically induced vasoconstriction and atropine-sensitive vasodilation. Pfluegers Arch 361:231-240
- Humphrey DR (1967) Neuronal activity in the medulla oblongata of the cat evoked by stimulation of the carotid sinus nerve. In: Kezdi P (ed) Baroreceptors and hypertension. Pergamon Press, Oxford, pp 131–168
- Humphreys PW, Joels N, McAllen RM (1971) Modification of the reflex response to stimulation of carotid sinus baroreceptors during and following stimulation of the hypothalamic defence area in the cat. J Physiol (Lond) 216:461-482
- Illert M, Gabriel M (1972) Descending pathways in the cervical cord of cats affecting blood pressure and sympathetic activity. Pfluegers Arch 335:109-124
- Illert M, Seller H (1969) A descending sympatho-inhibitory tract in the ventrolateral column of the cat. Pfluegers Arch 313:343-360
- Ingram WR, Dawkins EA (1945) The intramedullary course of afferent fibres of the vagus in the cat. J Comp Neurol 82:157-168

- Iriuchijima J, Kumada M (1963) Efferent cardiac vagal discharge of the dog in response to electrical stimulation of sensory nerves. Jpn J Physiol 13:599–605
- Iriuchijima J, Kumada M (1965) Activity in single vagal efferent fibres to the heart. Jpn J Physiol 14:479-487
- Janig W, Kümmel H (1977) Functional discrimination of postganglionic neurones to the cat's hindpaw with respect to skin potentials recorded from hairless skin. Pfluegers Arch 371:217-225
- Jankowska E, Roberts WJ (1972) An electro-physiological demonstration of the axonal projections of single spinal interneurones in the cat. J Physiol (Lond) 222:597-622
- Jansen J, Brodal A (eds) (1954) Aspects of cerebellar anatomy. Grundt Tanum, Oslo, 423 pp
- Jewett DL (1962) Activity of single vagal efferent cardiac fibres in the dog. J Physiol (Lond) 142:110-126
- Jewett DL (1964) Activity of single efferent fibres in the cervical vagus nerve of the dog with special reference to possible cardioinhibitory fibres. J Physiol (Lond) 175: 321-357
- Jordan D (1977) Studies on the distribution and excitability of sinus and aortic nerve afferent terminals. Ph.D. thesis, University of Birmingham
- Jordan D, Khalid MEM, Schneiderman N, Spyer KM (1979) Preganglionic vagal cardiomotor neurones in the rabbit; location and properties. J Physiol (Lond) 296:20–21
- Jordan D, Spyer, KM (1977a) Studies on the termination of sinus nerve afferents. Pfluegers Arch 369:65-73
- Jordan D, Spyer KM (1977b) Is presynaptic inhibition responsible for suppression of the baroreceptor reflex during the defence reaction? J Physiol (Lond) 258:58
- Jordan D, Spyer KM (1978a) The excitability of sinus nerve afferent terminals during the respiratory cycle. J Physiol (Lond) 277:66
- Jordan D, Spyer KM (1978b) The distribution and excitability of myelinated aortic nerve afferent terminals. Neurosci Lett 8:113-117
- Jordan D, Spyer KM (1979) Studies on the excitability of sinus nerve afferent terminals. J Physiol (Lond) 297:123-134
- Kahn N, Mills E (1967) Centrally evoked sympathetic discharge: A functional study of medullary vasomotor areas. J Physiol (Lond) 191:339-352
- Kalia M (1977) Neuroanatomical organisation of the respiratory centers. Fed Proc 36: 2405-2411
- Kannan H, Yagi K (1978) Supraoptic neurosecretory neurones: evidence for the existence of converging inputs both from carotid baroreceptors and osmoreceptors. Brain Res 145:385-390
- Katona PG, Jih F (1975) Respiratory sinus arrhythmia. Non-invasive measure of parasympathetic cardiac control. J Appl Physiol 39:801-805
- Katona PG, Tan KS (1975) Interaction of aortic and carotid sinus baroreceptors; effect of activation times. Am J Physiol 228:238-248
- Katona P, Poitras J, Barnett U, Terry B (1970) Cardiac vagal efferent activity and heart period in the carotid sinus reflex. Am J Physiol 218:1030-1037
- Katona PG, Lipson D, Dauchot PJ (1977) Opposing central and peripheral effects of atropine on parasympathetic cardiovascular control. Am J Physiol 232:146-151
- Katz DM, Karten HJ (9179) The discrete anatomical localization of vagal aortic afferents within a catecholamine-containing cell group in the nucleus solitarius. Brain Res 191:187-195
- Katz RL, Kahn N, Wang SC (1967) Brainstem mechanisms subserving baroreceptor reflexes, factors affecting the carotid occlusion response. In: Kezdi P (ed) Baroreceptors and hypertension. Pergamon Press, Oxford, pp 169-180
- Kaufman MP, Hamilton RB, Wallach JH, Petrik GK, Schneiderman N (to be published) Lateral subthalamic area as a mediator of bradycardia responses in rabbits. Am J Physiol

- Keith IC, Kidd C, Linden RJ, Snow HM (1975a) Modification of neuronal activity in the dog medulla oblongata by stimulation of the left atrial receptors. J Physiol (Lond) 245:80-81
- Keith IC, Kidd C, Penna PE (1975b) Modification of sympathetic chronotropic carotid sinus reflex responses by hypothalamic stimulation. J Physiol 232:77
- Kent BB, Drane JW, Manning JW (1971) Supraportine contributions to the carotid sinus reflex in the cat. Circ Res 29:534-541
- Kerr FWL (1962) Facial, vagal and glossopharyngeal nerves in the cat. Arch Neurol 6:264-281
- Kerr FWL (1967) Function of the dorsal motor nucleus of the vagus. Science 157: 451-452
- Kerr FWL (1969) Preserved vagal visceromotor function following destruction of the dorsal motor nucleus. J Physiol 202:755-769
- Kerr FWL, Alexander S (1964) Descending autonomic pathways in the spinal cord. Arch Neurol 10:249-261
- Kezdi P (ed) (1967) Baroreceptors and hypertension. Proceedings of the International Symposium Dayton, Ohio, 16–17th November 1965. Pergamon Press, Oxford
- Kezdi P, Geller E (1968) Baroreceptor control of postganglionic sympathetic nerve discharge. Am J Physiol 214:427–435
- Kimmel DL (1965) The termination of vagal and glossopharyngeal afferent nerve fibers in the region of the cat, rat and guinea pig. Anat Rec 151:371
- Kimmel DL, Kimmel DB (1964) Spinal distribution of glossopharyngeal and vagal afferent fibers in the cat, rat and guinea pig. Anat Rec 148:299-300
- Kirchheim HR (1976) Systemic arterial baroreceptor reflexes. Physiol Rev 56:100-176
- Kirchner F, Kirchner D, Polosa C (1975a) Spinal organisation of sympathetic inhibition by spinal afferent volleys. Brain Res 87:161-170
- Kirchner F, Wyszogrodski I, Polosa C (1975b) Some properties of sympathetic neuron inhibition by depressor area and intraspinal stimulation. Pfluegers Arch 357:349-360
- Klee MR (1966) Different effects on the membrane potential of motor cortex units after thalamic and reticular stimulation. In: Purpura DP, Yahr MD (eds) The thalamus. Columbia University Press, New York London, pp 287-322
- Klevans LR, Gebber GL (1970) Facilitatory forebrain influence on cardiac component of baroreceptor reflexes. Am J Physiol 219:1235-1241
- Koch E (1932) Die Irradiation der pressoreceptorischen Kreislaufreflexe. Klin Wochenschr 11:225-227
- Koepchen HP, Lux HD, Wagner PH (1961a) Untersuchungen über Zeitbedarf und zentrale Verarbeitung des pressoreceptischen Herzreflexes. Pfluegers Arch 273: 413-430
- Koepchen HP, Wagner PH, Lux HD (1961b) Über die Zusammenhänge zwischen zentraler Erregbarkeit reflektorischen Atemrhythmus bei der nervösen Steuerung der Herzfrequenz. Pfluegers Arch 273:443-465
- Koepchen HP, Langhorst P, Seller H, Polster J, Wagner D (1967) Neuronale Aktivität im unteren Hirnstamm mit Beziehung zum Kreislauf. Pfluegers Arch 294:40-64
- Koepchen HP, Langhorst P, Seller H (1975) The problem of identification of autonomic neurons in the lower brainstem. Brain Res 87:375-393
- Kötter V, Zerbst E, Schulze A (1970) Untersuchungen zur Bahnung und Konvergenz pressorezeptorischer Afferenzen mit Hilfe der Konditionstestreiztechnik. Pfluegers Arch 320:210-232
- Koizumi K, Sato A (1969) Influence on sympathetic innervation on carotid sinus baroreceptor activity. Am J Physiol 216:321–329
- Koizumi K, Seller H, Kaufman A, Brooks C McC (1971) Pattern of sympathetic discharges and their relation to baroreceptor and respiratory activities. Brain Res 27: 281-294

Korner PI (1971) Integrative neural cardiovascular control Physiol Rev 51:312–367

Kosaka K (1909) Über die Vaguskerne des Hundes, Neurol CBL 28:406-416

- Kosaka K, Yagita K (1905) Experimentelle Untersuchungen über den Ursprung des N. vagus und die zentralen Endigungen der dem Plexus nodosus entstammenden sensiblen Vagusfasern sowie über den Verlauf ihrer sekundären Bahn. Neurol Med Chir (Tokyo) 4:29-49
- Kumada M, Nakajima H (1972) Field potentials evoked in rabbit brainstem by stimulation of the aortic nerve. Am J Physiol 223:575–582
- Kumada M, Sagawa K (1974) Modulation of the baroreceptor reflex by central gray stimulation. J Physiol Soc Jpn 36:147-148
- Kumada M, Nogami K, Sagawa K (1975a) Modulation of carotid sinus baroreceptor reflex by sciatic nerve stimulation. Am J Physiol 228:1535-1541
- Kumada M, Schramm LP, Altansberger RA, Sagawa K (1975b) Modulation of carotid sinus baroreceptor reflex by hypothalamic defence response. Am J Physiol 228: 34-45
- Kunze DL (1972) Reflex discharge patterns of cardiac vagal efferent fibres. J Physiol (Lond) 222:1-15
- Kuypers HGJM, Maisky VA (1975) Retrograde axonal transport of horseradish peroxidase from spinal cord to brainstem cell groups in the cat. Neurosci Lett 1:9-14
- Kylstra PH, Lisander B (1970) Differentiated interaction between hypothalamic defence reaction and baroreceptor reflexes. Acta Physiol Scand 78:386-392
- Lam RL, Tyler HR (1952) Electrical responses in visceral afferent nucleus of the rabbit by vagal stimulation. J Comp Neurol 97:21-36
- Landgren S (1952) On the excitation mechanism of the carotid baroreceptors. Acta Physiol Scand 26:1-34
- Langhorst P, Werz M (1974) Concept of functional organization of brainstem "cardiovascular center". In: Umbach W, Koepchen HP (eds) Central rhythmic and regulation. Hippokrates, Stuttgart, pp 238-255
- Langhorst P, Stroh-Werz M, Dittmar K, Camerer H (1975) Facultative coupling of reticular neuronal activity with peripheral cardiovascular and central cortical rhythms. Brain Res 87:407-418
- Lawn AM (1964) The localisation by means of electrical stimulation of the origin and path in the medulla oblongata of the motor nerve fibres of the rabbit oesophagus. J Physiol (Lond) 174:323-344
- Lawn AM (1966) The nucleus ambiguous of the rabbit. J Comp Neurol 127:307-320
- Lebedev VP, Petrov VI, Skobelev VA (1980) Do sympathetic preganglionic neurones have a recurrent inhibitory mechanism. Pfluegers Arch 385:91-97
- Lee TM, Kuo JS, Chai CY (1972) Central integrative mechanism of the Bezold-Jarisch and baroreceptor reflexes. Am J Physiol 222:713-720
- Levy MN (1977) Parasympathetic control of the heart. In: Randal WC (ed) Neural regulation of the heart. Oxford University Press, New York, pp 95-130
- Levy MN, Zieske H (1969) Autonomic control of cardiac pacemaker activity and atrioventricular transmission. J Appl Physiol 27:465-470
- Levy MN, Degeest H, Zieske H (1966) Effects of respiratory centre activity on the heart. Circ Res 18:67-78
- Lipski J, Trzebski A (1975) Bulbospinal neurones activated by baroreceptor afferents and their possible role in inhibition of preganglionic sympathetic neurons. Pfluegers Arch 356:181-192
- Lipski J, McAllen RM, Spyer, KM (1972) Localisation of sinus nerve afferent endings in the brainstem. J Physiol (Lond) 225:30-31P
- Lipski J, McAllen RM, Spyer KM (1975) The sinus nerve and baroreceptor input to the medulla of the cat. J Physiol (Lond) 251:61-78
- Lipski J, McAllen RM, Spyer KM (1976a) Synaptic activation of cardiac vagal motoneurones. J Physiol (Lond) 256:68

- Lipski J, McAllen RM, Spyer KM (1976b) The carotid body chemoreceptor input to the inspiratory neurones of the nucleus of the tractus solitarious. J Physiol (Lond) 258:115-116
- Lipski J, McAllen RM, Trzebski A (1976c) Carotid baroreceptor and chemoreceptor inputs onto single medullary neurones. Brain Res 107:132–136
- Lipski J, Coote JH, Trzebski A (1977a) Temporal patterns of antidromic invasion latencies of sympathetic preganglionic neurons related to central inspiratory activity and pulmonary stretch receptor reflex. Brain Res 135:162-166
- Lipski J, McAllen RM, Spyer KM (1977b) The carotid chemoreceptor input to the respiratory neurones of the nucleus of the tractus solitarious. J Physiol (Lond) 269: 797-810
- Lisander B, Martner J (1971a) Cerebellar suppression of the autonomic components of the defence reaction. Acta Physiol Scand 81:84-95
- Lisander B, Martner J (1971b) Interaction between the fastigial pressor response and the baroreceptor reflex. Acta Physiol Scand 83:505-514
- Lisander B, Martner J (1973) Interaction between the fastigial pressor response and the defence reaction. Acta Physiol Scand 87:359-367
- Little R, Wennergren G, Öberg B (1975) Aspects of the central interaction of arterial baroreceptor and cardiac ventricular receptor reflexes in the cat. Acta Physiol (Scand) 93:85-96
- Löfving B (1961) Cardiovascular adjustments induced from the rostral cingulate gyrus. Acta Physiol Scand (Suppl) 184:1–82
- Loewy AD, Burton H (1978) Nuclei of the solitary tract. Efferent projections to the lower brainstem and spinal cord of the cat. J Comp Neurol 181:421-450
- Loizou LA (1969) Projections of the locus coerulus in the albino rat. Brain Res 15: 563-566
- Lopes OU, Palmer JF (1976a) Proposed respiratory 'gating' mechanism for cardiac slowing. Nature 264:454-456
- Lopes OU, Palmer JF (1976b) Hypothalamic inhibiton of vagal component of the sinus nerve cardiac reflex. J Physiol (Lond) 260:50
- Lopes OU, Palmer JF (1978) Mechanism of hypothalamic control of cardiac component of sinus nerve reflex. Q J Exp Physiol 63:231–254
- Magnes J, Moruzzi G, Pompeiano O (1961) Synchronization of the EEG produced by low frequency electrical stimulation of the region of the solitary tract. Arch Ital Biol 99:33-67
- Mannard A, Polosa C (1973) Analysis of background firing of single sympathetic preganglionic neurons of cat cervical nerve. J Neurophysiol 36:398-408
- Manning JW (1962) Lesions of the medullary reticular formation and cardiovascular reflexes. Fed Proc 21:107
- Manning JW (1965a) Intracranial representation of cardiac control In: Randall WC (ed) Nervous control of the heart. Williams & Wilkins, Baltimore, pp 16-33
- Manning JW (1965b) Cardiovascular reflexes following lesions in the medullary reticular formation. Am J Physiol 208:283-288
- Marguth H, Raule W, Schaeffer H (1951) Aktionsströme in zentrifugale Herznerven. Pfluegers Arch 254:224-245
- Marshall JM (1977) The cardiovascular response to stimulation of carotid chemoreceptors. J Physiol 266:48-49
- Martin GF, Humbertson AO, Laxson C, Parreton WM (1979) Evidence for direct bulbospinal projections to lamina IX, X and the intermediolateral cell column. Studies using axonal transport techniques in the North American opposum. Brain Res 170: 165-171
- McAllen RM (1973) Projections of the carotid sinus baroreceptors to the medulla of the cat. Ph D thesis, University of Birmingham
- McAllen RM (1976) The inhibition of the baroreceptor input to the medulla by stimulation of the hypothalamic defence area. J Physiol (Lond) 257:45-46

- McAllen RM, Spyer KM (1972) 'Baroreceptor' neurones in the medulla of the cat. J Physiol (Lond) 222:68-69
- McAllen RM, Spyer KM (1975) The origin of cardiac vagal neurones in the medulla of the cat. J Physiol 244:82-83
- McAllen RM, Spyer KM (1976) The location of cardiac vagal preganglionic motoneurones in the medulla of the cat. J Physiol (Lond) 258:187-204
- McAllen RM, Spyer KM (1977) Bradycardia produced by iontophoretic activation of preganglionic vagal motoneurones. J Physiol (Lond) 269:49
- McAllen RM, Spyer KM (1978a) Two types of vagal preganglionic motoneurones projecting to the heart and lungs. J Physiol 282:353-364
- McAllen RM, Spyer KM (1978b) The baroreceptor input to cardiac vagal motoneurones. J Physiol (Lond) 282:365-374
- McAllen RM, Jordan D, Spyer KM (1979) The carotid baroreceptor input to the brain. In: Koepchen HP, Hiltom SM, Trzebski A (eds) Central interactions between respiratory and cardiovascular control systems. Springer, Berlin Heidelberg New York pp 87-92
- McCall RS, Gebber GL, Barman SM (1977) Spinal interneurones in the baroreceptor reflex arc. Am J Physiol 232:H657–665
- Merrill EG (1974) Finding a respiratory function for the medullary respiratory neurons. In: Bellairs R, Gray EG (eds) Essays on the nervous system. Clarendon Press, Oxford, pp 451-486
- Mesulam MM (1978) Tetramethyl benzidine for horseradish peroxidase neurohistochemistry: A non-carcinogenic blue reaction product with superior sensitivity for visualizing neural afferents and efferents. J Histochem Cytochem 26:106-117
- Middleton S, Middleton HH, Grundfest H (1950) Spike potentials and cardiac effects of mammalian vagus nerve. Am J Physiol 162:553-559
- Middleton S, Woolsley CN, Burton H, Rose JE (1973) Neural activity with cardiac periodicity in the medulla oblongata of the cat. Brain Res 50:297-314
- Miller FR, Bowman JT (1915) The cardio-inhibitory center. Am J Physiol 39:149-153
- Miller R (1976) Identification of vagal efferent neurones; a horseradish peroxidase study. J Physiol (Lond) 256:69-70
- Mitchell GAG, Warwick R (1955) The dorsal vagal nucleus. Acta Anat 25:371-395
- Miura M (1975) Postsynaptic potentials recorded from the nucleus of the solitary tract and its subadjacent reticular formation elicited by stimulation of the carotid sinus nerve. Brain Res 100:437-440
- Miura M, Kitamura T (1979) Postsynaptic potentials recorded from medullary neurones following stimulation of carotid sinus nerve. Brain Res 162:261-272
- Miura M, Reis DJ (1968) Electrophysiological evidence that the carotid sinus nerve fibers terminate in the bulbar reticular formation. Brain Res 9:394-397
- Miura M, Reis DJ (1969a) Termination and secondary projections of the carotid sinus nerve in the cat brainstem. Am J Physiol 217:142-153
- Miura M, Reis DJ (1969b) Cerebellum: A pressor response elicited from the fastigial nucleus and its efferent pathway in the brainstem. Brain Res 13:595-599
- Miura M, Reis DJ (1970) À blood pressure response from the fastigial nucleus and its relay pathway in the brainstem. Am J Physiol 219:1330-1336
- Miura M, Reis DJ (1972a) The paramedian reticular nucleus: A site of inhibitory interaction between projections from fastigial nucleus and carotid sinus nerve acting on blood pressure. J Physiol (Lond) 216:441-460
- Miura M, Reis DJ (1972b) The role of the solitary and paramedian reticular nuclei in mediating cardiovascular reflex responses from carotid baro- and chemoreceptors. J Physiol (Lond) 225:525-548
- Mohlant M (1910) Le nerv vagus: Les connexions anatomiques et la valeur functioneue en noyar en vagus. Nevraxe 11:131

- Morest PK (1967) Experimental study of the projections of the nucleus of the tractus solitarius and the area postrema in the cat. J Comp Neurol 130:277-293
- Moruzzi G (1938) Azione del paleocerebellum sui riflessi vasomotori. Arch Fisiol 38: 36-78
- Moruzzi G (1940) Paleocerebellum inhibition of vasomotor and carotid sinus reflexes. J Neurophysiol 3:20–32
- Moruzzi G (1947) Sham rage and localised autonomic responses elicited by cerebellar stimulation in the acute thalamic cat. Proc. IVII Internat Congress Physiol Oxford, pp 114-115
- Moruzzi G (1950) Problems in cerebellar physiology. Thomas, Springfield, Ill
- Nathan M (1972) Pathways in the medulla oblongata of monkeys mediating splanchnic nerve activity. Electrophysiological and anatomical evidence. Brain Res 45:115-126
- Nathan MA, Reis DJ (1975) Fulminating arterial hypertension with pulmonary edema from release of adrenomedullary catecholamines after lesions of the anterior hypothalamus in the rat. Circ Res 37:226-235
- Neil E, Palmer JF (1975) Effects of spontaneous respiration on the latency of reflex cardiac chronotropic responses to baroreceptor stimulation. J Physiol (Lond) 247: 16
- Norgren R (1978) Projections from the nucleus of the solitary tract in the rat. Neuroscience 3:207-218
- Norgren R, Leonard CM (1973) Ascending central gustatory pathways. J Comp Neurol 150:217-238
- Nosaka S (1976) Responses of rat brainstem neurons to carotid distension. Am J Physiol 231:20-27
- Nosaka S, Yamamoto T, Yasunaga K (1979) Localization of vagal cardioinhibitory preganglionic neurons within the rat brain stem. J Comp Neurol 186:79-82
- Oberholzer RJH (1960) Circulatory centres in medulla and midbrain. Physiol Rev (Suppl 4) 40:179-195
- Öberg B, Thoren P (1973a) Circulatory responses to stimulation of medullated and non-medullated vagal afferents in the cardiac nerve of the cat. Acta Physiol Scand 87:121-132
- Öberg B, Thoren P (1973b) Circulatory responses to stimulation of left ventricular receptors in the cat. Acta Physiol Scand 88:8-22
- Okada H, Okamoto K, Nishida I (1961a) The activity of the cardioregulatory and abdominal sympathetic nerves during swallowing. Jpn J Physiol 11:44-53
- Okada H, Okamoto K, Nishida I (1961b) The activity of the cardioregulatory and abdominal sympathetic nerves in the cat in the Bainbridge reflex. Jpn J Physiol 11: 520:529
- Owsjannikow P (1871) Die tonischen und reflektorischen Zentren der Gefäßnerven. K. sächs. Ges der Wiss. Mathematischen-physische Klasse. Ber 23:135–147
- Paintal AS (1972) Cardiovascular receptors. In: Neil E (ed) Enteroreceptors. Springer, Berlin Heidelberg New York (Handbook of sensory physiology, vol III/1, pp 1-46)
- Paintal AS (1973) Vagal sensory receptors and their reflex effects. Physiol Rev 53: 159-227
- Palkovits M, Zaborsky L (1977) Neuroanatomy of central cardiovascular control. Prog Brain Res 47:9-34
- Peiss CN (1965) Concepts of cardiovascular regulation; past, present, future. In: Randall WC (ed) Nervous control of the heart. Williams & Wilkins, Baltimore, pp 154-197
- Pelletier CL, Clement DL, Shepherd JT (1972) Comparison of afferent activity of canine aortic and sinus nerves. Circ Res 31:557-568
- Perez-Gonzalez JF, Rojas JR (1976) Inhibition of the baroreceptor input to cardiac vagal preganglionic neurones by stimulation of the medullary reticular formation. J Physiol (Lond) 263:152

- Petras JM, Cummings JF (1972) Autonomic neurons in the spinal cord of the rhesus monkey: a correlation of the findings of cytoarchitectonics and sympathectomy with fiber degeneration following dorsal rhizotomy. J Comp Neurol 146:189-218
- Pitts RF, Bronk DW (1942) Excitability cycle of the hypothalamus-sympathetic neuron system. Am J Physiol 135:504-522
- Pitts RF, Larrabee MG, Bronk DN (1941) An analysis of hypothalamic cardiovascular control Am J Physiol 134:359–383
- Polosa C (1967) The silent period of sympathetic preganglionic neurons. Can J Physiol Pharmacol 45:1033-1045
- Polosa C (1968) Spontaneous activity of sympathetic preganglionic neurons. Can J Physiol Biochem 46:587-896
- Porszasz J, Proszasz-Gibischer K (1968) Effect of blood pressure on single unit activity in bulbar reticular formation. Acta Physiol Acad Sci Hung 34:249–258
- Porter R (1963) Unit responses evoked in the medulla oblongata by vagus nerve stimulation. J Physiol (Lond) 168:717-735
- Preiss G, Polosa C (1977) The relation between end-tidal CO<sub>2</sub> and discharge patterns of sympathetic preganglionic neurons. Brain Res 122:255–267
- Preiss G, Kirchner, F, Polosa C (1975) Patterning of sympathetic preganglionic firing by the central respiratory drive. Brain Res 87:363-374
- Preobrazhenski NN (1966) Microelectrode recordings of activity from neurons in vasomotor center. Fed Proc 25:T18-22
- Prichard BNC (1978)  $\beta$ -adrenergic receptor blockade in hypertension, past, present and future. Br J Clin Pharmacol 5:379–399
- Przybyla AC, Wang SC (1967) Neurophysiological characteristics of cardiovascular neurons in the medulla oblongata of the cat. J Neurophysiol 30:645-660
- Quest JA, Gebber GL (1972) Modulation of baroreceptor reflexes by somatic afferent stimulation. Am J Physiol 222:1251-1259
- Reis DJ, Cuenod M (1965) Central neural regulation of carotid baroreceptor reflexes in the cat. Am J Physiol 209:1267-1277
- Reis DJ, Doba N, Snyder DW, Nathan MA (1977) Brain lesions and hypertension; chronic lability and elevaton of arterial pressure produced by electrolytic lesions and 6-hydroxydopamine treatment of nucleus tractus solitari (NTS) in rat and cat. Prog Brain Res 47:169-188
- Rethelyi M (1972) Cell and neurophil architecture of the intermediolateral (sympathetic) nucleus of the cat spinal cord. Brain Res 46:203-213
- Rhoton AL, O'Leary JL, Ferguson JP (1966) The trigeminal, facial and glossopharyngeal nerves in the monkey. Arch Neurol 14:530-540
- Ricardo JA, Koh ET (1978) Anatomical evidence of direct projections from the nucleus of the solitary tract to the hypothalamus, amygdala, and other forebrain structures in the rat. Brain Res 153:1-26
- Richter DW, Seller H (1975) Baroreceptor effects on medullary respiratory neurones of the cat. Brain Res 86:168-171
- Richter DW, Keck W, Seller H (1970) The course of inhibiton of sympathetic activity during various patterns of carotid sinus nerve stimulation. Pfluegers Arch 317:110-123
- Richter DW, Heyde F, Gabriel M (1975) Intracellular recordings from different types of medullary respiratory neurons of the cat. J Neurophysiol 38:1162-1171
- Rijlant P (1936a) Le controle extrinsique de la frequence du battement cardiaque. CR Soc Biol (Paris) 123:99-101
- Rijlant P (1936b) L'arhythmie cardiaque respiratoire. CR Soc Biol (Paris) 123:997-1001
- Robertson TW, Wallach J, Schneiderman N, Neumann P (1976) Fiber origins of cervical vagus nerve in rabbits, cats and rhesus monkeys investigated by injection of horseradish peroxidase. Neurosci Abstr 2:175

Neural Organisation and Control of the Baroreceptor Reflex

- Rudomin P (1966) Pharmacolocial evidence for the existence of interneurones mediating primary afferent depolarisation in the solitary tract nucleus of the cat. Brain Res 2:181-183
- Rudomin P (1967a) Primary afferent depolarisation produced by vagal visceral afferents. Experientia 23:117-119
- Rudomin P (1967b) Presynaptic inhibiton induced by vagal afferent volleys. J Neurophysiol 30:964-981
- Rudomin P (1968) Excitability changes of superior laryngeal, vagal and depressor afferent terminals produced by stimulation of the solitary tract nucleus. Exp Brain Res 6:156-170
- Sakai K, Touret M, Salvert D, Leger L, Jouvet M (1977) Afferent projections to the cat locus coeruleus as visualized by the horseradish peroxidase technique. Brain Res 119:21-41
- Salmoiraghi GC (1962) "Cardiovascular" neurones in the brainstem of cat. J Neurophysiol 25:182-197
- Samodelov LF, Godehard E, Arndt JO (to be published) A comparison of the stimulus response curves of aortic and carotid sinus baroreceptors in decerebrate cats. Pfluegers Arch
- Saper C, Loewy AD, Swanson LW, Cowan WN (1976) Direct hypothalamo-autonomic connections. Brain Res 117:305-312
- Sato A, Schmidt RF (1973) Somato sympathetic reflexes: afferent fibers, central pathways, discharge characteristics. Physiol Rev 53:916-947
- Schaeffer H (1960) Central control of cardiac function. Physiol Rev (Suppl 4) 40: 213-231
- Scheibel ME, Scheibel AB, Mollica A, Moruzzi G (1955) Convergence and interaction of afferent impulses on single units of reticular formation. J Neurophysiol 18:309– 331
- Scherrer H (1966) Inhibition of sympathetic discharge by stimulation of the medulla oblongata in the rat. Acta Neuroveg 29:56-74
- Schmidt RF, Schönfuss K (1970) An analysis of the reflex activity in the cervical .sympathetic trunk induced by myelinated somatic afferents. Pfluegers Arch 314: 175-198
- Schramm LP Bignall KE (1971) Central neural pathways mediating active sympathetic muscle vasodilation in the cat. Am J Physiol 221:754-767
- Schulte FJ, Henatsch HD, Busch G (1959) Über den Einfluß der Carotissinus-Sensibilität auf die spinal motorischen Systeme. Pfluegers Arch 269:248-263
- Schwaber JS, Cohen DH (1978a) Electrophysiological and electron microscopic analysis of the vagus nerve of the pigeon, with particular reference to the cardiac innervation. Brain Res 147:65-78
- Schwaber JS, Cohen DH (1978b) Field potential and single unit analyses of the avian dorsal motor nucleus of the vagus and criteria for identifying vagal cardiac cells of origin. Brain Res 147:79-90
- Schwaber J, Schneiderman N (1975) Aortic nerve-activated cardioinhibitory neurons and interneurons. Am J Physiol 229:783-785
- Schweitzer A (1935) Zur Frage der respiratorischen Arrhythmie. Verh Dtsch Ges Kreislaufforsch 8:148-153
- Schweitzer A, Wrights S (1937) Effects on the knee jerk of stimulation of the central end of the vagus and of various changes in the circulation and respiration. J Physiol (Lond) 88:459-475
- Sears T (1964) Efferent discharges in alpha and fusimotor fibres of intercostal nerves of the cat. J Physiol (Lond) 174:295-315
- Seller H (1973) The discharge pattern of single units in thoracic and lumbar white rami in relation to cardiovascular events. Pfluegers Arch 343:317-330
- Seller H, Illert M (1969) The localisation of the first synapse in the carotid sinus baroreceptor reflex pathway and its alteration of the afferent input. Pfluegers Arch 306: 1-19

- Seller H, Richter DW (1971) Some quantitative aspects of the central transmission of the baroreceptor activity. In: Kao FF, Koizumi K, Vasalle M (eds) Research in physiology. Gaggi, Bologna, pp 541-545
- Seller H, Langhorst P, Richter D, Koepchen HP (1968) Über die Abhängigkeit der pressorezeptorischen Hummung des Sympathicus von der Atemphase und ihre Auswirkung in der Vasomotorik. Pfluegers Arch 302:300-314
- Shepherd JT (1973) Intrathoracic baroreflexes. Mayo Clin Proc 48:426-436
- Smith OA (1965) Anatomy of central neural pathways mediating cardiovascular functions. In: Randall WC (ed) Nervous control of the heart. Williams & Wilkins, Baltimore, pp 34-53
- Smith OA (1974) Reflex and central mechanisms involved in the control of the heart and circulation. Ann Rev Physiol 36:93-123
- Smith OA, Nathan MA (1966) Inhibition of carotid sinus reflex by inferior olive. Science 154:674-675
- Smith RS, Pearce JW (1961) Microelectrode recordings from the region of the nucleus solitarius in the cat. Can J Biochem 39:933-939
- Smolen AJ, Truex RC (1977) The dorsal motor nucleus of the vagus nerve of the cat; localisation of preganglionic neurons by quantitative histological methods. Anat Rec 189:555-566
- Snyder DW, Gebber GL (1973) Relationships between medullary depressor region and central vasopressor pathways. Am J Physiol 225:1129–1137
- Snyder DW, Nathan MA, Reis DJ (1978) Chronic lability of arterial pressure produced by selective destruction of the catecholamine innervation of the nucleus tractus solitarii in the rat. Circ Res 43:662-671
- Spyer KM (1969) The central nervous organisation of the carotid sinus baroreceptor reflex. Ph.D. thesis, University of Birmingham
- Spyer KM (1972) Baroreceptor sensitive neurones in the anterior hypothalamus of the cat. J Physiol (Lond) 224:245-257
- Spyer KM (1975) Organisation of baroreceptor pathways in the brainstem. Brain Res 87:221-276
- Spyer KM (1979) The baroreceptor control of vagal preganglionic activity. In: Brooks C Mc, Koizumi K, Sato A (eds) Integrative functions of the autonomic nervous system. University of Tokyo Press, Tokyo, pp 283-292
- Spyer KM, McAllen RM (1979) The interaction of central and peripheral inputs onto vagal cardiomotor neurones. In: Koepchen HP, Hilton SM, Trzebski A (eds) Central interactions between respiratory and cardiovascular systems. Springer, Berlin Heidelberg New York, pp 8–14
- Spyer KM, Wolstencroft JH (1971) Problems of the afferent input to the paramedial reticular nucleus, and the central connections of the sinus nerve. Brain Res 26: 411-414
- Stegemann J, Müller-Büton H (1966) Zur regeltheoretischen Analyse des Blutkreislaufs.
  I. Die zentrale Verrechnung der Signale aus den einzelnen Pressorezeptorenfeldern.
  Pfluegers Arch 287:247-256
- Stroh-Werz M, Langhorst P, Camerer H (1976) Neuronal activity with relation to cardiac rhythm in the lower brainstem of the dog. Brain Res 106:293-305
- Stroh-Werz M, Langhorst P, Camerer H (1977a) Neuronal activity with cardiac rhythm in the nucleus of the solitary tract in cats and dogs. 1. Different discharge patterns related to the cardiac cycle. Brain Res 133:65-80
- Stroh-Werz M, Langhorst P, Camerer H (1977b) Neuronal activity with cardiac rhythm in the nucleus of the solitary tract in cats and dogs. II. Activity modulation in relationship to the respiratory cycle. Brain Res 133:81-93
- Swanson LW (1977) Immunohistochemical evidence for a neurophysin-containing autonomic pathway arising in the paraventricular nucleus of the hypothalamus. Brain Res 128:346-353
- Szentagothai J (1952) The general visceral efferent column of the brain stem. Acta Morphol Acad Sci Hung 2:313-328

- Szulczyk P (1976) Descending spinal sympathetic pathway utilized by somato-sympathetic reflex and carotid chemoreflex. Brain Res 112:190–193
- Szulczyk P, Trzebski A (1977) Effects of carotid chemoreceptor and baroreceptor stimulation upon the sympathetic preganglionic and postganglionic cardiac nerve and single fiber activity in cats. Acta Neurbiol Exp (warsz) 37:15-26
- Taylor DG, Gebber GL (1973) Sympathetic unit responses to stimulation of cat medulla. Am J Physiol 225:1138-1146
- Taylor DG, Gebber GL (1975) Baroreceptor mechanisms controlling sympathetic nervous rhythms of central origin. Am J Physiol 228:1002–1013
- Thomas MR, Calaresu FR (1972) Responses of single units in the medial hypothalamus to electrical stimulation of the carotid sinus nerve in the cat. Brain Res 44:49–62
- Thomas MR, Calaresu FR (1973) Hypothalamic inhibition of chemoreceptor-induced bradycardia in the cat. Am J Physiol 225:201-208
- Thomas MR, Calaresu FR (1974a) Localization and function of medullary sites mediating vagus bradycardia. Am J Physiol 226:1344-1349
- Thomas MR, Calaresu FR (1974b) Medullary sites involved in hypothalamic inhibition of reflex vagal bradycardia in the cat. Brain Res 80:1–16
- Thomas MR, Ulrichsen RF, Calaresu FR (1977) Function of lateral reticular nucleus in central cardiovascular regulation in the cat. Am J Physiol 232:157–166
- Timms RJ (1977) Cortical inhibition and facilitation of the defence reaction. J Physiol (Lond) 266:98–99
- Todo K, Yamamoto T, Satomi H, Ise H, Takata H, Takahashi I (1977) Origins of vagal preganglionic fibers to the sino-atrial and atrio-ventricular node regions in the cat heart as studied by the horseradish peroxidase method. Brain Res 130:545-550
- Torvik A (1956) Afferent connections to the sensory trigeminal nuclei, the nucleus of the solitary tract and adjacent structures. J Comp Neurol 106:51-141
- Torvik A (1957) The spinal projection from the nucleus of the solitary tract. An experimental study in the cat. J Anat 91:314-332
- Trzebski A, Peterson LH (1964) The patterns of activity of the medullary respiratory neurones related to stimulation of the carotid body and carotid sinus receptors. In: Aviado DM (ed) Drugs and respiration. Pergamon Press, New York, pp 59-70
- Trzebski A, Peterson LH, Attinger F, Jones A, Tempest R (1962) Unitary responses in the medulla oblongata related to carotid sinus baroreceptor functions. Physiologist 5:222
- Wang SC, Borison HL (1974a) An analysis of the carotid sinus cardiovascular reflex mechanism. Am J Physiol 150:712-721
- Wang SC, Borison HL (1974b) Decussation of the pathways in the carotid sinus cardiovascular reflex. An example of the principle of convergence. Am J Physiol 150: 722-728
- Wang SC, Chai CY (1962) Central control of sympathetic cardioacceleration in the medulla oblongata of the cat. Am J Physiol 202:31-34
- Wang SC, Chai CY (1967) Central control of baroreceptor reflex mechanism. In: Kezdi P (ed) Baroreceptors and hypertension. Pergamon Press, Oxford, pp 117–130
- Ward DG, Baertschi AJ, Gann DS (1977) Neurones in medullary areas controlling ACTH: Atrial input and rostral projections. Am J Physiol 233:R116-126
- Warner HR, Cox A (1962) A mathematical model of heart rate control of sympathetic and vagus efferent information. J Appl Physiol 17:349–355
- Warzel H, Brattström A (1972) The effect of time of electrical stimulation of the carotid sinus on the amount of reduction in arterial pressure. Pfluegers Arch 337: 39-44
- Weidinger H, Hetzel R, Schaeffer H (1962) Aktionsströme in zentrifugalen vagalen Herznerven und deren Bedeutung für den Kreislauf. Pfluegers Arch 276:262–279
- Weiss GK, Crill WE (1969) Carotid sinus nerve; primary afferent depolarisation evoked by hypothalamic stimulation. Brain Res 16:269-272
- Weiss GK, Kastella KG (1972) Medullary single unit activity: response to periodic pressure changes in the carotid sinus. Proc Soc Exp Biol 141:314-317

- Werz M, Mengel E, Langhorst P (1974) Extracellular recordings of single neurons in the nucleus of the solitary tract. In: Umbach H, Koepchen HP (eds) Centralrhythmic and regulation. Hippokrates, Stuttgart, pp 259-265
- Widdicombe JG (1961) Action potentials in parasympathetic efferent fibres to the lungs of the cat. Arch exp Pathol Pharm 241:415-432
- Widdicombe JG (1966) Action potentials in parasympathetic and sympathetic fibres to trachea and lungs of dogs and cats. J Physiol (Lond) 186:56-88
- Wilson MF, Ninomiya I, Franz GN, Judy WV (1971) Hypothalamic stimulation and baroreceptor reflex interaction on renal nerve activity. Am J Physiol 221:1768– 1773
- Wurster RD (1977) Spinal sympathetic control of the heart. In: Randall WC (ed) Neural regulation of the heart. Oxford University Press, New York, pp 211-246
- Wurster RD, Trobiani S (1973) Effects of cervical sympathetic stimulation on carotid occlusion reflexes in cats. Am J Physiol 225:978–981
- Wyszogrodski I, Polosa C (1973) The inhibition of sympathetic preganglionic neurons by somatic afferents. Can J Physiol Pharmacol 51:29–38
- Yamashita H (1977) Effect of baro- and chemoreceptor activation on supraoptic nuclei neurons in the hypothalamus. Brain Res 126:551-556
- Ylitalo P, Karppanen H, Paasonen MK (1974) Is the area postrema a control centre of blood pressure? Nature 247:58-59
- Zanchetti A, Zoccolini A (1954) Autonomic hypothalamic outbursts excited by cerebellar stimulation. J Neurophysiol 17:475-483
- Zandberg P, Palkovits M, Jong W de (1977) The area postrema and control of arterial blood pressure; absence of hypertension after excision of the area postrema in rats. Pfluegers Arch 372:169-174

# Event-Related Slow (DC) Potentials in the Human Brain\*

MANFRED HAIDER \*\*, ELISABETH GROLL-KNAPP \*\*, and JOSEF A. GANGLBERGER \*\*\*

#### Contents

1	Introduction					
2	Terminology and Definitions 12					
3	Potential Characteristics and Methodology3.1The Biological Signal3.1.1Late Potentials3.1.2Contingent Potentials3.1.3Movement-Related Potentials3.2Reliability of Biological Signals3.3Variability of Biological Signals3.4Recording System3.4.1Characteristics of Electrodes for Registration of Slow (DC)	131 131 132 134 137 138 138				
	3.4.1       Characteristics of Electrodes for Registration of Slow (DC)         Potential Changes	138 139 139 140 141 141 144				
4	Electrogenesis and Physiological Significance4.1Event-Related Slow (DC) Potentials4.2Late Potentials4.3Contingent Potentials4.4Movement-Related Potentials	144 145 146 147 147				
5	Electrocorticography and Stereo-Electro-Encephalography5.1Electrocorticographical Studies5.1.1Late Potentials5.1.2Contingent Potentials5.1.3Movement-Related Potentials5.2Stereo-Electro-Encephalographic Studies5.2.1Late Potentials5.2.2Contingent Potentials5.2.3Movement-Related Potentials	148 148 149 149 150 151 152 154				

<sup>\*</sup> This research has been supported by the "Österreichischer Fonds zur Förderung der wissenschaftlichen Forschung".

<sup>\*\*</sup> Institut für Umwelthygiene der Universität Wien, Kinderspitalgasse 15, A-1095 Wien

<sup>\*\*\*</sup> Abteilung für funktionelle Neurochirurgie und klinische Neurophysiologie, Alserstr. 4, A-1090 Wien

6	Topographical Aspects (Scalp Distribution)6.1Late Potentials6.2Contingent Potentials6.3Movement-Related Potentials	157 157 159 160			
7	Maturational Aspects7.1Theoretical Problems of Developmental Studies7.2Late Potentials7.3Contingent Potentials7.4Movement-Related Potentials	161 161 162 163 166			
8	Psychological Correlates8.1Late Potentials as Information-Related Potentials8.2Contingent Potentials and Psychological Variables8.3Psychological Aspects of Movement-Related Potentials	167 167 170 173			
9	Psychiatric and Neurological Studies (Clinical Application)	174			
10	Drug Effects and Toxic Influences	178			
11	I       Concluding Remarks: a Hierarchical Model of Slow (DC) Potentials,         Activation and Behaviour       17				
Ref	ferences	183			

# Introduction

The study of event-related slow (DC) potentials of the human brain is at the same time fascinating and disappointing. It is fascinating because obvious correlations exist between these electrical phenomena of the brain with higher psychic functions and processes. But it is disappointing because these correlations remain "phenomenological", and little is known about the actual underlying brain mechanisms. Historically, *Caton* (1875) first observed event-related slow (DC) potentials in response to sensory stimulation and motor acts. Later studies ignored DC potentials due to the use of low frequency filters. Renewed interest in these problems started with *Davis* et al. (1939), who tried to correlate slow (DC) potentials with variations in depth of sleep. *Goldring* and *O'Leary* (1951a) studied stimulus-evoked slow (DC) potentials, and *Köhler* and *Held* (1949) and *Köhler* et al. (1952) demonstrated slow (DC) potentials connected with complex visual stimuli.

Event-related slow (DC) potentials occurring after external stimulation as well as prior to behavioural responses have been extensively described by *Caspers* (1961, 1963, 1965). DC potentials related to mental activity were found by *Bechtereva* (1967, 1968, 1974) in different subcortical structures of the brain. Slow potentials related to conditioned responses have been observed by *Rowland* (1960, 1967, 1968), and sound-evoked slow negative potentials were recorded from auditory cortex by *Gumnit* (1960, 1961). Late responses in the form of long latency, slow evoked potentials have been described as being associated with "information delivery" (*Sutton* et al. 1965) and with unpredictably occurring stimuli as a kind of central orienting response (*Haider* et al. 1968b; *Ritter* et al. 1968). These late responses have been variously designated "P 3" or "P 300" (*Ritter* et al. 1968); "late positive component" (LPC) (*Ritter* and Vaughan 1969) and "association cortex potential" (ACP) (Vaughan 1974).

Contingent responses have been observed as slow (DC) potential changes between a warning or conditional stimulus  $(S_1)$  and an imperative or unconditional one  $(S_2)$  with which the subject is "engaged" in the form of action or decision. *Walter* et al. (1964), in such an experimental design, observed a slow increase in surface negativity and called it "contingent negative variation" (CNV) or "expectancy wave" (E-wave).

Slow (DC) potentials preceding a voluntary movement have been observed by *Kornhuber* and *Deecke* (1965), who called this slow potential shift *Bereitschaftspotential* (BP) or "readiness potential" (RP). The potential changes associated with motor action have been named "motor potential" (MP) by *Gilden* et al. (1966) and *Vaughan* et al. (1968).

Our review deals with these three groups of phenomena, namely the late potentials associated with orienting and information processing, the contingent potentials associated with expectancy and the movement-related potentials associated with voluntary movement and motor control. First we will survey the terminologies and abbreviations which unfortunately are used very inconsistently in this field. Then we will describe some methodological aspects, which seem to be necessary since different methods for registration and analysis have been proposed. After comments on electrogenesis and physiological significance we will describe results of research with corticography and stereo-electro-encephalography. The later parts of the review will then be devoted to topographical and maturational aspects and related psychological processes, as well as the application in clinical studies and research on drug effects and toxic influences.

# 2 Terminology and Definitions

Concerning terminology, abbreviations and definitions we will try to work out some unifying principles and recommendations for further usage. A basic discrepancy already exists between researchers recording directly from the cortex, working mainly on the animal brain, and researchers recording from the human scalp. The first group has decided that the term DC potentials, derived from the application of directly coupled amplifiers, would be most unprejudiced and that this nomenclature should be adopted in future publications (*Caspers* 1974).

The second group, in the title of its congress papers, uses the term "event-related slow potentials" (*McCallum* and *Knott* 1973).

In our title we have combined both terminologies by adding DC in the sense of direct coupling, in parantheses, and also by using the term "event-related slow (DC) potentials". This combined term has already been used by *Lindsley* (1969). A general review of the different terminologies and abbreviations used in this field is given in Tables 1 and 2.

	Descriptive terms related to		
Technological terms	Potential characteristics	Events	
DC (directly coupled)	SP (slow potentials)	ERP (event-related potentials)	
DC (direct current)	SPC (slow potential change)		
DC recordings	SPS (slow potential shift)		
	DC change		
	DC shift		
	Standing potential		
	Steady potential		
	Sustained potential		

Table 1. General terminology and abbreviations for event-related slow (DC) potentials<sup>a</sup>

<sup>a</sup> Terms used as main headings in this paper and recommended for future usage are enclosed.

At the time terminologies must remain largely descriptive and not explanatory. We therefore use as main headings in this review and propose for further usage the general descriptive terms "slow potentials" (SP) and "event-related potentials" (ERP) and as specific descriptive terms, "late potential" (LP) or "information-related potential" (IRP), "contingent potential" (CP) and "movement-related potential" (MRP).

We are not concerned in this review with the early and middle components of evoked potentials. Late potentials (LP) may include positive as well as negative potentials between 200 and 500 ms, depending on the experimental situation. The best usage seems to us therefore to designate the main component of the late responses according to its polarity and latency. In many cases this will be a positive wave with a peak latency around 300 ms, as originally described by *Sutton* et al. (1965) and designated as P 3 or P 300 by *Ritter* et al. (1968) or LPC (late positive component) by

Phenomenological terms	Topographical terms	Descriptiv Potential characteristics	re terms related to Events
OP (orienting potential)		LP (late potential)	IRP (information- related potential)
	ACP (associa- tion cortex potential)	LPC (late positive component)	PWSP (post warning signal positivity)
	F	SNP (slow negative potential)	
		$P_3 = P_{300}$ (positive wave with peak latency 300 ms)	
E-wave (expectancy wave)			CP (contingent potential)
Imaginary potential			CNV (contingent negative variation)
Imagination potential			RCPV (rewarding contin- gent positive variation)
			PINV (post imperative negative variation)
I-wave (intention wave)			MRP (movement- related potential)
			RRP (response-related potential)
			AMP (average movement potential)
			RP (readiness potential, i.e. BP, Bereitschafts- potential)
			PMP (premotor positive potential)
			MP (motor potential)
			GDMP (goal-directed movement potential)

Table 2. Situation-specific terms and abbreviations for event-related slow (DC) potentials  $^{\rm a}$ 

<sup>&</sup>lt;sup>a</sup> Terms used as main headings in this paper and recommended for future usage are enclosed.

*Ritter* and *Vaughan* (1969). In other cases the main potential components may be a negative wave with latencies of about 250 ms, as described by *Haider* et al. (1968b). In other studies, still other prominent waves or wave complexes may be seen which could all be summed up under the descriptive heading of "late potentials". Since all components seem to be related to different aspects of information processing we propose the term "information-related potential" (IRP).

Similarly, the term "contingent potential" (CP) could be used for a whole group of responses described by *Walter* (1975) as "potential changes whose appearance is contingent on the association of signals and/or action or decision by the subject." The most obvious of these is the contingent negative variation (CNV). Again, this more specific term applies to a special situation in which a negative potential appears.

But sometimes the variation may be positive. *Marczynski* (1972) described a reward-contingent positive variation (RCPV). In some parts of the brain (frontal of the thalamus) even the typical experimental situation for a CNV leads to slow positive variations (*McCallum* et al. 1976).

Under the heading "movement related potentials" (MRP), at least three main components are described: a slow rising readiness potential (Bereit-schaftspotential), premovement phasic components and a late positive wave. Recently, Zielbewegungspotentiale (goal-directed movement potentials, GDMP) have been found over the precentral and parietal cortex (Grünewald-Zuberbier et al. 1978a; Grünewald-Zuberbier and Grünewald 1978). In these instances the premotion negativity increases and remains during the goal-directed movement until the target is reached.

It must be emphasized that some authors (Gilden et al. 1966; Vaughan et al. 1968) use the term "motor potential" for all potential components associated with movements, whereas other authors (Kornhuber and Deecke 1965; Deecke et al. 1969) call only the negative peak "motor potential". There is no agreement as to what part of the trace the term  $N_2$  applies and to what part, the term "motor potential". Gerbrandt et al. (1973) use the term "average movement potential". A group attending the symposium on motor control [Fourth International Congress on Event-Related Slow Potentials, Otto (ed.) 1978] therefore proposed the following taxonomy of movement-related brain macropotentials (Papakostopoulos, 1978).

- 1. Potentials related to the state of the sensory motor system at rest (resting potentials – RTPs)
- 2. Potentials related to functions during the preparatory period (preparatory period (preparatory potentials PPs)
- 3. Potentials related to the initiation period (initiation potentials IPs)
- 4. Potentials related to the execution of the movement (movement-execution potentials MEPs)

5. Potentials related to the termination of the movement (termination potentials - TPs)

Future research must show the extent to which these terms are useful in handling our knowledge of motor control from different disciplines. In this review we take the term "movement-related potentials" as the best descriptive term and propose it for further usage. How far the different components of these movement-related potentials, according to their time sequence, are related to preparation, initiation, execution and termination of movements will be discussed in this review.

#### 3 Potential Characteristics and Methodology

3.1 The Biological Signal

#### 3.1.1 Late Potentials

Sutton et al. (1965) described a rather slow late positive potential component with a peak latency of about 300 ms which appeared to be related more to complex psychological variables than to the physical characteristics of the stimulus. Further experiments showed that this phenomenon of late potential changes with peak latencies between  $\sim 200$  and  $\sim 400$  ms seem to be related to similar psychological variables (*Haider* et al. 1968b; *Ritter* et al. 1968). Multiple late potential components have also been described (*Harter* and *Salomon* 1972; *Squires* et al. 1975; *Courchesne* et al. 1975). Some characteristic wave forms of late potentials, drawn according to the published curves of different authors, are compared in Fig. 1.

There have also been reports of cerebral potentials occurring at about the time of an expected but absent stimulus. Stimulus absence provided information or is salient for the subject. These potentials consist mainly of a positive peak occurring about the same time as the p 300 and is referred to as an "emitted p 300" potential (*Klinke* et al. 1968; *Weinberg* et al. 1974; *Ruchkin* et al. 1975).

The original construct that one unifying psychological variable is responsible for these potential changes which are elicited in a wide variety of situations such as uncertainty, information delivery, signal significance, orienting, inhibition, selective recognition, awareness or salience, has been replaced by some authors with the concept of several independent late potential components. This second concept is supported mainly by topographical data.

The technical problems involved in defining LP on the basis of individual trials are considerable (see Sect. 3.6.2) and sometimes impossible (low signal-to-noise ratio).



Fig. 1 A-D. Late potentials found by different authors for unexpected stimuli compared with expected stimuli. — unexpected stimuli; ---- expected stimuli negativity upwards; ↑ stimulus onset. A Ritter et al. (1968); B Haider et al. (1968b); C Squires et al. (1973); D Courchesne et al. (1975)

Late potentials normally are extracted from the raw EEG by different methods of averaging (methods that increase the signal-to-noise ratio).

#### 3.1.2 Contingent Potentials

The contingent negative variation derives its name from being a slow potential shift in the EEG baseline which typically occurs on the association (contingency) of successive stimuli. As mentioned previously, it was first described by *Walter* et al. (1964), as developing between the warning signal and the imperative signal in a constant foreperiod reaction time task. The contingent negative variation appears within the intersignal interval as a surface negative shift with respect to a mastoid reference.

Meanwhile, it has been demonstrated that the CP can be elicited by many different experimental designs. It is not dependent on stimulus



Fig. 2. Contingent negative variation (CNV) in the standard paradigm (reaction time experiment with foreperiod) and the effect of time constant on the contingent variation. Note also the apparent variations in latency of the first peak of CNV. A warning signal  $(S_1)$  is followed by repetitive clicks which were terminated by the press of a button. Each trace is the average of 50 trials recorded simultaneously but with different time constants. Negativity upwards. (Cooper et al. 1974)

modality or intensity, nor are overt motor acts essential for a contingent variation. The reasons for the sometimes considerable changes in CP morphology and amplitude as well as for changes in peak latency variations are discussed in Sect. 8.

The CP is not readily seen in the raw EEG traces of most normal adults. Therefore, different averaging techniques are used to enhance CP amplitude relative to background EEG. In the last years some mathematical models have also been discussed for CP evaluation as alternatives to the averaging methods, which include some theoretical difficulties.

The maximum negative voltage, compared to the iso-electric direct current prestimulus baseline, ranges from  $10 \,\mu V$  to  $50 \,\mu V$  with an approximate mean of  $20 \,\mu V$  (*Walter* 1967b). The termination of the CP can be slow and inconsistent as well as immediate.

The terminating potential decline limb of the CP may overshoot the baseline. The time course for the development of CP may vary from 0.5 s to many seconds. It is reported to persist as long as 20 s under special conditions, if the subject is highly motivated. When using RC-coupled amplifiers the time constant may reduce the amplitude of slow potentials. This is demonstrated in Fig. 2, which presents a typical example of a CP simultaneously recorded with different time constants in a classic experimental paradigm.

# 3.1.3 Movement-Related Potentials

Voluntary movements are preceded and followed by slow electrical brain potentials of different latency and amplitude characteristics. There is general agreement as to the morphology of this MRP and its division in four components: a slowly rising negativity  $(N_1)$ , an inconsistently appearing small positivity  $(P_1)$ , a fast negative deflection  $(N_2)$  and a large, rather slow final positivity  $(P_2)$ .

Characteristic examples of movement-related potentials for right and left unilateral finger movements are shown in Fig. 3, which demonstrates that the fast negative deflection (motor potential, according to *Deecke* and *Kornhuber* 1977) occurs contralateral to the moving hand.

The N<sub>1</sub> component, generally called *Bereitschaftspotential* (BP), is a bilateral, slowly increasing surface negative potential. In scalp derivations it starts on the average about 0.8 s prior to the onset of a movement. The amplitude of the average BP is between 5 and 7  $\mu$ V. There is small intraindividual but large interindividual variability in latency. Amplitudes are dependent on several individual and experimental parameters.

Similar to the CP the BP occurs in preparatory situations. Therefore, the question sometimes arose whether CP and BP would not be the same electrophysiological phenomenon. There is now, however, general agree-



Fig. 3. Examples of voluntary movement-related potentials (*Deecke* et al. 1976b). Comparison of right unilateral (R) and left unilateral (L) index finger movements in the same experiment. With right index finger movement there is more negativity over the left precentral region (R); with left index finger movement there is more negativity in the right precentral lead (L). Bipolar recording (left versus right precentral) in the bottom trace

ment that CP and BP are correlates of different psychophysiological mechanisms. *Deecke* and *Kornhuber* (1977) ascribe different topographical distribution, morphology and amplitudes to CP and BP. Similar differences between BP and CP were described by *McCallum* (1978) for cortical derivations.

A small positive-going wave whose deflection time is 90-80 ms before the start of movement as defined by EMG onset is sometimes registered. Average amplitude of this wave is about 1.7  $\mu$ V. This P<sub>1</sub> component appears to be bilateral and widespread and is described as occurring inconsistently across and within subjects (*Gerbrandt* et al. 1973; *Vaughan* et al. 1968). *Deecke* and *Kornhuber* (1977) found this premotor positivity in about two-thirds of their subjects. *McCallum* (1978) was not able to analyse this potential component.

A fast negative deflection following the  $P_1$  component was described by *Kornhuber* and *Deecke* (1965) and is called  $N_2$  or motor potential (MP). The negative MP is restricted to the contralateral precentral area. It starts 60–50 ms prior to EMG onset and has an average amplitude of about 1.6  $\mu$ V. The MP starts before the onset of movement. The N<sub>2</sub> component described by *Deecke* and *Kornhuber* does not correspond to the N<sub>2</sub> component described by *Gerbrandt* et al. (1973), who found that this component arises after movement onset. This difference obviously depends on electrode linkage and position.

In cortical derivations during stereotactic surgery the motor potential has now been observed as a restricted, localized phenomenon at the proper motor cortex, area 4 (*Ganglberger* et al. 1980).

During smooth goal-directed hand movements the premotion negativity increases and remains during movement until the target is reached. An example is shown in Fig. 4.



Fig. 4. Negative potentials during goal-directed movements, Zielbewegungspotential  $(B \mod)$  reach double amplitudes of Bereitschaftspotential before movement  $(A \bigsqcup)$ . In B the right hand guides the rod to the visualized goal during maintained target  $\leftarrow$  fixation. Reference: linked mastoids. (Grünewald-Zuberbier et al. 1978a)

The negativity is very large and widespread and has maximal amplitudes over the vertex. Over the precentral cortex they are larger contralaterally, but over parietal cortex an ipsilateral preponderance is sometimes seen.

Potential changes during such goal-directed movements are of higher amplitude than potential changes during voluntary finger movements and very often may be seen in the raw EEG. *Grünewald-Zuberbier* et al. (1978a), *Grünewald-Zuberbier* and *Grünewald* (1978) were the first to describe the phenomenon under the term "goal-directed movement potential".

### 3.2 Reliability of Biological Signals

The reliability of a measurement is a crucial factor in determining its usefulness. Generally the reliability is expressed as the correlation between repeated measurements. Information on test-retest reliability (TRR) of averaged slow potential changes is limited, and the results of different investigations may not directly be compared. Different authors use various experimental designs and parameters for potential descriptions. This probably accounts for some of the diverging results.

The test-retest reliability of different EP components is reported to be between 0.87 and 0.97 (*Shagass* and *Schwartz* 1961; *Callaway* et al. 1965; *Kooi* and *Bagchi* 1964; *Ellingson* et al. 1973; *Soskis* and *Shagass* 1974).

Smaller reliability coefficients were found by *Roth* et al. (1975) (r = 0.78 amplitude; latencies 0.63). For longer test intervals smaller correlation coefficients in the range 0.55-0.66 were found. There is general agreement on higher reliability for earlier potential components than for the later ones. There is so far no agreement whether latency measurements or amplitude measurements are more reliable (*Kooi* and *Bagchi* 1964; *Roth* et al. 1975).

There are only few experimental data on CP reliability. *Straumanis* et al. (1969) found a rather low reliability of 0.32 for a normal population and 0.05 in a psychiatric population. It has to be mentioned, however, that different experimental designs were used as test and retest.

Cohen (1969) reports a reliability coefficient of 0.80 for "average maximum vertex amplitude", using test intervals of 2-8 days. The median subject consistency in retests separated from 5 min to 8 days was 0.68 as reported by *Roth* et al. (1975).

*Timsit-Berthier* et al. (1978) studied CP reliability using two groups of psychiatric patients (those with permanent behaviour disorders and those with important changes in their clinical state) and retest intervals of 1-2 months and 1-2 years. Correlation coefficients for amplitude measures were 0.66 (1-2 months) and 0.55 (1-2 years), respectively (P < 0.05). For qualitative evaluation of CNV termination, retest reliability was even higher (0.88-0.78).

Tests for personality traits which by definition are stable over time have TRRs ranging between 0.9 and 0.6 for intervals of several weeks. The CP reliability is therefore lower than expected for measurement of trait reliability but much higher than measurements of individual variables fluctuating with different states of the individual.

#### 3.3 Variability of Biological Signals

The interindividual variability of the EP component shows a slight but progressive increase for waves I–III and is very high for the late potential components. This occurs in the peak latencies as well as in amplitude measurements (*Storm van Leeuwen* et al. 1975).

There is considerable interindividual variability in phenomena related to contingent potential development. In raw EEG traces, CP's are to be seen in about 20% of subjects. The range of CP amplitude found among normal individuals is between 8 and 50  $\mu$ V in standard CP paradigms.

Changes in the experimental design (i.e. omission of  $S_2$ , distraction from  $S_2$ ) may evoke different reactions in different subjects, i.e. decrease in CP amplitude for some subjects and augmentation for others.

In summary, the interindividual variability of CP parameters in standard situations is already great. Introduction of supplementary behavioural aspects into the situation increases the interindividual variability considerably.

Average EPs in the same subject under identical conditions are very stable over a long period of time: the intra-individual variability is small. Studies of *Aunon* and *Cantor* (1977) have shown that the intrasession variability for EP components is significantly smaller than the intersession variability within the same subject. Interlaboratory standard deviation was found to be over twice as large as the intralaboratory standard deviation measured over an 8-week period.

#### 3.4 Recording System

### 3.4.1 Characteristics of Electrodes for Registration of Slow (DC) Potential Changes

Reversible electrodes should be used for registration of steady or slowly changing potential differences; i.e. electrodes having low values for  $R_F$ , the faradic resistance of the chemical change and the Warburg impedance. Using reversible electrodes a small increase or decrease of potential above or below the electrode potential will cause the discharge of ions at a considerable rate. This means that such electrodes have a low resistance to steady or slowly changing potentials.

Reversible electrodes commonly used in EEG investigations are reversible with respect to anions. In this case a metal is in contact with one of its insoluble salts immersed in a solution of a soluble salt of the same anion (*Geddes* 1972; *Cooper* et al. 1974).

The choice of the metal used for electrodes cannot only be determined by the susceptibility to polarization. Many metals cause inflammation when they are in contact with tissue for longer periods. Therefore, especially for chronical intracerebral recordings, metals such as stainless steel or gold must be used whose recording characteristics are poor when DC recordings have to be made and conventional EEG amplifiers are used. The low frequency activity is attenuated by a factor determined by the values of the electrode impedance and the amplifier impedance. EEG amplifiers with high input impedance (100 M $\Omega$ ) make possible the registration of slow potential changes with such electrodes without too much loss. In animal experiments, electrolyte-filled glass pipettes in connection with sintered Ag/AgCl pellets seem to be a good alternative (*Rebert* and *Irwin* 1973).

Electrode potentials often change with time. Slow drifts may be caused by several sources; for instance, differences between electrodes, skin potentials or potential changes at the skin as a result of the conducting jelly can be in the range of mV and therefore much higher than the EEG phenomenon under investigation. Because of these instabilities one should carefully consider whether DC recordings with all their difficult technical aspects are really required or whether AC amplifiers with very long time constants (25 s) will suffice.

# 3.4.2 Number and Location of Electrodes

There is general agreement that as many electrodes as possible should be used in EP experiments. The number and location of electrodes must be decided according to the specific goal of the experiment and the technical availabilities of the recording system. Further limitations are set by the capacity of the data-analysing equipment. For experiments dealing with the relationship between EPs and complex psychological processes, the following ranking of electrode placements in order of desirability was proposed by *Donchin* et al. (1977) if scalp electrodes are used. Electro-oculogram EOG and the encephalogram from the location  $C_Z$  should always be recorded followed by derivations from  $P_3$  and  $P_4$  ( $P_Z$ ),  $C_3$ ,  $C_4$ , then  $F_3$ ,  $F_4$  (or  $F_Z$ ),  $O_1$  and  $O_2$  and finally,  $T_5$  and  $T_6$ . For specification of electrode placement the international 10–20 electrode placement system (*Jasper* 1958) should be used. Electrode positions which vary for scientific reasons from the 10–20 system should also be specified in terms of the standard coordinate system.

# 3.4.3 Recording Montages

For deriving EEG recordings from an electrode array, three main methods are described:

1. In the bipolar method potential differences are measured between two electrodes, both of which are affected by EEG potentials ("active" electrodes).

- 2. In a common reference (unipolar, monopolar) derivation each amplifier is situated between one "active" electrode and the so-called common reference which is placed so as to minimize the possibility of picking up brain potentials ("inactive" electrode).
- 3. In a common average reference system (Goldman-Offner method) usually all the electrodes on the scalp (sometimes with the exclusion of some electrodes thought likely to produce artefacts) are connected through equal resistors to a single point which is then used as a common reference. The potential of this point will be the average of the potentials at the scalp electrodes. Each amplifier is then connected between one active electrode and this common average "inactive" reference).

It is not possible to make a general recommendation as to which recording method is preferable. In investigations of slow potential changes there seems to be, however, a preference for common reference derivations using "linked ears" as the "inactive" electrode site.

#### 3.5 Artefacts

"By definition an artefact in electroencephalography is any recorded electrical potential which does not originate in the brain" (*Cooper* et al. 1974). Artefacts arise because of two main factors: technical and physiological. Many technical artefacts arise at the junction between the electrodes and the patient's scalp. Movement of the electrode relative to the scalp results in a change of contact resistance or disturbance of the electrode potential. Imperfect electrode contact or contamination of the different junctions (electrode clipjacket) may cause further artefacts.

Several physiological potentials originating from extracerebral sources may be recorded from the scalp electrodes used to detect the EEG and the time-locked cerebral potential it contains. Some of them, such as muscle activity or artefacts from the EEG, are rather easily recognized in the raw EEG. Galvanic skin response and respiratory effects may seriously confound the activity originating from the brain, especially when event-related slow (DC) potential shifts are under investigation.

Low et al. (1966a) reported that a consistent pattern of involuntary eye movements could accompany behavioural tasks used to elicit the CNV or other CP.

The most serious difficulty, however, is undoubtedly presented by the corneoretinal potential which generates the EOG. The fact that the EOG is sometimes time-locked to stimuli hasled to serious errors, especially in slow potential investigations. This EOG potential is large in comparison to cerebral potentials and may significantly contaminate recordings taken from points as distant as the vertex and sometimes the occiput (*Rowland* 1968).

Different methods for EOG elimination have been proposed:

- 1. Requiring the subject to fix his gaze during the recording period is reported to be effective in about 50% of subjects (*Rowland* 1968; *Wasman* et al. 1970); *Hillyard* and *Galambos* (1970) found this method "generally" effective.
- 2. A variation of the gaze-fixation technique using a special device for stimulus presentation has been shown to virtually eliminate involuntary ocular rotations (*Borda* and *Hablitz* 1973).
- 3. Techniques which favour rejection of eye-movement contaminated trials are:
  - a) Electrographical monitoring of eye-movements and subsequent elimination from further analysis of trials during which eye movements occurred, based on either visual monitoring or automatic EOG recognition in off-line analysis (*Debecker* and *Carmeliet* 1974) with analogue equipment or on-line by a general purpose computer.
  - b) Rejection of mechanically detected eye-movement contaminated trials in on-line analysis using a mechanical transducer to sense movements of the eye equivalent to 3° or more has been suggested by *Papakostopoulos* et al. (1973).

The behavioural significance of such "cleaned" CPs must be discussed as it is very likely that in some situations CP trials accompanied by eye movements represent a special situation.

- 4. Mathematical adjustment of CNV amplitude based on the correlation between magnitude of eye movements and magnitude of "pseudo-CNV" recorded on the vertex (*Hillyard* and *Galambos* 1970).
- 5. On-line technique for separating eye movement potentials from EEG potentials by electronically subtracting a fraction of both the horizontal and the vertical EOG potential from the raw EEG (*Girton* and *Kamiya* 1973).

# 3.6 Data Analysis

# 3.6.1 Averaging and its Limitations

Since *Adrian* (1941) demonstrated that an evoked response to a flash in the ongoing activity of the EEG was tied in time to the stimulus, with latencies of about 100 ms one assumed that earlier signal-evoked events were also almost certainly present though masked by the ongoing EEG. The problem of selecting these evoked responses from the ongoing EEG is one of increasing the signal-to-noise ratio. A technique was consequently developed by which the recorded potential change is time locked to a pulse coincident with the stimulus. Repetition of the same signal situation and averaging the resultant record was, for a long time, the only method of enhancing the signal-to-noise ratio.

Different parameters of these averaged evoked responses are used for descriptive and statistical purposes. The parameters most often used in slow potential studies shall be discussed.

One way to define the size of LP components, contingent variation or movement-related potentials, is the measurement of amplitude. There is presently no general agreement as to whether the amplitude of EP components should be measured from a baseline or as an absolute distance between peaks of adjacent components (peak-to-peak measurement). Measurements from the baseline present some difficulties. First, one must define the baseline and, second, measurements from the baseline introduce special complexities in the interpretation of data, for example when negative-going potentials peak at levels more positive than the baselines.

When using peak-to-peak amplitude measures one need not be concerned by the fact that there are sometimes difficulties in determining a baseline and also that the baseline and the whole EP may be superimposed on a (very) slow wave. However, measuring peak-to-peak does not take into account that adjacent components may be differently affected by the introduced variables. Thus, those measurements may blur the true relationship.

Investigations of late potential components generally refer to a baseline prior to stimulus onset (5%-10%) of total sweep time is recommended). In both baseline measures and peak-to-peak measures, the points for amplitude evaluations are determined by the peak latencies and the number of late potential components.

In CP studies baseline considerations are dependent upon the general definition of the contingent variation which can be either (a) the absolute level of negativity/positivity, measured in relation to an absolute baseline (in this case prestimulus baseline is used); or, alternatively (b) a potential change with respect to the level upon which the warning stimulus falls. Here baseline would be adjusted from a point 400 ms following the first stimulus (*Rebert* and *Knott* 1970).

The difference between these two definitions of CP becomes important whenever the evoked response to the first stimulus is associated with a pronounced positive shift, followed by a small negative-going potential shift which in fact may never exceed the absolute baseline (i.e. the stimulus baseline).

The use of the late positive-going potential shift of the first signal for baseline evaluation or peak-to-peak measurements would imply that this component is not affected by the same variables as the contingent variation. Some investigations, however, have shown that such activity evoked by the first stimulus has considerable effect on the CNV (*Otto* and *Leifer* 1973).
There are two different ways for amplitude evaluation commonly used in contingent variation studies. Some workers measure amplitude just before or at the time of the presentation of the imperative stimulus (*McCallum* and *Walter* 1968), whereas others use maximum amplitude wherever it happens to occur in the interstimulus interval.

Further amplitude measures used in most CNV studies are the values for the post-warning signal positivity measured either in relation to absolute baseline or as peak-to-peak measure in relation to previous negative potential peak.

Amplitude measures of movement-related potential changes refer to a prepotential baseline and also to peak-to-peak measures. Because the onset time of the slowly developing *Bereitschaftspotential* (readiness potential) changes according to different parameters of the performed movement, no general assumption can be made for the length of the necessary premovement registration time. (In experiments using the general experimental designs of voluntary finger movement 2-s premovement and 1-s postmovement registration should be enough for baseline evaluation and measurement of slow potential components.)

No general solution can be advanced when amplitude measures are of concern. Each experiment, however, should examine these data both as measured from the baseline prior to stimulus onset and also from "peak-to-peak".

Low and McSherry (1968) proposed to use the integration of the area above or under the baseline between arbitrarily defined points as a measurement of the size of potential components. This method should only be used when digital computation is available. There are some typical integration limits, generally used by experimentors, which correspond in some way to the most frequently used points of amplitude measurement. Because less data are discarded by the use of area measures than by amplitude measurement, this method offers some advantages.

Neither amplitude nor area measures are able to take the shape of the contingent variation or movement-related potentials. As a matter of fact a great variety of wave shapes exists. For the CNV, *Tecce* (1972) has discussed at least three major types, each showing a large individual variability.

Some evidence, however, is emerging that the slope of the ascending limb might be equally as important as other CNV measures. In this connection the use of "latency to peak" or some other rise time measures could be helpful. Attempts to relate the cut-off or descending part of the curve with diagnostic measures have already been successful, and it will be necessary to perform further work on these problems.

The concept of signal averaging implies that a signal must be extracted which may not be observed in its entirety in the raw data. It implies at the same time that each reaction in the nervous system in response to the same stimulus should be the same. This assumption is certainly not true, since the stimuli occur at different times and are therefore presented to a different state of the nervous system. Different reactions should result. For some parameters, systematic changes during successive presentations of the same event are known. In those cases, variability may be thought of as a dependent factor.

The most important limitation of signal averaging is the inability of this method to give specific information about the specific influence of the single trials on the central tendency both with respect to amplitude and latency differences which may exist.

Knowledge of the consistency of potential changes is necessary, however, to any statement of reliability. Therefore measures of variability are now often used as routine qualification of the average. The most commonly used measure is "variability of voltage at the same time". But as long as those measurements refer to the mean or the sum of potentials, the variability in the amplitude of the signal may not be distinguishable from variability in latencies.

#### 3.6.2 Alternatives to Averaging

One alternative to signal averaging is the method of superimposition, which assumes that the same variability would emerge with an equal number of randomly selected single trials. Superimposition of single trials, on the other hand, does not yield results if the signal-to-noise ratio is low. In both, the eye is used as a device for estimating mean and variability.

Discriminant and factor analysis techniques were proposed. These methods are based on the same assumption as average methods, namely that single trials represent variability in respect to a "real signal".

Fourier analysis has also been used for the analysis of averages as well as for that of raw data. Fourier analysis gives very little information about the time when the signal is occurring, but this difficulty may be partially solved by using successively smaller epochs within the record of interest.

#### 4 Electrogenesis and Physiological Significance

The discussion of electrogenesis and physiological significance will be kept brief as *Caspers* and *Speckmann* will give an extensive account in one of the following volumes of "Reviews of Physiology". At present such a discussion can be only hypothetical. The main problem is to bridge the gap between our knowledge of sensory, neuron and membrane physiology as well as neurochemical mechanisms on the one hand and the slow DC potential phenomena on the other hand.

## 4.1 Even-Related Slow (DC) Potentials

Slow surface potentials of evoked responses are assumed by some authors (Watanabe et al. 1966; Creutzfeld and Kuhnt 1966, 1967) to reflect the summated postsynaptic potentials of cortical cells as well as the potentials originating in synchronous afferent and efferent fibre activity. Expanding this theory to cover slow DC potentials McSherry (1973) assumes a combination of prolonged superficial axodendritic excitatory postsynaptic potential volleys and prolonged deep axosomatic inhibitory postsynaptic potential volleys, and he presumes distant cortical and subcortical centres to supply the steady barrage of inhibiting and excitatory input. In accordance with this view, McSherry and Borda (1973), in a study using monkeys, found intracortical, positive correlates to the slow rising surface negativity. *Rebert* (1973) described different active and inactive brain regions during the training of monkeys on a reaction time task. Some stereotactic studies using humans demonstrated the involvement of different subcortical brain structures in the genesis of slow DC potentials (*Haider* et al. 1968a, 1972; Groll-Knapp et al. 1968, 1977; McCallum et al. 1976).

Other authors stress the possibility that neurons and glia may contribute to slow DC potentials. *Goldring* (1974) concludes that evoked DC shifts will reflect either sustained hyperpolarization of neurons in cortical depth, predominantly glial depolarization, or a blend of both neuronal and glial depolarization.

Since gross recordings of slow DC potentials probably reflect the average potential fluctuations in a large number of individual units, investigations of the origin of DC potentials have focussed on special experimental conditions, such as seizure activity, in which the majority of individual generators are forced into synchronized action. Studies with such models have shown that DC changes are paralleled by membrane-potential changes of single neurons and of glia cells (*Karahashi* and *Goldring* 1966; *Glötzner* and *Grüsser* 1968; *Grossman* and *Hampton* 1968; *Caspers* and *Speckmann* 1970; *Speckmann* et al. 1972). Since *Kuffler* and *Nichols* (1966) pointed out that glial depolarization will occur due to local release of potassium as a result of neural activity, it seems possible that a linkage between neuronal and glial activity represents a complex generation of slow DC potentials. The extent to which such models may be applied to late responses, contingent responses and movement-related potentials awaits further study.

The possibility of an involvement of cholinergic mechanisms in the genesis of slow DC potentials is suggested by laminar DC studies of acetyl-

choline-activated epileptiform discharges in cerebral cortex (*Ferguson* and *Jasper* 1971) and the cholinergic influences on reward-contingent positive variations (*Marczynski* et al. 1971). Obviously many questions with respect to neurochemical mechanisms and their association with slow DC potentials in different brain regions need further explication. Because an "ascending cholinergic reticular system" from the brain stem, with dorsal and ventral tegmental pathways extending to the anterior thalamus and septal regions with important projections to the limbic system and cerebral cortex has been described (*Shute* and *Lewis* 1967; *Lewis* and *Shute* 1967), it may be speculated that thalamic slow DC potentials as observed by *Haider* et al. (1968a, 1972), *Rebert* (1972), *McCallum* et al. (1976) and *Groll-Knapp* et al. (1977) may also be cholinergically mediated.

It should be mentioned that some authors (*Wurtz* and *O'Flaherty* 1967; *Besson* et al. 1970) assume a vascular origin for DC potential shifts related to sleep, arousal and drug influences. The time course of vascular changes is however much too slow to account for the event-related slow DC potentials discussed herein.

### 4.2 Late Potentials

Assumptions about the physiological significance of late potentials are complicated by the fact that different experimental situations lead to different topographical distributions of these responses. Therefore it cannot easily be concluded that they all depend on an unitary physiological mechanism arising from a single cerebral generator system. Some authors (Waszak and Obrist 1969; Karlin et al. 1970) noted an augmentation of localized late responses in situations with an inhibitory context. Papakostopoulos et al. (1976) and McCallum et al. (1976) explain these findings as being due to an increased spatial distribution in the cortex, resulting in an augmentation of amplitude when recorded from the scalp, and they interpret late responses as electrophysiologically indicating an inhibitory brain function. These authors also find a spread of late responses generally located parietally to prefrontal cortex in situations demanding alterations of a preset plan and discuss this as being compatible with frontal lobe functions and with the special perceptual motor deficits after frontal lobe damage. Desmeth and Debecker (1979a, b) interpret the P<sub>350</sub> as a postdecision event resulting from a phasic inhibition exerted by prefrontal cortex on the mesencephalic reticular formation at the closure of a cognitive processing epoch.

### 4.3 Contingent Potentials

In speculating about the physiological significance of contingent responses, Walter (1975) suggests that the underlying electrochemical processes may act as cortical "primers", ensuring prompt and economical firing of the central effector mechanisms. He mentions that such slow DC potential phenomena arise in brain regions closely connected to the motor area and that the amplitude of such potentials is related inversely to motor reaction times. During the 200–300 ms before the imperative stimulus, when the contingent response is at its peak, the amplitude of responses to intracerebral electrical stimuli is 30% larger than at other times, the latency of responses to interpolated sensory stimuli is slightly shorter at this phase and the threshold for the H-reflex is lower (*Papakostopoulos* et al. 1973).

Concerning the connection of contingent responses and the energy exchange system of the brain, *Walter* (1975) maintains that a local rise in available oxygen starts about 10 s after the response and peaks at about 20 s. Such local variations in oxygen availability may be measured with noble metal electrodes (*Cooper* et al. 1966). On the hypothesis that they are due to changes in blood flow and that this is mediated by variations in  $CO_2$  as a result of glycolysis, it is supposed that the electrical phenomena of contingent responses are associated with local increases in brain metabolism.

### 4.4 Movement-Related Potentials

The physiological significance of movement-related potentials has been proposed on the basis of different timing for the main components. The readiness potential (RP, or *Bereitschaftspotential*) obviously resembles the contingent responses (*Kornhuber* and *Deecke* 1965; *Gilden* et al. 1966) and represents physiological activity related to conation or preparation for performance of a motor act (*Low* et al. 1966a). *Vaughan* (1975) showed that the negative shift recorded in the classic situation for contingent responses conforms to the topographical distribution of the readiness potential and is more closely time locked to the motor response than to the stimulus.

On the other hand it has to be pointed out that slow (DC) potentials may be observed even when no motor response is required. The subject may only anticipate a meaningful presentation (*Cohen* 1969) or the "response" may only be a decision (*Haider* et al. 1968b). Some clarification may be found in the observation that tasks requiring sensory discrimination are associated with potential distributions consistent with their generation in sensory areas, whereas tasks with establishment of a preparatory

motor set are characterized by potentials arising from motor cortex (Vaughan and Costa 1968; Vaughan 1969). The premovement phasic components are considered to represent cortico-subcortico-motor-cortical loops with cerebellar neurons being active prior to the motor cortex as well as the corticospinal discharge immediately preceding motor contraction. With subdural electrodes during stereotactic surgery in humans a "localized motor potential" is observed at area 4 over the representation of hand or finger movement (Ganglberger et al. 1980). The corticomuscular delay, i.e. the interval between onset of cortical potentials and electromyograph (EMG) activity varies according to the length of neural pathways between brain and contracting muscle (Ganglberger 1962; Vaughan et al. 1968). The late positive wave of movement-related potentials has been considered to represent kinesthetic feedback (Kornhuber and Deecke 1965). Later, Vaughan et al. (1968) have shown that deafferentiation of the upper extremity failed to alter the configuration of motor potentials. Sometimes, especially in mental patients, the late positive components are absent and sometimes they last very long. It may be postulated therefore that these later portions of movement-related potentials may reflect primarily central mechanisms.

## 5 Electrocorticography and Stereo-Electro-Encephalography

## 5.1 Electrocorticographical Studies

The term "electrocorticography" refers to the recording of electrical activity through the dura (the term "electrodurogram" is seldom used), directly from the cortical surface, using electrodes placed in different cortical layers or with a transcortical electrode arrangement.

## 5.1.1 Late Potentials

Late potentials have been observed localized mainly over the parietal and frontal cortex during stereotactic brain surgery (*Groll-Knapp* et al. 1968; *Papakostopoulos* and *Crow* 1976). These studies show that late potentials are cerebral events, independent of stimulus modality and with a localization slightly different from contingent responses. In the most frequently used experimental situation the maximal amplitude of LP is localized over the parietal cortex. Under special circumstances, for instance in situations where the subject is asked to alter a preset plan, the late potentials may also be recorded over more frontal and prefrontal localizations.

## 5.1.2 Contingent Potentials

Contingent potentials, in most situations, are electrocortigraphically recorded over a widespread central and frontal area (Haider et al. 1968c; Papakostopoulos and Crow 1976; McCallum 1978). The distribution for contingent responses is usually similar though not identical to the distributions of late potentials and readiness potentials. Contingent potentials are symmetrically distributed and slightly more prominent anteriorly then late responses and readiness potentials. In animal studies under special circumstances, for instance during waiting periods (Donchin et al. 1971, 1973) or following periods of food deprivation (Borda 1973), some authors observed a central or postcentral dominant potential. In the usual stimulus-response paradigm, most authors (Low et al. 1966b; Rebert 1972, 1973; Borda 1973) found a frontal premotor dominant potential. McSherry and Borda (1973) concluded from their work with animals that contingent potentials are not uniformly distributed throughout the frontal lobe, but rather are a local phenomenon and consist of a superficial cortical negative shift or an intracortical positive shift or, at some locations, both a superficial negative and an intracortical positive shift.

## 5.1.3 Movement-Related Potentials

Cortical recordings of movement-related potentials should be instrumental in providing additional insight into the questions related to preparation, initiation, execution and termination of movement. At the moment the information is meagre and partially contradictory. McCallum (1978) demonstrated that the readiness potential is distributed weakly asymmetrically, showing increased negativity over the hemisphere contralateral to the operative muscles, the largest values occurring pre-Rolandically. In no case was a premotion positivity observed by McCallum. Papakostopoulos and Cooper (1976) found cortical negativity to be completely paralled with EMG recordings up to the point of movement and concluded that these potentials may be the result of movement. Groll-Knapp et al. (1977) and Haider et al. (1979) extracted some patterns of movement-related cortical potentials: one early positive potential shift in the parietal and the frontopolar cortical region, a slow negative potential change occurring on the premotor region and marked phasic potential shifts around the time of movement in the proper motor cortex.

A premovement positivity followed by a negative small "motor potential" can be observed locally at the region of area 4 indicating hand or finger movement. If the electrode is near or at the postcentral area, only a prominent positive wave is seen. These specific localized changes are depicted in Fig. 5 in which the different curves are grouped according to the different representation fields.



Fig. 5. Voluntary movement-related potentials from different localizations in area 4. The eight individual traces are grouped and superimposed according to representation fields. Negativity upwards, button trace: averaged EMG activity.  $\downarrow$  trigger point at time of the button press

#### 5.2 Stereo-Electro-Encephalographical Studies

The term stereo-electro-encephalography (SEEG) has been proposed for the recording of electrical activity from different intracranial levels with the help of stereotactic procedures for localizing the different brain structures (*Bancaud* 1975). The localization procedures that make it possible to determine intracranial and especially subcortical brain structures rather precisely in animals cannot be transposed directly to man. Functional stereotactic investigations in man were dependent on the publication of atlases providing the spatial coordinates of the different brain structures. With the help of special methods and electrodes, it is possible in any patient to reach the intracranial targets previously determined. Investigations are based either on chronically implanted electrodes or recordings made during acute stereotactic surgery. In both situations location of electrodes and duration of investigations are limited according to the medical indication.

# 5.2.1 Late Potentials

Late potentials for different subcortical structures have not been systematically studied up to now. A series of experiments by *Groll-Knapp* et al. (1968) and *Haider* (1970) showed in man, however, that evoked potentials recorded in nuclei of motor thalamus after unexpected acoustic stimuli clearly differentiate from evoked potentials after regular stimulation. The



Fig. 6. Evoked potentials on vertex (scalp), in area 6 (subdural) and in the motor thalamus (nucleus ventralis oralis anterior, *v.o.a.*) after unexpected (---) and expected (---) stimuli. Reference: linked mastoids; negativity upwards.

- V.o.a. = Nc. ventro-oralis anterior
- V.o.p. = Nc. ventro-oralis posterior

Zo.in = zona incerta

- Ce = Nc centralis thalami
- (centre median) Pu = Pulvinar thalami
- M = Nc. medialis thalami
- D.im.1 = Nc. dorsalis intermedius thalami
- D.o.l. = Nc. dorsalis oralis thalami
- Cd = Nc. caudatus
- CA = Commisura anterior
- Cp.in = Capsula interna
- Cz = Vertex

differences consisted of longer latencies and higher amplitudes of the prominent wave in the orienting situation. The late potentials in the motor thalamus demonstrate the involvement of the central motor system in orienting reactions. Fig. 6 gives examples of late potentials in motor thalamus, premotor cortex and scalp derivation.

Acoustically evoked potential shifts in motor thalamus have also been reported by *Ganglberger* et al. (1971) and by *Fukushima* et al. (1976).

The differential effect of attention and task relevance on late subcortical somatically evoked potentials (SEPs) was previously mentioned by *Ervin* and *Mark* (1964) and recently studied by *Velasco* et al. (1975) in a group of Parkinson's disease patients. *Ervin* and *Mark* (1964) reported SEPs at the medial thalamus (CM) with long-lasting rhythmic after-discharges. Significant amplitude reduction of late SEP components was found by *Velasco* et al. (1975) when patients shifted from attention to distraction or habituation. No such changes occurred in early components. In contrast to the early SEP components the late components occurred rather widespread in thalamic lemniscal, prelemniscal and reticular regions.

### 5.2.2 Contingent Potentials

For several years contingent responses have been generally regarded as cortical phenomena, primarily because it was possible to record contingent potentials directly from the cerebral cortex (see Sect. 5.2.1). Recordings taken from thalamic nuclei in a standard CNV paradigm during acute stereotactic surgery indicated, however, a close relationship between the thalamic and the cortical electrophysiological event connected with expectancy (*Haider* et al. 1972, 1979; *Groll-Knapp* et al. 1970). These results are shown, together with similar results reported by *Iliukina* (1977) and *Tsubokawa* and *Moriyasu* (1978), in Fig. 7.

In Fig. 7 contingent responses reported by different authors for different patients during stereotactic brain operations can be seen. The vertex and cortical leads show the typical CNV pattern. In the thalamus, clear local differences may be observed. In the motor thalamus [ventrooralis posterior (v.o.p.) and ventrolateralis (VL) regions] only late potentials after the first stimulus (orienting potentials) and between the second stimulus and the response occur, but no slow rising negativity is seen.

In the nucleus medialis, or dorsomedialis, as well as in the centre median and intralaminar nuclei a clear slow negative potential shift is seen in most cases. This potential shift has a short deflection time and precedes the slow rising negativity of cortical and vertex derivations in its sequential development. This may show that in humans the whole medial thalamus and the mediothalamic-frontocortical system are involved in the generation and control of slow DC potentials of the cortex.

#### Event-Related Slow (DC) Potentials in the Human Brain



Fig. 7. Contingent potentials in a CNV paradigm with a warning and an imperative stimulus (Cl-Cl-R), found by different authors in vertex, cortex, motor thalamus (V.o.p., ventro-oralis posterior, VL, ventrolateralis), medial (M) or dorsomedial (DM) nucleus and non-specific thalamic projection nuclei (CM, centre median; Lam, lamella medialis). Negativity upwards;  $\downarrow$  trigger point; R, time of reaction

These conclusions are supported by the observation of *Tsubokawa* and *Moriyasu* (1978) that ventrolateral thalamotomy has no effect on the amplitude of cortical contingent potentials, whereas medial thalamotomy causes suppression of contingent potentials. One could speculate that this mechanism of suppressing attention and expectancy for exogenous stimuli may be involved in the relief of intractable pain after medial thalamotomy.

*McCallum* and his colleagues (1976) studied DNV distribution with multiple implanted gold electrodes in brain stem structures. Widespread slow (DC) potential changes were observed for brain stem derivations, those from the most rostral electrodes being positive and those from the most caudal (mesencephalic electrodes) negative. These slow subcortical potential shifts look very similar to the contingent responses recorded from some parts of the cortex. In thalamic nuclei little evidence of contingent responses was found.

Some common features in all studies seem to be that complex cortical events are mediated at subcortical levels. Many thalamic nuclei show no slow DC potential changes such as contingent responses, but in some thalamic structures, especially the medial thalamus, slow potential shifts accompanying the beginning and the duration of the cortical slow (DC) events are observed.

Several experiments using animals support the still rather limited findings in human studies. Slow potentials were recorded in different animals from a variety of subcortical nuclei during different experimental paradigms. *Rebert* (1972, 1973, 1976), in macaque monkeys, found negative potential shifts in several non-specific projection nuclei (midbrain reticular formation, nucleus reuniens, posterior hypothalamus), whereas positive shifts developed in the caudatum, amygdala and nucleus dorsomedialis, thus confirming earlier assumptions that positive shifts occur in rhinencephalic structures. For some structures the coincidence of negative shifts with increased multiple unit activity and of positive shifts with decreased unit activity could be demonstrated. The earliest indication of learning (defined by the occurrence of slow potentials) was found in non-specific ascending reticular structures and not in cortical regions.

A series of studies in cats (Skinner 1971; Skinner and Lindsley 1973; Yingling and Skinner 1977; Skinner and Yingling 1977) demonstrate the involvement of the mediothalamic frontocortical system in the generation of cortical slow DC potentials. Bilateral cryogenic blockade in the inferior thalamic peduncle (ITP) with interruption of connections between medial thalamus and frontal cortex abolished all SP changes in the cortex after novel stimuli as well as SP after strong cutaneous stimulation or SP in a tone-shock conditioning paradigm. Slow potential changes in nucleus reticularis thalami are unequally affected. While conditioned slow potential shifts and slow potentials evoked in the reticular thalamic nucleus after medial thalamic stimulation are abolished during ITP blockade, SPs after medioreticular formation stimulation and after strong cutaneous shock remain unaffected.

## 5.2.3 Movement-Related Potentials

The distribution of movement-related potential shifts throughout the midbrain and brain stem was studied with implanted gold electrodes (*McCallum* et al. 1976). Similar to their CNV results the authors found little evidence for movement-related potential changes in thalamic nuclei. In the rostral derivations MRP were positive, whereas in the caudal derivations negative shifts could be demonstrated.

In recent studies *Groll-Knapp* et al. (1977) and *Haider* et al. (1979) reported voluntary movement-related slow potential changes in different subcortical nuclei. Subcortical target points – according to the indication for stereotactic surgery – were in different thalamic nuclei (Nuclei medialis, centre median, v.o.a, V.o.p, L.Po., V.c.pc; see Fig. 8 for explanation of abbreviations), in the tractus spinothalamic, fornix and amygdala [nomenclature of *Hassler* (1959), stereotactic atlas of *Schaltenbrand* and *Bailey*].

Voluntary movement-related slow (DC) potential changes for various thalamic specific and non-specific nuclei and other subcortical structures are shown in Fig. 8.



Fig. 8. Voluntary movement-related potentials from various thalamic nuclei and other subcortical structures. *M.fa.p.*, Medialis fasciculus posterior; *Ce*, nucleus centralis or centre median of *Luys; V.c.pc*, ventrocaudalis parvocellularis; Fx, fornix; *Z.im*, nucleus centralis intermedius; *V.o.p.*, ventro-oralis posterior; *Z.i*, zona incerta; *Am.m.*, nucleus medialis amygdalae. The different beginnings of the traces are due to the different tape speeds during the instantaneous opisthochronic analysis; *R*, reaction and trigger point

In the specific motor relay nuclei and the zona incerta the negativity starts shortly before motor action and is followed by a relatively sharp increase. The peak is reached after the trigger mark produced by thumb press. In the nucleus medialis or dorsomedialis, as well as in the centre median, a clear slow negative potential shift starts earlier and shows a higher amplitude than those recorded from the motor thalamus. In recent studies we observed greatly enhanced motor thalamic potentials during goal-directed movements, which were sustained until the target was reached.

According to our findings four patterns of cortical and subcortical event-related slow potential changes may be differentiated:

- 1. Early positive potential shifts in parietal and frontopolar cortex associated with beginning negative potential changes in medial thalamus;
- 2. Widespread slow negative potential rises in premotor and parietal cortex, increasing during goal-directed movements;
- 3. Localized potential changes on proper motor and sensory cortex, motor thalamus and other subcortical nuclei around the time of movement. Some of these potentials are greatly enhanced and sustained during goal-directed movements; and
- 4. Positive potential declines in most cortical and subcortical electrode sites after movements, decisions or responses.

On the basis of time relationships of these four possible patterns it may be speculated that designs or "ideations" are accompanied by early potential changes in parietal cortex and the mediothalamic-frontocortical system. These potential changes, together with widespread negative potential rises over premotor and parietal cortex (readiness potentials), constitute the preparatory period for judgements, decisions and responses. In the initiation period a premotion positive decline over premotor cortex as well as localized potential changes in motor cortex and motor thalamus are sometimes seen. In the part of area 4 representing hand or finger movements, premotion positivity (PMP) and motor potentials (MP) are seen (Ganglberger et al. 1980). Near or at the sensory cortex only a prominent positive wave occurs. In the upper and lower parts of area 4 a slowly rising negativity reaching its peak around the time of actual movement or shortly afterwards is observed. These negative peaks of motor cortical and motor thalamic potentials are greatly enhanced and sutained during goaldirected movements until the target is reached. Since the motor thalamus is integrated in motor feedback loops from the cerebellum and basal ganglia to the motor cortex, these potentials may be related to the control and internal monitoring of programming activities during initiation and execution of movement. The final post-movement-positive potential decline may represent central mechanisms related to termination of movements and reafferent signals from the periphery.

Many of these interpretations are still highly speculative, but our studies, based on acute stereotactic surgery, demonstrate some mechanisms of summation and integration of the multiple local generators in the cortex to the common patterns of event-related responses emerging from studies using scalp electrodes. Moreover, these studies reveal subcortical involvement in the regulation of cortical activity and the interrelationship between subcortical and cortical electrophysiological events.

### 6 Topographical Aspects (Scalp Distribution)

#### 6.1 Late Potentials

Studies by Vaughan and Ritter (1970) suggested for LPs a dipole layer source with axes perpendicular to the surface of the scalp and centred upon the parietotemporal association area. In accordance with this suggestion the largest amplitude for late potentials has been reported at the parietal electrodes (Donchin et al. 1975). The same modality non-sepcific distribution with a maximum over the parietal region has been found for the "emitted potentials" from missing stimuli. The distributions and presumptive sources of negative and positive components of visually evoked potentials and for potentials after missing stimuli are shown in Fig. 9.

The topographical analyses seem to show that the earlier components have distributions compatible with sources in and near the primary cortical projection area, whereas the potential components for missing stimuli similar to all late potentials have a distribution compatible with a cortical origin in the inferior parietal lobule. The main localization for late potentials has also been found in electrocorticographical studies to be at the parietal cortex (see Sect. 5). Because the topographical distribution suggests an origin in secondary cortical projection areas and in association cortex, *Vaughan* (1975) uses the term "association cortex potential (ACP)".

Since late potentials are not a uniform wave component but consist of a variety of negative and positive waveforms, we must also take into account the possibility of different sites of generation, depending for instance on the task and experimental design (see Sect. 8.1). Late potential components, with distributions largest over posterior regions, have been described during an acquisition task (*Poon* et al. 1974). On the other hand a large frontal positive component following a negative orienting potential was observed for novel stimuli (*Courchesne* et al. 1975). One may speculate, therefore, that different topographical field distributions for late potentials reflect different functions of cortical and subcortical sites with respect to novelty and information processing.

Stereo-electro-encephalographical studies (see Sect. 5) clearly demonstrate that the source of late potentials is not purely cortical. Therefore one has to consider that either subcortical or combined cortical-subcortical sources may account for the reported scalp distribution. Intracranial



Fig. 9. Averaged responses (VER) to 1-s light flash (S) and brain potentials missing stimulus potentials (MSP) associated with unexpected deletion of a flash (S). Potentials are grand means of 11 responses obtained from eight subjects. Isopotential maps on left depict the negative  $(N_1)$  and late positive  $(P_2)$  components of the VER, respectively, together with the estimates of intracranial sources inferred from these data. Maps on the right indicate the distribution and presumptive sources of the missing stimulus paradigm (Vaughan 1975)

studies during stereotactic surgery (*Groll-Knapp* et al. 1968, 1970; *Haider* 1970; *Ganglberger* et al. 1971; *Velasco* et al. 1975) have shown that quickly habituating late potentials for unexpected (orienting) stimuli are widespread in different thalamic nuclei, even in the motor thalamus. This may explain to a certain degree why late potentials show modality non-specific distributions without polarity inversions in scalp and cortical depth recordings and with maximum amplitudes deep in the brain and not near the cortical surface (*Wood* et al. 1979).

#### 6.2 Contingent Potentials

There may be three approaches to the analysis of differences in contingent potentials, recorded from different localizations of the scalp. Some authors consider contingent potentials to be unitary phenomena, while others see them as being composed of several different waves, converging on local recording sites. A third group assumes that each potential is characteristically generated at the local site from which it is recorded (*Weinberg* and *Papakostopoulos* 1975).

Early studies (*Walter* et al. 1964; *Low* et al. 1966a) reported contingent potentials with maximal amplitudes at frontal sites. Later studies reported a gradient of change in the anterior-posterior axis. Many authors (*Cant* et al. 1966; *Walter* 1967a, b; *Cohen* 1969; *Gullickson* 1970) suggested a bilateral symmetrical distribution with maximum at the vertex, being smaller in the frontal region and smallest in posterior areas.

In tasks involving the processing of verbal and spatial information, the amplitudes of contingent potentials were larger at parietal areas compared to frontal and temporal sites (*Marsh* and *Thompson* 1973).

In conditioning experiments differences in location have been found related to the different stages of training. In early training phases a central dominant potential was seen; during overtraining periods a frontal dominant wave was observed (*Borda* 1970; *Hablitz* 1973).

That contingent potentials may indicate functionally different processes in more frontal or more posterior areas is suggested by different studies. Task dependency has been demonstrated by frontal dominant potentials in an auditory discrimination task and central dominant potentials prior to motor movements (*Järvilehto* and *Fruhstorfer* 1970).

Correlation studies of contingent potentials and reaction times showed poor matching of frontal and vertex waveforms contrasting with good matches of central and parietal with vertex waveforms. Moreover, there was a lack of correlation between contingent potentials and reaction times in prefrontal areas but some correlation in central-parietal areas (*Papako*stopoulos and Fenelon 1975). Similarly, Weinberg and Papakostopoulos (1975) used a pattern recognition technique to determine the form differences of contingent potentials in frontal, parietal and vertex sites. Frontal potentials were the smallest and were different in form from vertex, central and parietal sites.

One conclusion to be gained from all these studies seems to be that the wave-form of contingent potentials to a certain degree maps the processing of information under the recording area. In this respect contingent potentials show similarities to late potentials. But besides topographical differences there are differences in the kind of information processing and its outcome. Late potentials are post stimulus and therefore are most sensitive to immediate outcomes of information processing, whereas contingent potentials occur prior to critical stimuli and may reflect changes in different cerebral sites as a result of accumulated information.

#### 6.3 Movement-Related Potentials

The topography of voluntary movement-related potentials was first studied systematically by *Vaughan* et al. (1968) and *Deecke* et al. (1969). *Gerbrandt* et al. (1973), *Becker* et al. (1973), *Deecke* et al. (1973, 1979), and *McCallum* et al. (1976) have also investigated this problem.

There is still some controversy about the topographical distribution of movement-related potentials. Different spatial distribution of maximal amplitudes and onset of the main three potential components preceding the movement, as well as for the post-movement positivity, have been found.

The individual differences in both the prominence and degree of symmetry of the MRP are substantial and greater than for the CNV. These variations could perhaps partly account for some of the discrepancies in the above-mentioned studies. Differences in experimental design and especially in the measurement technique may also explain diverging results.

Deecke et al. (1969, 1973), Deecke and Kornhuber (1977) and Becker et al. (1973) report that the slow surface negative potential (readiness potential, RP) is maximal at the vertex and that this potential is nearly symmetrical in the pre- and postcentral region. It is sometimes larger over the contralateral than over the ipsilateral precentral region, mainly when the dominant hemisphere is involved in the movement. The readiness potential declines in amplitude in more anterior derivations. In frontal and basal leads, however, Deecke and Kornhuber (1977) found this potential to be positive.

The difference in the distribution of the RP compared to that of the CNV indicates that the slow surface negative potential reflects the general facilitating process which pre-activates the brain regions needed in the ensuing situation. During a premovement time the frontal lobe might be inactive (no negative shift) or even inhibited (positive **RP**).

Premovement positivity was also found to be symmetrically distributed and rather widespread over the scalp. In contrast the additional rising negativity occurring shortly before the onset of EMG activity (motor potential, MP) had its maximum over the contralateral precentral motor region, sometimes may only be recorded there and is a very circumscribed phenomenon. This has now been directly demonstrated by our corticographic studies as shown in Fig. 5 (Sect. 5.1.3).

There is rather good agreement among different authors that the final component of the movement-related potential, the relatively large and complex positive  $(P_2)$ -negative wave occurring after the movement, is largest and most asymmetrical predominantly over the motor cortex and that these components are mainly due to the input from joint receptors. Several authors (Vaughan et al. 1968; Deecke et al. 1969; and Gerbrandt et al. 1973) mention the close relationship between the  $P_2$  peak and the "vertex potential" of the somatically evoked potential. The authors have developed their concepts about physiological origin and the physiological relevance of the potential components according to scalp distribution and the time relations of the movement-related potential components. Vaughan et al. (1968) report that movement-related potential components are directly connected to motor cortex activity. In contrast, Gerbrandt et al. (1973) states that the motor origin of the different components of the "average movement potential" components indicates different aspects in the preparation and execution of a motor movement. Taking into account the results of cortical and subcortical investigations of humans (Groll-Knapp et al. 1977; McCallum 1978) mentioned in the previous section this concept now seems the most appropriate.

### 7 Maturational Aspects

### 7.1 Theoretical Problems of Developmental Studies

Developmental studies in slow potential research involve some theoretical problems. Age is generally taken as the independent variable in developmental studies, but the question remains to what extent chronological age is a relevant parameter for a process, such as slow potential changes, which is so clearly linked to behaviour. The great interindividual differences, mainly in children, between chronological age and behavioural age point out this difficulty. There exists no complete parallel between psychological and behavioural maturation, on the one hand, and biological and neurol-

M. Haider et al.

ogical development on the other. One should therefore try to find other criteria, perhaps based on behavioural tests, to form homogeneous groups for developmental investigations.

Compared to the general explosion of research relating to the CNV since its discovery, only a few investigations have attempted to study ontogenetic aspects of SP. It is only in the last years that the number of relevant publications has increased, due mainly to the fact that SP research was introduced to psychiatric research. In this connection one expects that the study of SP in normal subjects can provide the normal data base necessary for comparative evaluation of SPs in psychiatric populations and for the development of adjuvant procedures for the evaluation of higherorder functions in clinical populations.

### 7.2 Late Potentials

The various long latency waves (> 100 ms) seem to be unequally affected by age. Peak latencies of wave components between 140 and 300 ms are reported to be shortest for teenagers and young adults (*Dustman* and *Beck* 1969; *Lüders* 1970; *Schenkenberg* 1970, unpublished work; *Buchsbaum* et al. 1974). Young children (*Dustman*, 0–4 years; *Buchsbaum*, 6–9 years) and older people show longer latencies. *Courchesne* (1979) did not find any change in latencies of these potential components between 6 and 36 years of age. Amplitudes of these components generally decline at higher ages (*Dustman* and *Beck* 1969; *Lüders* 1970; *Buchsbaum* et al. 1974; *Courchesne* 1979).

Wave components occurring after 300 ms were observed by *Kurtzberg* et al. (1979) for children in paradigms similar to  $P_3$  paradigms used for studies of adults.

Recently *Courchesne* (1979) reported on a series of experiments with subjects between 6 and 36 years of age. When events occurred which were not explicitly pre-categorized, subjects of different ages produced ERPs dominated by different waves. In adults  $P_3$  waves were the most prominent, while large negative components with peak latencies at about 700 ms and positive peaks at about 1000 ms were characteristic for 6-month-old infants (*Courchesne* et al. 1977) and for children between 6 and 17 years (*Courchesne* 1979). In this latter group, the  $P_3$  component, which was not clearly developed in infants, had peak latencies at about 700 ms and decreased with age to approximately 400 ms.

In studies in which subjects are actively involved, significant age-dependent changes were not seen (*Marsh* 1978). Higher activity for older subjects was found if a cumulative voltage measure was taken between 200– 500 ms (*Schenkenberg* 1970, unpublished work). Linear correlation coefficients were calculated for ten normal children  $(5.7-12.7 \text{ years}; \overline{X} = 9.5 \text{ years})$  between age and cumulative voltage measures referred to prestimulus baseline for positive shifts occurring during the encodement interval in a three-stimulus paradigm "protracted p 300 waves" (*Otto* et al. 1976). Substantial negative correlations were found for all measurement epochs during the  $S_2-S_3$  (encodement interval) and for the 320-ms epoch preceding  $S_2$ . Young children tended to show positive shifts during the warning and encodement interval while older children showed negative shifts similar to CNV.

Positivity after the warning signal in a classic CNV paradigm was found to be developmentally related (*Karrer* and *Ivins* 1975, 1976). Postwarning stimulus positivity (PWSP) was determined in relation to prestimulus baseline and occurred with a latency of about 300–500 ms. There were significant differences between the two groups under experiment. Younger children (9–12 years;  $\bar{X} = 11.1$ ) had significantly larger PWSP than older children (13–18 years;  $\bar{X} = 15.1$ ) on the vertex. The same trends were not significant in frontal areas. The larger PWSP in young children is interpreted within the p 300 conceptualization as reflecting greater conscious effort to control irrelevant activity and greater uncertainty in responding.

Late negative postwarning stimulus activity was observed during foreperiod experiments by *Loveless* and *Sanford* (1974). These authors found a negative-going potential shift peaking about 1 s after the warning signal. No significant difference in this component was found between two age groups, one with a mean age of 21.0 (20.2-22.7) and a second with a mean of 66.9 years (58.1-75.1). Form and latency of this negative peak were not affected by the use of different sets of foreperiods, and this uniformity strongly suggests that this negative peak is a constant effect of the warning signal and is interpreted by the authors to be part of the orienting response.

Similar postwarning signal activity showed an age dependence for younger subjects in the study by *Klorman* (1975) insofar as this component tended to habituate over several trial blocks in 19-year-olds, but not in 10- and 14-year-olds.

#### 7.3 Contingent Potentials

Compared to the few reports on the age dependence of late potentials the literature on contingent potentials is more systematic and covers a broader span of age. *Walter* (1967a, b), using a constant foreperiod reaction time paradigm, could not observe CNVs in children under 3 years. Nevertheless, it seems possible to elicit contingent variations in very young and immature children (i.e. 3 years). In a systematic study of preschool children, clear

CNVs were recorded by the use of coloured patterns and novel auditory patterns as  $S_2$  without the occurrence of a motor response. Probably the children's general interest, attention, motivation and cooperation were higher due to the different experimental design (*Gullickson* 1973a).

Between the ages of 5 and 15, CNV is rather unstable. Either no CNVs, in the usual sense, are observed or only small inconsistent negative shifts, sometimes even a positive-going shift in the  $S_1-S_2$  interval and a post- $S_2$  negativity (Low and Stoilen 1973; Timsit-Berthier and Hausman 1973). Papani and Zappoli (1973) found "tent-like" CNVs for this age.

In general there seems to exist a positive correlation between CNV amplitude and age during childhood up to the time when the adult contingent variation is established. Table 3 (*Cohen* 1973) demonstrates the age dependency of the CNV.

Age of child	Vertex	Frontal		Central		Parietal	
		L	R	L	R	L	R
6	11	6	6	11	10	12	11
7	13	9	8	12	13	12	11
8	13	9	9	12	13	12	12
9	15	11	10	13	12	13	12
10	14	10	11	13	12	12	11
11	16	12	13	13	14	11	11
12	15	12	14	14	13	11	12
13	16	14	12	13	14	11	10
14	17	13	13	15	14	12	12
15	21	15	13	15	16	12	13
16	20	14	15	17	17	13	12
17	21	15	14	17	18	14	13
18	22	16	15	18	19	13	12

Table 3. Mean amplitude of peak CNV in 6-18-year-old subjects in frontal, vertex and parietal derivations

No agreement has so far been reached about the age at which "adultlike CNV" can be observed. Several authors assume a rather clearly maturation of CNV, at an age of about 10–11 years (Low et al. 1966a; Timsit-Berthier and Hausman 1972, 1973; Low and Stoilen 1973; Lancry 1976; Karrer and Ivins 1976; Karrer et al. 1978). These authors found either immature CNV forms for children younger than 10 years; and the usual configuration for older ones, or no correlation between age and CNV parameters after age 12.

A later age for the maturation of the CNV is suggested by *Klorman* (1975), who did not find the usual CNV configuration in subjects younger than about 14. In the investigations of *Cohen* and his group the adult peak

amplitude of approximately  $20-22 \mu V$  and pattern was reached by age 15 (*Cohen* 1973). In earlier studies an even later age was proposed (*Cohen* 1970, 16–18 years; *Cohen* et al. 1967a, 20 years).

Whereas in adults CNV typically declines abruptly to baseline upon responses to  $S_2$ , in children this termination of CNV is usually gradual and has been suggested as an index of maturation (*Cohen* et al. 1967b; *Low* and *Stoilen* 1973).

Investigations of CNV in older populations were done by *Loveless* and *Sanford* (1974a), *Marsh* and *Thompson* (1973) and *Marsh* (1978). *Loveless* and *Sanford* have shown equal orienting responses for a group with a mean age of 21 and for one with a mean age of 66.9, but much reduced CNV responses at long foreperiod intervals (up to 15 s) for older subjects. The main age difference consisted in a rise of slow potential initiated either immediately following the warning signal or after a brief period and then sustained at a fairly constant but low level throughout the foreperiod.

Marsh and Thompson (1973) compared CNV parameters of a young group (19-21 years) with those of an elderly group (68-80 years). The absence of any consistent age difference agrees with the findings of Thompson and Nowlin (1971), but conflicts with the results of Loveless and Sanford (1974a). If one assumes that CNV reflects the level of arousal, the data suggest that elderly subjects are capable of showing central arousal levels similar to those of the young when the experimental situation requires a high arousal level.

There is an interesting relationship between age and CNV distribution over the scalp. As previously discussed the usual topography of the CNV consists of maximum amplitude on the vertex and higher CNV amplitudes in frontal areas, compared to posterior regions. The anterior-posterior distribution and the amplitude changes with age can also be seen in Table 3. The CNV attains its earliest peak amplitude in younger children in the parietal region. There is a shift to central and frontal maximum amplitude for older children and adults. After 11 years of age the adult pattern of higher CNV in the frontal areas emerged (Cohen 1973). This shift in topographical distribution is due to an increase in frontal CNV, whereas parietal CNV amplitude remains essentially the same between 6 and 10 years of age. Other investigations support these findings, showing clearer CNV in posterior brain areas of young people than in adults (Walter 1967b) and a higher correlation for older children (between frontal and vertex derivation) (Karrer and Ivins 1976). Low and Stoilen (1973) occasionally found parieto-occipital dominant waves in children age 10 and over, though the configuration of CNV was already classified as mature.

As in adults contingent potentials in children appear to be bilaterally symmetrical over the hemispheres (*Cohen* 1973).

#### 7.4 Movement-Related Potentials

There has been little study of the development of voluntary movementrelated potential changes. A readiness potential preceding the motor act was clearly seen by *Gullickson* (1973b) in 3-year-old children. For children up to 14 years, he found premovement negativity to be smaller than the postmovement positivity. Between the age of 12-14, the preparatory potential becomes the dominant negative potential. *Karrer* et al. (1978) also found different wave-forms for children 6-8 years old as compared to young adults (16-18 years). Whereas the slowly rising negativity in adults begins 500-700 ms before EMG onset, in young children a positive-going wave was initiated at this time. This wave was followed by a negative shift peaking about 150-300 ms pre-EMG and a second positive shift.

Three mechanisms are possibly responsible for this clearly divergent wave-form in young children in contrast to the wave-form of adults in polarity and amplitude. First, the second positivity could be a correlate of the requirement of greater activity for the initiation and may reflect an enhanced PMP. Second, the positivity occurring in adults after the movement could be shifted forewards. A third speculation is that the young



Fig. 10. Mean Bereitschaftspotential (BP) amplitude in different age groups. White columns, BP amplitude at movement onset (first action potential in the EMG); hatched columns, BP amplitude 150 ms prior to EMG onset. (Deecke et al. 1978)

child may need some verbal control to organize his motor response. Activity preceding speech has a relatively large positive component.

Only *Deecke* et al. (1978, 1979) have systematically investigated about movement-related potential development in higher age groups. The average BP amplitude is relatively constant from age 17 until the end of the 4th decade (39 years) and then gradually declines. In the higher age groups very small BP amplitudes are observed; above age 60 positive BP can occur. Such positive BP are found in younger people only in frontal leads. For subjects between 30 and 69 the correlation coefficient between age and amplitude  $P_{150}$  was r = 0.58 (p < 0.01) and for  $P_0$ , r = 0.66 (p < 0.001). In contrast, no significant reduction of the motor potential was observed (see Fig. 10).

Gullickson (1973b) describes the anterioposterior distribution of change occurring with age. The maximum preparatory potential was observed at the vertex for all ages. In young children the frontal response was larger than the parietal, whereas in subjects between 12 and 16 years, parietal response surpasses the frontal in magnitude.

Age-dependent changes in the topographical distribution were also found by *Karrer* et al. (1978), who described an increase in the coherence of BP measures between different derivations at higher ages.

## 8 Psychological Correlates

Many psychological processes have been related to slow (DC) brain potentials, and the whole field is as yet ill defined. It is impossible to cover the literature completely and therefore only the main lines of scientific developments and neuropsychological theorizing or model building will be outlined.

## 8.1 Late Potentials as Information-Related Potentials

Substantial increase of late potential components of evoked potentials to various stimuli have been related by many investigators to the amount of information carried by the stimulus (*Chapman* and *Bragdon* 1964; *Sutton* et al. 1965, 1967). Depending on the experimental design the late changes were elicited in a variety of situations and have been interpreted in terms of decision making (*Hillyard* 1969; *Smith* et al. 1970), cognitive evaluation (*Ritter* and *Vaughan* 1969), template matching (*Squires* et al. 1973, 1975, 1977), reduction of arousal (*Karlin* 1970), stimulus uncertainty (*Paul* and *Sutton* 1972), change in preparatory set (*Karlin* and *Martz* 1973)

and selective attention (*Hillyard* et al. 1973). The late potential in all these studies is interpreted as being dependent upon the recognition of known, task-relevant stimuli. Some authors, however, found no differences in  $P_3$  latency and amplitude between task-relevant and task-irrelevant signals (*Roth* et al. 1976).

In another development some authors observed late potentials evoked by novel, rare or unexpected stimuli. This was related to the concept of orienting responses or "what is it" reactions. Examples are demonstrated in Fig. 1. The orienting potentials were independently described by Haider et al. (1968b) and Ritter et al. (1968). Haider et al. observed a late negativepositive wave complex with negative peak latencies between 250 and 350 ms evoked by unexpected changes of stimuli in a time series as well as unexpected changes of sense modalities (Fig. 1A). Ford et al. (1973) found a modification of late negative waves with peaks between 190 and 280 in response to relevant or irrelevant stimuli from a relevant modality or to novel stimuli. Ritter et al. (1968) as well as Ritter and Vaughan (1969), observed P<sub>3</sub> waves correlated with unpredictable changes in pitch (Fig. 1B). Demaire and Coquery (1977) showed that in the absence of any task to be performed by the subjects after the occurrence of stimuli, a  $N_2 - P_3$  complex is the most conspicuous component of the evoked potential to be differentially affected by stimuli to expected and unexpected modalities. Prominent negative-positive wave complexes with latencies between 210 and 450 ms have been described by Squires et al. (1975 and 1977). The negative peaks occurred in response to rare stimuli, regardless of whether these were task relevant (Fig. 1D). The positive component P<sub>350</sub> is enhanced when the stimulus is rare and task relevant. A slow wave component following the  $P_{350}$  is related to the same variables but with a different scalp distribution. Courchesne et al. (1975) found a negative late potential component, similar to that described by *Haider* et al. (1968b) as largest for novel stimuli, which are most likely to produce an orienting response (Fig. 1C). This negative wave was followed by a large frontal positive component (330-360 ms). Another, more posteriorly evoked positivity (380-430 ms) was evoked by recognizable, simple stimuli. Courchesne mentions cases of an atypically high amplitude negative process (onset at 200 ms and a duration of 450 ms) similar for all types of unexpected stimuli. Other researchers have reported large negative deviations in response to complex visual patterns (Cohen and Walter 1966; Lifschitz 1966; Symmes and Eisengart 1971). An orienting potential with an onset latency of 300-350 ms and a peak about 450-460 ms following the warning signal in a classic CNV paradigm has been interpreted by Loveless and Sanford (1974) as a cortical component of the orienting response to the warning signal. This orienting response was not affected by instructions such as "sensory set" or "motor set". Finally it has been shown that orienting

which contains motor elements in the sense of readiness for movements enhances the late negative potential. This is demonstrated through experiments in which subjects had to make a direct or a delayed response (*Kok* 1980; *Ritter* and *Vaughan* 1980).

The great variety of situations evoking late potential changes and the correlations between psychological processes and different potential components are extensively discussed in recent review articles by *Donchin* (1979) and *Sutton* (1979).

It may be concluded from all these studies that the late potential is not a unitary phenomenon, but consists of multiple components, each related perhaps to different aspects of cognitive behaviour. We therefore proposed in Sect. 2 that all these potential components may be summed up under the heading of "information-related potentials". This term may also include the "emitted potentials" occurring at about the time of an expected but absent stimulus when the event of stimulus absence provided information or was salient for the subject (Rusinov 1959; Haider 1965; Sutton et al. 1967; Klinke et al. 1968; Barlow 1969; Weinberg et al. 1970, 1974; Ruchkin and Sutton 1973; Picton and Hillyard 1974). Emitted and evoked potential components are similarly affected by variations in event probability (Rushkin et al. 1975).

Concerning the differences of polarities, latencies and topographies of information-related potentials one may speculate that the negative components and the frontally localized positive late component constitute a central orienting response, with motor readiness being most prominent in situations using rare stimuli, with novelty and unpredictability, and even without task relevance. Such late potential components have been observed by us in motor thalamus and motor as well as frontal cortex (Fig. 6).

Especially in motor thalamus a quick habituation occurred, showing that the whole motor control system is involved in the response of the organism to novelty. The different positive components located more posteriorly at the vertex and parietal regions seem more related to task-relevant information processing probability aspects of the stimuli and stimulus categorization. In some situations different negative and positive potential components may produce potential peaks which follow each other in a sequence of cognitive processing; in others, one component may outweigh the others. The potential peaks occurring in a certain situation depend on many factors. The peak latency is dependent on the internal event to which it is related, which may at least be partly independent of stimulus and response. The polarity is dependent on the basic DC level, which may to a certain degree represent the level of cortical activation.

#### 8.2 Contingent Potentials and Psychological Variables

Contingent potentials occur in a great variety of situations in which perceptual, cognitive and motor acts are prepared, anticipated or conditioned. The paradigms in which CPs are generated may tentatively be classified into four general types: (1) holding a motor response in readiness; (2) preparing for a perceptual judgement; (3) anticipation of a positive or negative reinforcer; and (4) preparing for a cognitive decision (*Hillyard* 1971, 1973). In highly motivated and stressful situations contingent potentials are clearly seen in EEG records without averaging. One such example, from a musician who had to play a horn solo in an open performance, is demonstrated in Fig. 11.



Fig. 11a-c. Contingent variation in on-line EEG of a horn player prior to his solo performance during a concert. a EEG (P<sub>3</sub>-linked mastoid), negativity upwards; b ECG; c Sonogram. (*Haider* and *Groll-Knapp* 1971)

For the interpretation of empirical results, psychological constructs and theoretical concepts related to learning and memory, expectancy, conation, motivation and attention have been used.

Slow potential shifts related to learning and conditioning are reported by *Rowland* (1960, 1967). Cortical slow potential shifts to non-reinforcing stimuli disappeared with loss of novelty and to reinforcing stimuli diminished with drive reduction. In learning the slow (DC) potentials to non-reinforcing stimuli are acquired in some cortical loci and become drive dependent. It has recently been shown that the slow (DC) potentials may be conditioned operantly in humans by appropriate visual and acoustical feedback (*Birbaumer* et al. 1978; *Bauer* and *Lauber* 1979).

The contingency of reinforcement was also important in early theorizing on CNV. The basic paradigm for generating a CNV involved a first stimulus (the warning stimulus, or  $S_1$ ) followed by a second stimulus (the imperative stimulus, or  $S_2$ ). When  $S_2$  was omitted, CNV amplitude was reduced or "exstincted" after several trials (*Walter* et al. 1964; *Low* et al. 1966b). This was interpreted in terms of "expectancy processes". Expectancy was defined as "subjective probability" or relative certainty that  $S_2$ will follow  $S_1$ , and the CNV was phenomenologically called "expectancy or E-wave" (*Walter* 1964; *Walter* et al. 1964). Obviously this expectancy hypothesis leaves unexplained the many changes occurring in contingent potentials with unchanged statistical association between  $S_1$  and  $S_2$ .

That the intention to perform an act ("conation") is a main determinant for the development of contingent potentials is demonstrated by the finding that the amplitude of such potentials is significantly elevated when a motor response is given to  $S_2$  compared with the absence of a response (Irwin et al. 1966; Jus et al. 1968; Low et al. 1966a; Peters et al. 1970; Small and Small 1970; Straumanis et al. 1969). In addition the potential magnitude was found to be directly proportional to be amount of anticipated force needed for a response (Rebert et al. 1967; Low and McSherry 1968). Furthermore, amplitude changes proportional to the anticipated speed of movement in response to  $S_2$  were described by *Grünewald* et al. (1979a). When the response terminates  $S_2$  and helps to avoid a shock, CP amplitude is significantly enhanced (Peters et al. 1970; Cant and Bickford 1967). All these results provide evidence that motivational states and response intentions are important for the development of contingent potentials. But it has been shown in other experiments that systematic changes in such potentials may occur without changes in the response parameters or even with no overt response (Irwin et al. 1966; Gullickson 1970).

Attention as an anticipatory selective function has been related to CNV development by different lines of experimentation. The correlation between fast reaction times (RT) and large CNVs has been interpreted to signify that CNV is a sign of attention (*Hillyard* and *Galambos* 1967). But later work on the relations of RT to CNV shows conflicting results. Some authors (*Walter* et al. 1964; *Hillyard* and *Galambos* 1967; *Rebert* and *Sperry* 1973) reported significant correlations. Others (e.g. *Waszak* and *Obrist* 1969) failed to find such correlations, and *Rebert* and *Tecce* (1973) concluded in their review that CNV and RT are essentially unrelated events. *Papakostopoulos* and *Fenelon* (1975) found few intra-individual

correlations, but some correlations in average data across subjects in the central-parietal areas, contrasting with a lack of correlation in prefrontal areas.

A second line of evidence for the influence of attentional factors on CP comes from correlations found between CNV amplitudes and measures of perceptual accuracy. Such results have been attained in auditory signal detection (*Hillyard* 1969, 1971), auditory intensity discrimination (*Wilkinson* and *Haines* 1970), visual pattern recognition (*Cohen* 1973) and visual position discrimination (*McAdam* and *Rubin* 1971). But there have also been studies in which no association between perception and CNV in auditory discrimination (*Järvilehto* and *Fruhstorfer* 1970) and auditory signal detection (*Paul* and *Sutton* 1972) was found.

A third group of results involves changes of attention to  $S_2$ , for instance by reducing the intensity of this expected signal to near threshold level. In these cases higher CNV amplitudes have been found (*Low* et al. 1967; *Rebert* et al. 1967; *Faidherbe* et al. 1969). The increased CNV may indicate a facilitation of sensory processing in these instances, but other factors such as motivation and activation or arousal may also be involved.

A fourth group of findings is that distracting stimuli produce concurrent reduction in CNV and response effectiveness. CNV magnitude has been decreased by such extra stimulation as conversation and reading (Walter 1964), irrelevant tones (McCallum and Walter 1968), presenting and remembering letters (Tecce and Hamilton 1973) and background music (Miller et al. 1973). In the latter study, CNV amplitude was decreased only when  $S_2$  had a different sense modality than the extra stimulation. The results of all these studies have been interpreted as CNV distraction effects and as support for an attention hypothesis of CNV development (Tecce 1972; Tecce and Hamilton 1973; Tecce et al. 1976). Studies with sustained cognitive activity produced, in addition to the CNV changes, significant elevations in heart rate levels and also increased frequency of eye blinks (Tecce et al. 1976). In the frame work of a distraction-arousal hypothesis these result were interpreted as indicating heightened autonomic arousal which accompanies and mediates the disruption of CNV development.

In conclusion one has to admit that the CNV does not in general predict subsequent behavioural performance with a high degree of reliability (*Hillyard* 1973). Since it occurs in preparation for very diverse types of perceptual, motor and cognitive acts, the results may be conceptualized in a hierarchical model of activation and related behavioural events (*Haider* 1969, 1970). In Sect. 11 we will try to elaborate such a hierarchical theoretical model of slow (DC) potentials, activation and behavioural states.

The topographical differences in scalp derivations (Sect. 6.2) as well as in cortical and subcortical leads described in Sect. 5 seem to show that

contingent potentials are not a unitary process but rather may be differentiated into several task-specific event-related potential components. Järvilehto and Fruhstorfer (1970) distinguished a central premotor dominant and a frontal dominant CNV accompanying auditory discrimination, and Cohen (1973) reported a more posteriorly distributed negativity preceding a visual discrimination. Electrocorticographical studies on animals (McSherry and Borda 1973) and man (Groll-Knapp et al. 1977; Haider et al. 1979; Papakostopoulos and Crow 1976) demonstrated that contingent responses are not uniformly distributed over the cortex but consist of a complex of rather local phenomena. The common pattern emerging from studies with scalp electrodes obviously indicates an integration of various cortical and subcortical mechanisms. Since the results of split brain experiments showed only slight deviations of bilateral symmetry it has been proposed (Hillyard 1973) that each type of human contingent potentials contains a bilaterally symmetrical, diffuse component, possibly under reticular and non-specific thalamic control, to which is added a process-specific negativity distributed across the cortex according to which thalamocortical pathways are active.

The return to baseline after the motor performance has been called CNV resolution. Some variables have been shown to accelerate this return. Using complex tasks and a paradigm with more than two stimuli, it has been shown that CNV resolution was caused neither by the overt response, nor by the final decision concerning the nature of the stimulus, but probably by the first coarse identification of the stimulus (*Wilkinson* and *Spence* 1973). It was also shown that the negativity was sustained during periods of unconfounded expectant attention, and it was suggested that a sustained negativity, sometimes seen in behaviour pathology, may be attributable to the attempt of subjects to continue processing information after S<sub>2</sub> and the response have occurred (*Weinberg* 1973).

## 8.3 Psychological Aspects of Movement-Related Potentials

It has been mentioned in the last section that the amplitude of contingent potentials depends on response variables. The amplitude is elevated when a movement is made and especially when the response has an operant quality and motor readiness potential occur together and form a complex, "hybrid" wave. But when subjects are required to perform a voluntary movement in the absence of external stimuli, the motor readiness potential is influenced by similar psychological variables such as the contingent potential, which increases with greater involvement in the experimental task (Kornhuber and Deecke 1965; Kornhuber et al. 1969) and varies with different motivational and attentional states. The amplitude can, for instance, be elevated by monetary reward (*McAdam* and *Seales* 1969) and correlates with perceptual accuracy (*McAdam* and *Rubin* 1971). The readiness potential preceding a verbal response seems to be largest over Broca's area (*McAdam* and *Whittaker* 1971). During a retrieval task, differences between words recognized on a list and words not recognized occurred over the temporal lobe. Effects of physical aspects of movements (speed, duration and force) on the movement-related potentials have been demonstrated by different authors (*Hazemann* et al. 1968; *Kutas* and *Donchin* 1977; *Becker* and *Kristeva* 1980; *Grünewald* et al. 1979a).

With forearm extensions against different forces opposing the movement, a surface negative-positive deflection appeared after movement began (*Wilke* and *Lansing* 1973). The greater force condition produced greater amplitude of this wave. Similarly, slow waves following movements were seen after clenching the fist or articulating words (*Lelord* et al. 1973). With imperative instructions, contingent potentials could be conditioned to the slow activity evoked during and after movements.

Recently it has been reported that during smooth goal-directed hand movements the premotion negativity increases and remains during movement until the target is reached. In a feedback task this goal-directed movement potential showed an increased negativity and a large positivity after the feedback stimulus. It is suggested that the bilateral component of the negative slow potential shift during goal-directed movements is related to processes of intention and expectancy and that the lateralized components reflect cortical activity specific to the movement execution (*Grünewald-Zuberbier* et al. 1978a, b; *Grünewald* et al. 1979b).

The motor thalamus exhibits only a small wave during short ballistic movements at the time of movement. This is greatly enhanced and extended during the period of goal-directed movements, thus showing that thala-mocortical interactions are important in the execution of aimed and goal-directed motor acts (*Knapp* et al. 1980).

# 9 Psychiatric and Neurological Studies (Clinical Application)

Many clinical psychiatric and neurological investigations have been carried out, the results being somewhat different, sometimes equivocal, sometimes even contradictory. Consequently the diagnostic value of studying the various forms of slow potential phenomena remains open for discussion. Several authors even discuss the possibility whether the differences found are caused only by varying degrees of attention of the subjects.

Some of the various results attained by different groups of investigators may be explained by the means of selection of psychiatric patients and also by national differences in classification of psychopathology. Differences in method may also play a role.

In the earliest stages of CNV investigation, some irregularities were noted in mentally disturbed patients and neurotics. Patients with anxiety or phobias were reported to develop CNVs very slowly and with smaller amplitudes, the E-waves being more sensitive to equivocal presentations. Psychopathic delinquents showed "little or no sign of an E-wave", but rather an augmentation of the non-specific (late) components of the EP (*Walter* 1964; *McCallum* 1966, 1967a).

Patients with compulsive-obsessive disorders showed irregularities, especially in relation to the effect of the operant response, i.e. slow return to baseline after  $S_2$ , and single subjects even showed what was later called a postimperative negativity while failing to develop a CNV (*Walter* 1964, 1966a, b; *Timsit-Berthier* et al. 1971).

In highly anxious patients smaller CNVs were found and distraction considerably attenuated the amplitude, which was also seen to a lesser degree in normal controls. Obsessive neurotics had slightly higher CNVs and showed faster recovery of amplitude after distraction (*Walter* 1964, 1966a, b, 1967; *McCallum* 1967b; *McCallum* and *Walter* 1967a, b); the higher amplitude was also confirmed by other groups (*Dongier* and *Bostem* 1967; *Timsit* et al. 1970).

Slightly smaller CNVs had been found in chronically anxious neurotics and in schizophrenic patients (*McCallum* 1966, 1967a). When mild distraction was introduced (irregular tones as a background), the amplitude could be reduced considerably. After elimination of distraction, normal controls recovered quickly, recovery being delayed in highly anxious neurotics and schizophrenics. Obsessive neurotics, on the other hand, showed recovery of the CNV similar to that of non-patients. After successful treatment of highly anxious neurotics the CNV amplitude increased and also became more resistant to distraction. The probability was pointed out "that the common psychological factor shared" by neurotics and schizophrenics with small CVNs "is a failure of attention" (*McCallum* 1967a).

No correlation of anxiety with amplitude of CNV was found by others (*Low* et al. 1967; *Bostem* et al. 1967; *Dongier* and *Bostem* 1967), while attenuation of amplitude in a high anxiety group was found only under stress (*Knott* and *Irwin* 1968, 1973), females being more sensitive to stress than males (*Knott* and *Peters* 1974).

The tendency of mental patients to show prolonged CNVs has been reported by McCallum (1969), who had also published traces of post-imperative negativity in a schizophrenic patient, after-distraction showing an independent post-imperative negativity. The significance of this was pointed out later by *Dongier* et al. (1973), who also stated the necessity of averaging epochs of 8 s (*Timsit* et al. 1970) for studying prolonged CNV or

post-imperative negative variation (PINV). At the same time the question has been raised whether a PINV is necessarily a prolonged CNV.

*Timsit* et al. (1970) compared CNVs of normal controls, neurotics (hysterics and obsessives) and psychotics (early and chronic schizophrenics and manic-depressives). Amplitude of CNV was found to be generally larger in obsessives than in hysterics (in agreement with *Walter* 1966a), while the highest incidence of prolonged CNV was found in psychotics (in agreement with *McCallum* 1969) and the lowest in normal controls. The authors concluded that the probability of the occurrence of a prolonged CNV increases with the severity of the psychological disturbance.

No difference in amplitude between neurotics and normals was later reported (*Timsit-Berthier* et al. 1971) and no prolongation of the *Bereit-schaftspotential* was found in these cases. Psychosomatically ill patients were found to have a greater CNV amplitude than normals (*Dongier* and *Koninckx* 1970).

Different types of CNVs, including post-imperative negativity, were worked out by *Timsit-Berthier* et al. (1973). The highest incidence of small CNVs was found in psychotics (in agreement with *McCallum* 1966, 1967b; *McCallum* and *Walter* 1968; *Small* and *Small* 1971; *McCallum* and *Abraham* 1973; *Abraham* et al. 1976). The highest incidence of prolonged CNVs was found in psychotics (in agreement with *McCallum* 1969; *Timsit* et al. 1970).

Studying the motor readiness potential (RP) in normals, neurotics and psychotics, the Liege group found, mainly in psychotics, and less frequently in neurotics, a smaller motor RP with slow and only gradual return to baseline after the voluntary movement.

*Dongier* et al. (1973) found the highest incidence of abnormal and prolonged CNVs in psychotics. The prolongation was reported to shorten with clinical improvement.

Contrary to earlier statements of the Liege group the difference in amplitude of CNVs between hysterical and obsessive neurotics was not confirmed in this series, which also found that psychotics tended to have a prolonged motor RP. Also not found by this group was the small amplitude of CNV in psychopathic criminals, contrary to the findings of the group in Bristol, which may be explained by a different selection of subjects.

In subjects suffering from phobic neurosis, CNV was found to be of higher amplitude and of longer duration when phobogenic stimuli were used, while at the same time RT after  $S_2$  was found to be shortened (*Barbas* et al. 1978).

Schizophrenics showed slightly attenuated amplitudes together with a high trial-to-trials variability (*McCallum* 1966, 1967a, b, 1969). Another study confirmed these earlier observations (*McCallum* and *Abraham* 1973).

Especially florid cases of early schizophrenia showed small mean CNV amplitudes in the "acquisition condition", together with further reduction in the "distracting condition". The initial amplitude was smaller and the reduction by distraction was more pronounced in cases with more severe symptoms. In a more recent study (*Abraham* et al. 1974, 1976) using another group of schizophrenics, acquisition and distraction amplitude, prolongation and parietal spread [amplitude difference between vertex ( $C_z$ ) and postcentral midline derivation ( $P_z$ )] were investigated, the subjects being without medication at the first test. The earlier observations were confirmed: prolongation tended to disappear with treatment and improvement. Patients with more florid symptoms showed less differences in amplitude values between  $C_z$  and  $P_z$  than normal controls.

That post-imperative negativity could be selectively attenuated by hyperventilation without seriously affecting CNV amplitude and duration was shown by *Dubrovsky* et al. (1973).

Total sleep loss for one night reduced CNV amplitude considerably; sleep loss for two nights abolished it (*Naitoh* et al. 1971; *Naitoh* and *Hilbert* 1976).

Contingent negative variation in cases with known brain lesions was investigated by several groups. *McCallum* et al. (1970) and *McCallum* and *Cummins* (1973) reported reduction of amplitudes over unilateral lesions, while cases with bilateral lesions and bilateral Parkinson's disease patients showed generally small amplitudes, the latter sometimes showing a tendency to inverse polarity.

The *Bereitschaftspotential* in Parkinson's diesease patients was found to be severely attenuated over both hemispheres in bilateral cases, while hemi-Parkinson's diesease subjects showed severe attenuation contralaterally to the akinesia-affected side of the body (*Deecke* et al. 1976a).

Zappoli et al. (1976) reported that absent CNV in cases with frontal space-occupying lesions could be restored after surgery in some cases. This group also investigated patients who had previously undergone extensive prefrontal lobotomy. Of the small number of eight patients, five had "fairly typical" CNVs. In these subjects the actual extent of the interruption of the mediothalamic-frontocortical system was deduced only from surgical reports, the anatomical evidence of the extent of interruption remaining open (Zappoli et al. 1976, 1980).

Children with poor abilities to concentrate showed longer latencies and smaller amplitudes of the late components of EPs and smaller CNVs (*Grünewald-Zuberbier* et al. 1978b; *Grünewald* et al. 1978).

In dyslexic children CNV was found to be attenuated or even diminished when a visually presented word was used for  $S_2$  (Fenelon 1968).

Differences in CNV between normals and stutterers were reported to be found over inferiofrontal regions by Zimmermann and Knott (1974).

There was no difference in vertex CNV when stutterer spoke normally, but a less hemispheric asymmetry was found in stutterer over inferiofrontal regions. When stuttering occurred no vertex CNV was seen.

In an examination of aphasic, dyslexic and normal children, using a three-stimulus paradigm, *Otto* et al. (1976) found greater positivity in aphasic and dyslexic children in the interval between warning and encodement.

#### 10 Drug Effects and Toxic Influences

Some observations of drug effects on CNV were initially mentioned by *Walter* (1964). Deprivation of (the accustomed intake of) caffein attenuated the CNV which was restored by administration of caffeine sodium citrate. The same result was found with amphetamine. In one subject administered 100  $\mu$ g LSD-25, the E-wave following single flashes "was enormously augmented and spread to nearly all regions" (*Walter* 1964).

Low (1969) saw no apparent effect on CNV amplitude in subjects given amphetamine or chlordiazepoxide. An increase of CNV amplitude was seen by McCallum (1969) in those administered methedrine. After administration of large doses of dextro-amphetamine *Tecce* and *Cole* (1974) found reduced CNV amplitudes in subjects who showed a paradoxical drowsiness in the 1st hour. Others, with increased alertness, showed increased CNV amplitudes.

The influences of marijuana and alcohol, methamphetamine and secobarbital have been studied by Kopell et al. (1972), Low et al. (1973), and by Roth et al. (1977). Flurazepam was found to cause a marked reduction of the EP to S<sub>1</sub> while the CNV amplitude was only slightly attenuated (Hablitz and Borda 1973). Ashton et al. (1974) compared the effects of caffeine, nitrazepam and cigarette smoking on the CNV. The mean amplitude of the CNV was clearly increased by administration of 300 mg caffeine citrate; it was significantly decreased by 2.5 mg nitrazepam. In smokers the change of CNV amplitude differed between individual smokers, some smokers showing an increase and others a decrease in amplitudes. In a later study (Ashton et al. 1978), the effect of nicotine was studied after smoking and after intermittent intravenous administration of 150  $\mu$ g nicotine in volunteers. Initial small doses increased CNV amplitude while repeated small doses decreased the amplitude. The biphasic effect of nicotine (small doses stimulating, larger doses acting as a depressant) was found also during cigarette smoking and was dependent on individual smoking habits:

Increase of the late components of visual evoked potentials (VEP) during tobacco smoking was found by *Friedmann* et al. (1974), while the auditory evoked potential (AEP) tended to be reduced.
Low doses of carbon monoxide were found to attenuate the CNV amplitude, though this effect was not as clearly seen in a more recent study (*Groll-Knapp* et al. 1972, 1978), which also showed that with low level carboxyhemoglobin (COHb) the VEP remained unchanged while there is a marked effect on the somatosensory evoked potential (SEP), which was lessened with higher COHb levels.

In a collaborative pilot study between investigators in Bristol and Newcastle-upon-Tyne, the CNV was studied under interference with the action of some putative neurotransmitters by intramuscular administration of antagonists and a placebo (physiological saline solution). Atropine was used as antagonist of acetylcholine, thymoxamine against noradrenalin, and metoclopramide as an antagonist of dopamine.

Atropine and metoclopramide produced a steady fall in CNV amplitude, a minimum amplitude was reached earlier with atropine, followed by partial recovery during the experiment. No significant change of mean CNV was produced by thymoxamine or the placebo. At the same time of the  $S_2$  response reaction time (RT) was measured. There was no change with atropine and placebo. Reaction time was lengthened with metoclopramide and shortened by thymoxamine, which had no effect on CNV amplitude.

The handicap of the very short plasma life of thymoxamine was also discussed. It was suggested that there is more than one putative neuro-transmitter involved in the genesis of the CNV (*Thompson* et al. 1978).

In shoe industry workers exposed to chronic poisoning with adhesive solvents the CNV proved to be of no clinical value, while the raw EEG showed slowed  $\alpha$ -rhythms and diffuse abnormalities, and the motor nerve conduction velocity was decreased in accordance with the clinical polyneuropathy (*Zappoli* et al. 1978b).

In conclusion it may be stated that the evidence concerning the relation between biochemical and toxicological effects and slow (DC) potential changes in the brain is currently scarce and inconsistent. One may hope that more biochemists and toxicologists become interested in event-related brain potentials and contribute to a more coherent theoretical and empirical approach in the future.

# 11 Concluding Remarks: a Hierarchical Model of Slow (DC) Potentials, Activation and Behaviour

In reviewing the research field of event-related slow (DC) potentials one gets the impression that much data have been accumulated, but that the experimentor is still faced with problems and questions similar to those of 20 years ago. We obviously need more principal data and new theoretical



fied manner some relationships between different, long- and short-term slow (DC) potentials (left), some activation mechanisms Fig. 12. Hierarchical model of slow (DC) potentials, activation and behaviour, provided as an aide in conceptualizing in a very simpli-(middle) and some behavioural states (right) approaches, and we must try to analyse psychological data which is as complex as neurophysiological data. On the one hand we must try to understand the basic neurophysiological mechanisms to be able to use event-related potentials as better indicators for psychological theory. The potentials clearly depend to some degree on psychological states and processes, but the question is how specifically they represent them. Obviously we are able to attend to certain stimuli and to learn and perform motor acts without reliably occurring slow potential changes. This may be disappointing but it demonstrates on the other hand the great adaptability of our neural functions. The brain may solve different problems by using different strategies. All we can expect therefore is to find dynamic patterns of activity with some sequential development in time.

Within a certain period of time we assume that different slow (DC) potentials occur in the brain with different slopes, different time courses and different amplitudes. The potential changes, recorded in a specific situation, depend on many variables such as the state of the brain and the experimental design. We must try to conceptualize the functional relations of all the different slow (DC) potentials in some kind of a theoretical model and to combine the study of general, long-term potential changes with event-related short-term changes.

Many investigators interpreted slow (DC) potentials to be a consequence of altered levels of cortical activation (Goldring and O'Leary 1951; Caspers 1965; O'Leary and Goldring 1964; Walter et al. 1964; Rowland 1960, 1967). Findings on event-related potentials suggest that more differentiated activation processes are involved (Routtenberg 1968; Rebert 1973; Tecce 1972; Desmedt and Debecker 1979a, b) and results of stereo-electroencephalographic studies, summarized in Sect. 5, demonstrate the importance of subcortical, reticular, thalamic and striatal mechanisms.

In Fig. 12 we show a first approach for a hierarchical model of slow (DC) potentials in relation to concepts of activation and behaviour. A hierarchical system of activation was proposed earlier (*Haider* 1969, 1970).

First we have some general activation or arousal mechanisms, regulating wakefulness and sleep. Related to these are long-term slow DC potentials which occur with gross transitions from one state of the brain to another, such as the sleep-wakefulness cycle (*Caspers* 1961, 1963) or seizure activity (*Speckmann* and *Caspers* 1979).

We then have "tonic" activation with long latencies and durations, slowly changing the state of the organism between low and high arousal; reticular and limbic activation mechanisms mediate these changes. Related to this may be middle term (tonic) slow (DC) potentials, observed, for instance, in some studies of *Bechtereva* (1974).

Next there are "phasic" activation or arousal mechanisms with shorter latencies and durations. They are mediated by the diffuse thalamic projection system (Jaspers 1960) and the truncothalamic system, including a striatopallidal neuronal chain (Hassler 1978). Event-related (phasic) slow potentials related to expectancy, attention and motor readiness (contingent and readiness potentials) may be strongly connected to the functioning of these systems as well as to the mediothalamic-frontocortical system. It is now well established by studies of different authors that during stereotactic surgery in humans contingent potentials (CNVs) occur in all parts of these systems (Groll-Knapp et al. 1977; Haider et al. 1968a, 1979; Iliukina 1977; McCallum et al. 1976; Tsubokawa and Moriyasu 1978).

Finally we have to consider a highly "differentiated and selective" activation related to selective perceptual, cognitive and motor acts, without any gross changes in arousal level. For instance, some components of information-related and motor potentials may depend on the more specific thalamocortical and striatocortical mechanisms involved in these processes. These mechanisms for the most part are feedback mechanisms so that one should really use the terms "thalamocortical-thalamic" or "striatothalamiccorticostriatal", etc. Striatocortical mechanisms include pathways to the neurons of the pallidum which send off ascending non-specific or semispecific pathways which travel through the rostral pole of the thalamus and run through specific and non-specific afferents to almost all cortical fields (Hassler 1978). By stimulating the pallidum or the motor thalamus (v.o.a.) or the anterior thalamus during stereotactic surgery a "psychmotor advertance mechanism" may be elicited. This consists of turning the head, directing attention to the contralateral side and dilating the pupil. This may be related to "orienting" potentials which we found in the motor thalamus.

Motor thalamus and motor cortex also show clear motor potentials which at the proper motor cortex (area 4) are demonstrable only very specifically and are localized on the parts representing hand- or finger movements (*Ganglberger* et al. 1980). Concerning the focussing of attention it should be mentioned that a well-organized corticostriatal feedback mechanism with glutamate as the transmitter has been described. This mechanism, especially in relation to the function of the putamen, may be involved in focussing attention to one single perceptive, cognitive or motor act and fading out all other events (*Hassler* 1978). Information- and movement-related potentials may in this connection indicate which thalamocortico-thalamic or striatothalamic-corticostriatal pathways are open and active during a specific mental act.

In a hierarchical system of activation it must be assumed that each level of the hierarchy influences and determines the next one and vice versa. To be able to perform selective perceptional, cognitive and motor acts we need a waking organisms with optimal (medium) arousal and with attentiveness to a specific class of stimuli. Physiologically the upper levels of the hierarchy determine the polarity of potential changes at lower levels by regulating the basic activation-related DC level (*Speckmann* and *Caspers* 1979). Low levels of epicortical DC negativity (low activation) lead to predominantly negative event-related slow potentials and high levels to predominantly positive components.

Empirically the generality or specificity of activation may be operattionally defined by the number of variables differentiating the different levels. The more indicators vary with a certain level the more general it is. The change from sleeping to waking is accompanied by a great variety of physiological changes, amongst others by changes in body temperature, pulse rate, EEG and slow (DC) potentials. Distracting influences on contingent potentials may be an effect of heightened autonomic arousal, as shown by the elevations in pulse rate (*Tecce* et al. 1976). Some eventrelated slow (DC) potential changes are accompanied by phasic changes in EEG activity. For instance, an inverse relation between  $\alpha$ -activity and contingent potentials was found (*McCallum* and *Walter* 1968; *Pfurtscheller* and *Aranibar* 1977).

Finally, some information- and movement-related potential components are obviously related to specific mechanisms without any change in the intrinsic EEG rhythm or in other indications of arousal.

Many of the assumed relationships in the hierarchical model outlined here are arbitrary and speculative.

With refined methods and experimentation the functional relations will be more precisely definable in future experimentation. What we need in the whole field for the future is not so much phenomenology but a more theoretical and neuroscientific framework within the broad context of brain systems and brain functions, including event-related slow (DC) potentials together with psychological states and processes.

#### References

- Abraham P, McCallum WC, Docherty T, Newton P, Fox A (1974) The CNV in schizophrenia. Electroencephalogr Clin Neurophysiol 36:217-218
- Abraham P, McCallum W, Gourlay I (1976) The CNV and its relation to specific psychiatric syndromes. In: McCallum WC, Knott JR (eds) The responsive brain. Wright, Bristol, pp 144-149
- Ashton H, Millman IE, Telford R, Thompson IW (1974) The effect of coffeine, nitrazepam and cigarette smoking on the contingent negative variations in man. Electroencephalogr Clin Neurophysiol 37:59-71
- Ashton H, Millman IE, Rawlins MD, Telford R, Thompson IW (1978) Stimulant and depressant effects of cigarette smoking, nicotine and certain other drugs on brain activity in man as measured by the contingent negative variation (CNV). In: Otto D (ed) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-43 397-400. Triangle Park Research Center, North Carolina

- Aunon JI, Cantor F (1977) VEP and AEP variability: interlaboratory vs intralaboratory and intersession vs introsession variability. Eelectroencephalogr Clin Neurophysiol 42:705-708
- Bancaud J (1975) Stereoelectroencephalography. In: Rémond A (ed) Handbook of electroencephalography and clinical neurophysiology, vol 10/B. Elsevier, Amsterdam
- Barbas H, Dubrovsky B, Solyom L (1978) Characteristics of the CNV in patients suffering from specific phobias. In: Otto D (ed) Multidisciplinary perspectives in eventrelated brain potential research. EPA 600/9-77-043. 369-372. Triangle Park Research Center, North Carolina
- Barlow JS (1969) Some observations on the electrophysiology of timing in the central nervous system. Electroencephalogr Clin Nuerophysiol 27:545
- Bauer H, Lauber W (1979) Operant conditioning of brain steady potential shifts in man. Biofeedback and selfregulation. 4:145–154. Plenum Press, New York
- Becker W, Kristeva R (to be published) Cerebral potentials prior to various force developments. In: Kornhuber H, Deecke L (eds) Motivation, motor and sensory processes of the brain. Progr Brain Res
- Becker W, Hoehne O, Iwase K, Kornhuber HH (1973) Cerebral and ocular muscle potentials preceding voluntary eye movements in man. Electroencephalogr Clin Neurophysiol (Suppl) 33:99-104
- Bechtereva NP (1967) On the principles of studying the neurophysiological bases of mental activity (in Russian). Vestn Akad med Nauk SSSR 1:3-11
- Bechtereva NP (1968) Some neurophysiological aspects of mental activity in man. In: Proctor D (ed) Proceedings of the International Symposium on biocybernetics of the CNS. Washington, DC, pp 63-76
- Bechtereva NP (1974) DC Changes associated with the sleep-wakefulness cycle. In: Rémond A (ed) Handbook of electroencephalography and clinical neurophysiology, vol 10/A, pp 25-32
- Besson JM, Woody CD, Aleonard P, Thompson HK, Albefessard D, Diarshall WH (1970) Correlations of brain d.c. shifts with changes in cerebral blood flow. Am J Physiol 218:284-291
- Birbaumer N, Elbert Th, Lutzenberger W, Rockstroh B (1978) Biofeedback of slow cortical potentials. Proceedings of the Biofeedback Society of America. 9th annual meeting. 210-214. Biofeedback Society of America, Denver, Colorado
- Borda RP (1970) The effect of altered drive states on the contingent negative variation (CNV) in rhesus monkey. Electroencephalogr Clin Neurophysiol 29:173-180
- Borda RP (1973) The relationship of CNV to behavior in animals: a review. Electroencephalogr Clin Neurophysiol (Suppl) 33:249-256
- Borda RP, Hablitz JJ (1973) Use of a simple visual display to reduce eye movement artefacts in CNV recordings. Electroencephalogr Clin Neurophysiol 34:433-436
- Bostem F, Rousseau JC, Degossely M, Dongier M (1967) Psychopathological correlations of the non-sepcific portion of visual and auditory evoked potentials and the associated contingent negative variation. Electroencephalogr Clin Neurophysiol (Suppl) 26:131-138
- Buchsbaum MS, Henkin RI, Christiansen RL (1974) Age and sex differences in averaged evoked responses in a normal population, with observations on patients with gonal dysgenesis. Electroencephalogr Clin Neurophysiol 37:137-144
- Callaway E, Jones RT, Layne RS (1965) Evoked responses and segmental set of schizophrenia. Arch Gen Psychiatry 12:38-39
- Cant BR, Bickford RG (1967) The effect of motivation on the contingent negative variation (CNV) (abstr). Electroencephalogr Clin Neurophysiol 23:594
- Cant BR, Pearson JE, Bickford RG (1966) The mechanism of the expectancy wave in man. Electroencephalogr Clin Neurophysiol 21:622
- Caspers H (1961) Changes of cortical DC potentials in the sleep wakefulness cycle. In: Wolstenholme GEW, O'Connor M (eds) The nature of sleep. Churchill, London, pp 237-259

- Caspers H (1963) Relations of steady potential shifts in the cortex to the wakefulnesssleep spectrum. In: Brazier MAB (ed) Brain function. University of California Press, Berkeley Los Angeles, pp 177–200
- Caspers H (1965) Shifts of the cortical steady potential during various stages of sleep. Editions du Centre National de la Recherche Scientifique (Paris) 127:213-229
- Caspers H (1974) In: Rémond A (ed) Handbook of electroencephalography and clinical neurophysiology, vol 8/A. Elsevier, Amsterdam, p 3
- Caspers H, Speckmann EJ (1970) Postsynaptische Potentiale der einzelnen Neurone und ihre Beziehungen zum EEG Z EEG EMG 1:50-65
- Caton R (1875) The electrical currents of the brain. Br Med J II:278
- Chapman RM, Bragdon HR (1964) Evoked responses to numerical and non-numerical usual stimuli while problem solving. Nature 203:1155-1157
- Cohen J (1969) Very slow brain potentials relating to expectancy: the CNV. In: Donchin E, Lindsley DB (eds) Averaged evoked potentials, methods, results, and evaluations. US Government Printing Office, Washington, DC, pp 143–198
- Cohen J (1970) Maturation of the contingent negative variation. Electroencephalogr Clin Neurophysiol 28:90-105
- Cohen J (1973) The CNV in children with special reference to learning disabilities. Electroencephalogr Clin Neurophysiol (Suppl) 33:151-154
- Cohen J, Walter WG (1966) The interaction of responses in the brain to semantic stimuli. Psychophysiology 2:187-196
- Cohen J, Offner F, Palmer C (1967a) Development of the contingent negative variation in children. Electroencephalogr Clin Neurophysiol 23:77-78
- Cohen J, Offner F, Blatt S (1967b) Psychological factors in the production and distribution of the contingent negative variation (CNV). Electroencephalogr Clin Neurophysiol. Elsevier, Amsterdam 22:190
- Cooper R, Crow HJ, Walter WG, Winter AL (1966) Regional control of cerebral vascular reactivity and oxygen supply in man. Brain Res 3:174-191
- Cooper R, Osselton JW, Shaw JC (1974) EEG technology. Butterworth, London
- Courchesne E (1977) From infancy to adulthood: the neurophysiological correlates of cognition. In: Desmedt JE (ed) Cognitive components in cerebral event-related potentials and selective attention. Progress in clinical neurophysiology, vol 6, pp 224-242. Karger, Basel
- Courchesne E, Hillyard SA, Galambos R (1975) Stimulus novelty, task relevance and the visual evoked potential in man. Electroencephalogr Clin Neurophysiol 39:131–143
- Courchesne E, Hillyard SA, Courchesne RY (1977) P3 waves to the discrimination of targets in homogeneous and heterogeneous stimuli sequences. Psychophysiology 14:590-597
- Creutzfeldt O, Kuhnt U (1966) The visual evoked potentials in normal human subjects and neurological patients. Electroencephalogr Clin Neurophysiol 20:999
- Creutzfeldt OD, Kuhnt U (1967) The visual evoked potential: physiological, developmental and clinical aspects. Electroencephalogr Clin Neurophysiol (Suppl) 26:29-41
- Davis H, Davis PA, Loomis AL, Harvey EN, Hobart G (1939) Electrical reactions of the human brain to auditory stimulation during sleep. J Neurophysiol 2:500-515
- Debecker J, Carmeliet J (1974) Automatic suppression of eye movement and muscle artefacts when averaging tape recorded cerebral evoked potentials. Electroencephalogr Clin Neurophysiol 37:513-515
- Deecke L, Kornhuber HH (1977) Cerebral potentials and the initiation of voluntary movement. In: Desmeth E (ed) Attention voluntary contraction and event-related cerebral potentials. Progress in Clinical Neurophysiology vol 1, pp 132-150. Karger, Basel
- Deecke L, Scheid P, Kornhuber HH (1969) Distribution of readiness potentials, premotion positivity and motor potentials of the human cerebral cortex preceding voluntary finger movements. Exp Brain Res 7:158-168

- Deecke L, Becker W, Grözinger G, Scheid P, Kornhuber H (1973) Human brain potentials preceding voluntary limb movements. Electroencephalogr Clin Neurophysiol (Suppl) 33:87-94
- Deecke L, Kornhuber HH, Schmidtt G (1976a) Bereitschaftspotential in Parkinson patients. In: McCallum W, Knott JR (eds) The responsive brain. Wright, Bristol, pp 169-171
- Deecke L, Grözinger B, Kornhuber HH (1976b) Voluntary finger movement in man: cerebral potentials and theory. Biol Cybern 23:99-119
- Deecke L, Englitz HG, Schmitt G (1978) Age-dependance of the Bereitschaftspotential. In: Otto D (ed) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-043. 330-332. Triangle Park Research Center, North Carolina
- Deecke L, Grözinger B, Kornhuber HH, Kriebel I, Kristeva R (1979) Cerebral potentials related to voluntary movement, speach production and respiration in man. In: Speckmann EJ, Caspers H (eds) Origin of cerebral field potentials. Thieme, Stuttgart, pp 132-140
- Demaire C, Coquery JM (1977) Effects of selective attention on the late components of evoked potentials in man. Electroencephalogr Clin Neurophysiol 42:702-704
- Desmeth JE, Debecker J (1979a) Wave form and neural mechanism of the decision P350 elicited without pre-stimulus CNV or readiness potential in random sequences of near-threshold auditory clicks and finger stimuli. Electroencephalogr Clin Neurophysiol 47:648-670
- Desmeth JE, Debecker J (1979b) Slow potential shifts and decision P350 interactions in tasks with random sequences of near-threshold clicks and finger stimuli delivered at regular intervals. Electroencephalogr Clin Neurophysiol 47:671-679
- Donchin E (1979) Event-related brain potentials: a tool in the study of human information processing. In: Begleiter H (ed) Evoked brain potentials and behavior. Plenum Press, New York, pp 13-88
- Donchin E, Otto D, Gerbrandt LK, Pribram KH (1971) While a monkey waits: electrocortical events recorded during the foreperiod of a reaction time study. Electroencephalogr Clin Neurophysiol 31:115–127
- Donchin E, Otto D, Gerbrandt LK, Pribram K (1973) Studies in the physiology of electrocortical events recorded during fore period of a reaction time study. Electroencephalogr Clin Neurophysiol 31:115-127
- Donchin E, Tueting P, Ritter Q, Kutes M, Hoffley E (1975) On the independence of the CNV and the P 300 components of the human averaged evoked potential. Electroencephalogr Clin Neurophysiol 38:449-461
- Donchin E, Callaway E, Cooper R, Desmet JE, Goff WR, Hillyard SA, Sutton S (1977) Publication criteria for studies of evoked potentials (EP) in man. In: Desmet E (ed) Attention voluntary contraction and event-related cerebral potentials. Karger, Basel, pp 1-11
- Dongier M, Bostem F (1967) Essais d'application en psychiatrie de la variation contingente négative. Acta Neurol Belg 67:640-645
- Dongier M, Koninckx N (1970) Present-day neurophysiological models of mind-body interaction. Psychother Psychosom 18:123-129
- Dongier M, Timsit-Berthier M, Koninckx N, Delanoy J (1973) Compared clinical significance of CNV and other slow potential changes in psychiatry. Electroencephalogr Clin Neurophysiol (Suppl) 33:321-326
- Dubrovsky B, Garcia-Rill E, Dongier M (1973) Effect of hyperventilation on abnormally prolonged CNV. Electroencephalogr Clin Neurophysiol 34. Extra number, p 740
- Dustman RE, Beck EC (1969) The effect of maturation and aging on the wave form of the visual evoked potentials. Electroencephalogr Clin Neurophysiol 26:2-11
- Ellingson RJ, Lathrop GH, Danahy T, Nelson B (1973) Variability of visual evoked potentials in human infants and adults. Electroencephalogr Clin Neurophysiol 34: 113-124

- Ervin FR, Mark VH (1964) Studies of the human thalamus. IV. Evoked responses. Ann NY Acad Sci 112:18-92
- Faidherbe J, Deliege M, Devaux-Pasque N (1969) Manuelles recherches sur la signification psychophysiologiques de certaines composantes lentes de potentiels evoques électroencéphalographiques: contribution à une psychophysiologie de la potensité.
  In: Dargent J, Dongier M (eds) Variations contingentes négatives. Congrès et Colloques de l'Université de Liège, vol 52, pp 114-137
- Fenelon B (1968) Expectancy waves and other complex cerebral events in dyslexic and normal subjects. Psychosom Sci 13:253-254
- Ferguson JH, Jasper HM (1971) Laminar D.C. studies of acetylcholine activated epileptiform discharge in cerebral cortex. Electroencephalogr Clin Neurophysiol 30: 377-390
- Ford JM, Roth WT, Dinko SJ, Kopell BS (1973) Evoked potential correlates of signal recognition between and within modalities. Science 181:465–466
- Friedman J, Goldberg H, Horvath TB, Meares RA (1974) The effect of tobacco smoking on evoked potentials. Clin Exp Pharmacol Physiol 1:249–258
- Fukushima T, Mayanagi Y, Bouchard G (1976) Thalamic evoked potentials to somatosensory stimulation in man. Electroencephalogr Clin Neurophysiol 40:481-490
- Ganglberger JA (1962) Wirkungen umschriebener Reizungen im menschlichen Zwischenhirn auf das EMG. IIIrd Neurobiological Meeting Kiel
- Ganglberger JA, Gestring GF, Groll-Knapp E, Haider M (1971) Auditory evoked potentials from cortex and various subcortical centers. Arch Klin Exp Ohr Nas Kehlk Heilkd 198:41-49
- Ganglberger JA, Haider M, Knapp E, Schmid H (to be published) Subdural recordings of the cortex-motor potential. In: Kornhuber H, Deecke L (eds) Motivation, motor and sensory processes of the brain. Progr Brain Res
- Geddes LA (1972) Electrodes and the measurement of bioelectric events. Wiley, New York
- Gerbrandt LK, Goff WR, Smith DB (1973) Distribution of the human average movement potentials. Electroencephalogr Clin Neurophysiol 34:461-474
- Gilden L, Vaughan HG Jr, Costa LD (1966) Summated human EEG potentials with voluntary movement. Electroencephalogr Clin Neurophysiol 20:433-438
- Girton DG, Kamiya J (1973) A simple on-line technique for removing eye movement artefacts from the EEG. Electroencephalogr Clin Neurophysiol 34:212-216
- Glötzner F, Grüsser OJ (1968) Membran Potential und Entladungsfolgen corticaler Zellen, EEG und corticales Bestandspotential bei generalisierten Krampfanfällen. Arch Psychiatr Nervenkr 210:313-339
- Goldring S (1974) DC shifts released by direct afferent stimulation. In: Rémond A (ed) Handbook of electroencephalography and clinical neurophysiology, vol 10/A. Elsevier, Amsterdam, pp 12–18
- Groll-Knapp E, Ganglberger JA, Haider M (1968) Thalamo-corticale Wechselwirkungen bei zentralen Orientierungsreaktionen. In: Irle M (ed) Bericht über den 26. Kongr d Deutschen Gesellschaft für Psychologie. Hogrefe, Göttingen, pp 195–198
- Groll-Knapp E, Ganglberger JA, Haider M (1970) Relations between thalamic, cortical and subperiostal derivations of expectancy waves and orienting potentials. Electroencephalogr Clin Neurophysiol 28:322-325
- Groll-Knapp E, Wagner H, Hauck H, Haider M (1972) Effects of low carbonmonoxide concentrations on vigilance and computer-analysed brain potentials. Staub Reinhalt Luft 32:64-68
- Groll-Knapp E, Ganglberger JA, Haider M (1977) Voluntary movement-related slow potentials in cortex and thalamus in man. In: Desmeth JE (ed) Attention, voluntary contraction and event-related cerebral potentials. Prog Clin Neurophysiol, vol 1, Karger, Basel, pp 164-173
- Groll-Knapp E, Haider M, Höller H, Jenkner G, Neuberger M, Stidl H (1978) Neuroand psychophysiological effects of moderate CO doses. In: Otto D (ed) Multidisci-

plinary perspectives in event-related brain potential research EPA 600/9-77-043. Triangle Park Research Center, North Carolina, pp 424-430

- Grossman RG, Hampton T (1968) Depolarization of cortical glia cells during electrocortical activity. Brain Res 11:316-324
- Grünewald G, Grünewald-Zuberbier E, Netz J (1978) Late components of average evoked potentials in children with different abilities to concentrate. Electroencephalogr Clin Neurophysiol 44:617-625
- Grünewald G, Grünewald-Zuberbier E, Netz J, Hömberg V, Sander G (1979a) Relationships between the late component of the contingent negative variation and the Bereitschaftspotential. Electroencephalogr Clin Neurophysiol 46:538-545
- Grünewald G, Grünewald-Zuberbier E, Hömberg V, Netz J (1979b) Cerebral potentials during smooth goal-directed hand movements in right-handed and left-handed subjects. Pfluegers Arch 381:39-46
- Grünewald-Zuberbier E, Grünewald G (1978) Goal-directed movement potentials of human cerebral cortex. Exp Brain Res 33:135-138
- Grünewald-Zuberbier E, Grünewald G, Jung R (1978a) Slow-potentials of the human precentral and parietal cortex during goal-directed movements (Zielbewegungspotentiale). J Physiol 284:181-182
- Grünewald-Zuberbier E, Grünewald G, Rasche A, Netz J (1978b) Contingent negative variation and alpha attenuation responses in children with different abilities to concentrate. Electroencephalogr Clin Neurophysiol 44:37-47
- Gullickson GR (1970) The contingent negative variation in the preschool child. Ph D Dissertation, Institute of Technology, Illinois
- Gullickson GR (1973a) CNV and behavioral attention to a glide-tone warning of interesting non-moving or kaleidoscopic visual or auditory patterns in 2 and 3-years-old children. Electroencephalogr Clin Neurophysiol 33:145-150
- Gullickson GR (1973b) Ontogenesis of readiness and motor potentials. Electroencephalogr Clin Neurophysiol 34:106
- Gumnit RJ (1960) D.C. potential changes from auditory cortex of cat. J Neurophysiol 23:667–675
- Gumnit RJ (1961) The distribution of D.C. responses evoked by sounds in the auditory cortex of the cat. Electroencephalogr Clin Neurophysiol 13:889-895
- Hablitz JS (1973) Operant conditioning and slow potential changes form monkey cortex. Electroencephalogr Clin Neurophysiol 34:399–408
- Hablitz JJ, Borda RP (1973) The effects of Dalmane (flurazepam hydrochloride) on the contingent negative variation. Electroencephalogr Clin Neurophysiol 33:317– 320
- Haider M (1965) Erwartungswirkung, Antizipation und Cortical Evozierte Potentiale. In: 6th International Congress of Electroencephalography and Clinical Neurophysiology, Vienna, pp 167–168
- Haider M (1969) Elektrophysiologische Indikatoren der Aktivierung. In: Schönpflug W (ed) Methoden der Aktivierungsforschung. Huber, Bern Stuttgart, pp 125–156
- Haider M (1970) Neuropsychology of attention, expectation and vigilance. In:

Mostofsky DI (ed) Attention: contemporary theories and analysis. Appleton-Century-Crofts, New York, pp 419-430

- Haider M, Groll-Knapp E (1971) Psychophysiologische Belastung eines Musikers in einem Symphonieorchester. In: Piperek M (ed) Stress und Kunst. Braumüller, Wien, pp 15-37
- Haider M, Ganglberger JA, Groll-Knapp E (1968a) Neuropsychologische Untersuchungen über Erwartungsvorgänge und ihre thalamo-corticalen Reaktionen. 26. Kongress der Deutschen Gesellschaft für Psychologie, Göttingen, pp 189–195
- Haider M, Groll-Knapp E, Studynka G (1968b) Orientierungs- und Bereitschaftspotentiale bei unerwarteten Reizen. Exp Brain Res 5:45-54
- Haider M, Ganglberger JA, Gestring GF, Groll-Knapp E (1968c) Zentralnervöse Steuerungsmechanismen für Erwartungsvorgänge und sensomotorische Integration. Jahrestagung der Deutschen Gesellschaft für Biophysik, Verlag der Wr Med Akad, pp 49–51

- Haider M, Ganglberger JA, Groll-Knapp E (1972) Computer analysis of subcortical and cortical evoked potentials of slow potential phenomena in humans. Confin Neurol 34:224-229
- Haider M, Ganglberger JA, Groll-Knapp E, Schmid H (1979) Averaged cortical and subcortical potentials during stereotactic operations in humans. In: Speckmann E, Caspers H (eds) Origin of cerebral field potentials. Thieme, Stuttgart, pp 141–150
- Harter MR, Salomon LE (1972) Intra modality selective attention and evoked potentials to randomly presented patterns. Electroencephalogr Clin Neurophysiol 32: 605-613
- Hassler R (1959) Anatomie des Thalamus Anatomy of the thalamus. In: Schaltenbrand G, Bailey P (eds) Einführung in die stereotaktischen Operationen mit einem Atlas des menschlichen Gehirns, vol I. Thieme, Stuttgart, pp 230–290
- Hassler R (1978) Striatal control of locomotion, intentional actions and of integrating and perceptive activity. J Neurol Sci 36:187–224
- Hazemann PS, Metral S, Lille F (1978) Influence of physical parameters of movement (force, speed and duration) upon slow cortical potentials in man. In: Otto D (ed) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-043. Triangle Park Research Center, North Carolina, pp 107-111
- Hillyard SA (1969) The CNV and the vertex evoked potential during signal detection. A preliminary report. In: Donchin E, Lindsley DB (eds) Averaged evoked potentials, methods, results and evaluation. NASA-SP-191. US Government Printing Office, Washington DC, pp 349-353
- Hillyard SA (1971) The psychological specificity of the contingent negative variation and late evoked potential (P 300). Electroencephalogr Clin Neurophysiol 31:302– 303
- Hillyard SA (1973) The CNV and human behavior (a review). Electroencephalogr Clin Neurophysiol 33:161-171
- Hillyard SA, Galambos R (1967) Effects of stimulus and response contingencies on a surface negative slow potential shift in man. Electroencephalogr Clin Neurophysiol 22:297-304
- Hillyard SA, Galambos R (1970) Eye movement artefacts in the CNV. Electroencephalogr Clin Neurophysiol 28:173-182
- Hillyard SA, Hink RF, Schwent UL, Pieton TW (1973) Electrical signs of selective attention in the human brain. Science 182:177-180
- Iliukina WA (1977) Slow-bio-electric events in the human brain (in russian). Academy of Sciences, USSR
- Irwin DA, Knott JR, McAdam DW, Rebert CS (1966) Motivational determinants of the "contingent negative variation". Electroencephalogr Clin Neurophysiol 21: 538-543
- Järvilehto T, Fruhstorfer H (1970) Differentiation between slow cortical potentials associated with motor and mental acts in man. Exp Brain Res 11:309–317
- Jasper HH (1958) The ten twenty electrode system of the International Federation of Societies for Electroencephalography and Clinical Neurophysiology. Electroencephalogr Clin Neurophysiol 10:370--375
- Jasper HH (1960) Unspecific thalamo-cortical relations. In: Magoun HW, Hall VE (eds) Handbook of physiology, sect 1 (Neurophysiology, vol 2). American Physiological Society, Washington, DC, pp 1307-1321
- Jus K, Kiljan A, Kubacki A, Rzepecki J, Wilczsak H, Jus A (1968) Studies on the effect of psychological factors on the expectancy wave. Pol Med J 7:1235-1241
- Karahashi Y, Goldring S (1966) Intracellular potentials from "idle" cells in cerebral cortex of cat. Electroencephalogr Clin Neurophysiol 20:600-607
- Karlin L (1970) Cognition, preparation and sensory evoked potentials. Psychol Bull 73:122-126
- Karlin L, Martz MJ (1973) Response probability and sensory evoked potentials. In: Kornblum S (ed) Attention and performance IV. Academic Press, New York, pp 25-33

- Karlin L, Martz, MJ, Mordkoff AM (1970) Motro performance and sensory evoked potentials. Electroencephalogr Clin Neurophysiol 28:307–313
- Karrer R, Ivins J (1975) Steady potentials accompanying perception and response in mentally retarded and normal children. In: Karrer R (ed) Developmental psychophysiology of mental retardation. Thomas, Springfield, Ill, pp 361-417
- Karrer R, Ivins J (1976) Post-warning signal positivity in relation to development, RT performance and warning signal compunding. In: McCallum WC, Knott JR (eds) The responsive brain. Wright, Bristol, pp 132-135
- Karrer R, Warren C, Ruth R (1978) Slow potentials of the brain preceding cued and noncued movements: effects of development and mental retardation. In: Otto D (ed) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-043. Triangle Park Research Center, North Carolina, pp 322-329
- Klinke R, Fruhstorfer H, Finkenzeller P (1968) Evoked responses as a function of external and stored information. Electroencephalogr Clin Neurophysiol 25:119–122
- Klorman R (1975) Contingent negative variation and cardiac deceleration in a long preparatory interval: a developmental study. Psychophysiology 12/4:609-617
- Knapp E, Schmid H, Ganglberger JA, Haider M (to be published) Cortical and subcortical potentials during goal directed and serial goald directed movements in humans. In: Kornhuber HH, Deecke L (eds) Motivation, motor and sensory processes of the brain. Prog Brain Res
- Knott JR, Irwin DA (1968) Anxiety, stress and the contingent negative variation (CNV). Electroencephalogr Clin Neurophysiol 24:286–287
- Knott JR, Irwin DA (1973) Anxiety, stress and the contingent negative variation. Arch Gen Psychiatry 29:538-541
- Knott JR, Peters JF (1974) Changes in CNV amplitude with progressive induction of stress as a function of sex. Electroencephalogr Clin Neurophysiol 36:47-51
- Köhler W, Held R (1949) The cortical correlate of pattern vision. Science 110:414-419
- Köhler W, Held R, O'Connel DW (1952) An investigation of cortical currents. Proc Am Philos Soc 96:290–330
- Kok A (to be published) Changes in components of the event-related potential (ERP) during visual recognition: effects of stimulus degradation and stimulus familiarity.
   In: Kornhuber HH, Deecke L (eds) Motivation, motor and sensory processes of the brain. Progr Brain Res
- Kooi KA, Bagchi BK (1964) Visual evoked responses in man: normative data. Ann NY Acad Sci 112:270–280
- Kopell BS, Tinklenberger JR, Hollister LW (1972) Contingent negative variation amplitudes: marihuana and alcohol. Arch Gen Psychiatry 27:809-811
- Kornhuber HH, Deecke L (1965) Hirnpotentialänderungen bei Willkürbewegungen und passiven Bewegungen des Menschen: Bereitschaftspotential und reafferente Potentiale. Arch Ges Physiol 284:1-17
- Kornhuber HH, Becker W, Träumer R, Hoehne O, Iwase K (1969) Cerebral potentials accompanying voluntary movements in man: readiness potential and reafferent potentials (abstr). Electroencephalogr Clin Neurophysiol 26:439
- Kuffler SW, Nicholls JG (1966) The physiology of neuroglia. Ergeb Physiol 57:1-90
- Kurtzberg D, Vaughan HG, Kreuzer J (1977) Task related cortical potentials in children. In: Desmedt JE (ed) Cognitive components in cerebral event-related potentials and selective attention. Prog Clin Neurophysiology, vol 6, Karger, Basel,
- pp 216-223 Kutas M, Donchin E (1977) The effect of handedness of the responding hand and the response force on the contralateral dominance of the readiness potential. In: Desmeth E (ed) Attention, voluntary contraction and event-related cerebral potentials. Prog Clin Neurophysiol, vol 1, Karger, Basel, pp 189-210
- Lancry A (1976) Variation contingente négative chez l'enfant de 10 ans. Rélatin avec le type de performance dans une tâche de localisation spatiale. Electroencephalogr Clin Neurophysiol 40:561-570

- Lelord G, Laffont F, Sauvage D, Jusseaume P (1973) Etude chez l'homme d'activités evoquées lentes succédant au movement volontaire et au language articulé. Electroencephalogr Clin Neurophysiol 35:113–124
- Lewis PR, Shute CCD (1967) The cholinergic limbic system: projections to hippocampal formation, medial cortex, nuclei of ascending cholinergic reticular system and the subfornical organ and supra-optic crest. Brain 90:521-540
- Lifschitz K (1966) The average evoked cortical response to complex visual stimuli. Psychophysiology 3:55-68
- Lindsley DB (1969) Neurophysiological mechanisms of attention and perception: their role in information processing in the visual system. In: Kimble DP (ed) Readiness to remeber, vol 2, Gordar & Breach, New York, pp 345-507
- Loveless NE, Sanford AJ (1974) Slow potential correlates of preparatory set. Biol Psychology 1:303-314
- Loveless N, Sanford A (1974a) Effects of age on the contingent negative variation and preparatory set in a reaction time task. J Gerontol 29:52-63
- Low MD (1969) Discussion of: Application a la pathologie. In: Dargent J, Dongier M (eds) Variations contingentes négatives. Congrès et colloques de l'Université de Liège, vol 52, p 163
- Low MD, McSherry JW (1968) Further observations of psychological factors involved in CNV genesis. Electroencephalogr Clin Neurophysiol 25:203-207
- Low MD, Stoilen L (1973) CNV and EEG in children: maturational characterisitcs and findings in the MCD syndrome. Electroencephalogr Clin Neurophysiol 33:139-143
- Low MD, Borda RP, Frost JD Jr, Kellaway P (1966a) Surface-negative slow-potential shift associated with conditioning in man. Neurology 16:771-782
- Low MD, Borda RP, Kellaway P (1966b) "Contingent negative variation" in rhesus monkeys: an EEG sign of a specific mental process. Percept Mot Skill 22:443-446
- Low MD, Coats AC, Rettig GM, McSherry JW (1967) Anxiety, attentiveness-altertness: a phenomenological study of the CNV. Neuropsychologia 5:379–384
- Low MD, Klonoff H, Marcus A (1973) The neurophysiological basis of the marihuana experience. Can Med Assoc J 108:157-165
- Lüders H (1970) The effects of aging on the wave form of the somatosensory cortical evoked potential. Electroencephalogr Clin Neurophysiol 29:450-460
- Marczynski TJ (1972) Electrophysiological correlates of positive reinforcement: postreinforcement synchronization, modulation of sensory input, and steady potentials. In: Karczmar AG, Eccles JC (eds) Brain and human behavior, Springer, Berlin Heidelberg New York, pp 303-321
- Marczynski TJ, Hackett CJ, Sherry CJ, Allen SL (1971) Diffuse light input and quality of reward determine the occurrence of "reward contingent positive variation" (RCPV) in cat. Brain Res 28:57-70
- Marsh GR (1978) Aging and average evoked potentials. In: Otto D (ed) Multidisciplinary perspectives in event-related potential research. EPA 600/9-77-043. Triangle Park Research center, North Carolina, pp 333-336
- Marsh GR, Thompson LW (1973) Effect of verbal and non-verbal psychological set on hemispheric asymmetries in the CNV. Electroencephalogr Clin Neurophysiol 33: 195-200
- McAdam DW, Rubin EH (1971) Readiness potential, vertex positive wave, contingent negative variation and accuracy of perception. Electroencephalogr Clin Neurophyiol 30:511-517
- McAdam DW, Seales DM (1969) Bereitschaftspotential enhancement with increased level of motivation. Electroencephalogr Clin Neurophysiol 27:73-75
- McAdam DW, Whittaker HA (1971) Language production: electroencephalographic localization in the normal human brain. Science 172:499–502
- McCallum WC (1966) The EEG: has it a futur in social and clinical psychology? Proc Univ London Psychol Soc, Nat Psychol Conf, London, 16 pages
- McCallum WC (1967a) New waves in the brain. New Scientist 36:592-594

- McCallum WC (1967b) The contingent negative variation: an experimental study of a slow potential change in the electroencephalogram of normal subjects and psychiatric patients. Ph D Dissertation, University of Bristol
- McCallum WC (1969) The contingent negative variation as a cortical sign of attention in man. In: Evans CR, Mullholland TB (eds) Attention in neurophysiology. Butterworths, London, pp 40–63
- McCallum WC (1978) Relationships between Bereitschaftspotential and contingent negative variation. In: Otto D (ed) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-043. Triangle Park Research Center, North Carolina pp 124-130
- McCallum WC, Abraham P (1973) The contingent negative variation in psychosis. Electroencephalogr Clin Neurophysiol 33:329-335
- McCallum WC, Cummins B (1973) The effect of brain lesion on the contingent negative variation in neurosurgical patients. Electroencephalogr Clin Neurophysiol 35: 449-456
- McCallum WC, Knott JR (1973) Event-related slow potentials of the brain: their relations to behavior. Electroencephalogr Clin Neurophysiol 33:390
- McCallum WC, Walter WG (1968) The effects of attention and distraction on the contingent negative variation in normal and neurotic subjects. Electroencephalogr Clin Neurophysiol 25:319-329
- McCallum WC, Walter WG, Winter A, Scotton L, Cummins B (1970) The contingent negative variations in cases of known brain lesions. Electroencephalogr Clin Neurophysiol 28:210
- McCallum WC, Papakostopoulos D, Griffith HB (1976) Distribution of CNV and other slow potential changes in human brainstem structures. In: McCallum WC, Knott JR (eds) The responsive brain. Wright, Bristol, pp 205-211
- McSherry JW (1973) Physiological origins: a review. Electroencephalogr Clin Neurophysiol 33:53-61
- McSherry JW, Borda RP (1973) The intracortical distribution of the CNV in rhesus monkey. Electroencephalogr Clin Neurophysiol 33:69-74
- Miller LH, van Veen W, Sandman CA, Knott JR (1973) Intermodal and intramodal distraction and CNV. Electroencepahlogr Clin Neurophysiol (Suppl) 33:209-211
- Naitoh P, Hilbert RP (1976) CNV-heart rate response under gradual sleep reduction. In: McCallum WC, Knott JR (eds) The responsive brain. Wright, Bristol, pp 60-63
- Naitoh P, Johnson LC, Lubin A (1971) Modification of surface negative slow potential (CNV) in the human brain after total sleep loss. Electroencephalogr Clin Neurophysiol 30:17-22
- O'Leary JL, Goldring S (1964) DC-potentials of the brain. Physiol Rev 44:91-125
- Otto D (ed) (1978) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-043. Triangle Park Research Center, North Carolina
- Otto DA, Leifer LH (1973) The effect of modifying response and performance feedback parameters on the CNV in humans. Electroencephalogr Clin Neurophysiol (Suppl) 33:29-37
- Otto D, Houck K, Finger H, Hart S (1976) Event-related slow potentials in aphasic, dyslexic and normal children during pictorial and letter-matching. In: McCallum WC, Knott JR (eds) The responsive brain. Wright, Bristol, pp 172-177
- Papakostopoulos D (1978) The present state of brain macropotentials in motor control research: a summary of issues. In: Otto D (ed) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-043. Triangle Park Research Center, North Carolina, pp 77-83
- Papakostopoulos D, Crow HJ (1976) Electrocorticographic studies of the contingent negative variation and "P 300" in man. In: McCallum WC, Knott JR (eds) The responsive brain. Wright, Bristol, pp 201-204
- Papakostopoulos D, Fenelon B (1975) Spatial distribution of the contingent negative variation (CNV) and the relationship between CNV and reaction time. Psychophysiology 12:74-78

- Papakostopoulos D, Winter A, Newton P (1973) New techniques for the cntrol of eye potential artefacts in multichanal CNV recordings. Electroencephalogr Clin Neurophysiol 34:651-653
- Paini M, Zappoli R (1973) The "expectancy wave" (CNV) in normal children. Electroencephalogr Clin Neurophysiol 35:438
- Paul DD, Sutton S (1972) Evoked potential correlates of response cirteria in auditory signal detection. Science 177:362-364
- Peters JF, Knott JR, Miller LH, van Veen WJ, Cohen S (1970) Response variables and magnitude of the contingent negative variation. Electroencephalogr Clin Neurophysiol 29:608-611
- Pfurtscheller G, Aranibar A (1977) Event-related cortical desynchronization detected by power measurements of scalp EEG. Electroencephalogr Clin Neurophysiol 42: 817-826
- Pfurtscheller G, Aranibar A (to be published) Changes in the central EEG activity in relation to voluntary movement. 1. Normal subjects. In: Kornhuber H, Deecke L (eds) Motivation, motor and sensory processes of the brain. Prog Brain Res
- Picton TW, Hillyard SA (1974) Human auditory evoked potentials. II. Effects of attention. Electroencephalogr Clin Neurophysiol 36:191–199
- Poon LW, Thompson LW, Williams RB, Marsh GR (1974) Changes of anterior-posterior distribution of CNV and late positive components as a function of information processing demands. Psychophysiology 11:660-673
- Rebert CS (1972) Cortical and subcortical slow potentials in the monkey's brain during a preparatory interval. Electroencephalogr Clin Neurophysiol 33:389-402
- Rebert CS (1973) Elements of a general cerebral system related to CNV genesis. Electroencephalogr Clin Neurophysiol 33:63-67
- Rebert CS (1976) Slow potential changes in the monkey's brain during reaction time foreperiod. In: McCallum WC, Knott JR (eds) The responsive brain. Wright, Bristol, pp 191–194
- Rebert CS, Irwin DA (1973) Simple electrode configurations for chronic or acute recording of d.c. potentials from subcortical nuclei of the brain. Electroencephalogr Clin Neurophysiol 34:440-442
- Rebert CS, Knott JR (1970) The vertex non-specific evoked potential and latency of contingent negative variation. Electroencephalogr Clin Neurophysiol 28:561-565
- Rebert CS, Sperry KG (1973) Subjective and response-related determinants of CNV amplitude. Psychophysiology 10:139-144
- Rebert CS, McAdam DW, Knott JR, Irwin DA (1967) Slow potential change in human brain related to level of motivation. J Comp Physiol Psychol 63:20–23
- Ritter W, Vaughan HG (1969) Average evoked responses in vigilance and discrimination: areassessment. Science 164:326-328
- Ritter W, Vaughan HG (to be published) Topographic analysis of auditory ERP's. In: Kornhuber H, Deecke L (eds) Motivation, motor and sensory processes of the brain. Prog Brain Res
- Ritter W, Vaughan HH,Costa LD (1968) Orienting and habituation to auditory stimuli: a study of short term changes in average evoked responses. Electroencephalogr Clin Neurophysiol 25:550-556
- Roth WT, Kopell BS, Tinklenberg JR, Huntsberger GE, Kraemer HC (1975) Reliability of the contingent negative variation and the auditory evoked potential. Electroencephalogr Clin Neurophysiol 38:45-50
- Roth WT, Ford JM, Lewis SJ, Kopell BS (1976) Effects of stimulus probability and task relevance of event-related potentials. Psychophysiology 13:311-317
- Roth WT, Tinklenberg JR, Kopell BS (1977) Ethanol and marihuana effects on eventrelated potentials in a memory retrieval paradigm. Electroencephalogr Clin Neurophysiol 42:381-388
- Routtenberg A (1968) The two-arousal hypothesis: reticular formation and limbic system. Physiol Rev 75:51-80

- Rowland V (1960) Electrographic responses in sleeping conditioned animals. In: Wolstenholme GEW, O'Connor M (eds) The nature of sleep. Little, Brown, Boston, Ma, pp 284-306
- Rowland V (1967) Steady potential phenomena of cortex. In: Quarton GC et al. (eds) The neurosciences: a study program. Rockefeller University Press, New York, pp 482-498
- Rowland V (1968) Cortical steady potential (direct current potential) in reinforcement and learning. In: Stellar E, Sprague JM (eds) Progress in physiological psychology, vol 2. Academic Press, London New York, pp 2–78
- Ruchkin DS, Sutton S (1973) Visual evoked and emitted potentials and stimulus significance. Bull Psychos Soc 2:144–146
- Ruchkin D, Sutton S, Tueting P (1975) Emitted and evoked potentials and variation in stimulus probability. Psychophysiology 12:591–599
- Rusinov VS (1959) Electroencephalographic studies in conditioned reflex formation in man. In: Brazier MAB (ed) The central nervous system and behavior. Second Macy Conference, Madison, NJ, pp 249-256
- Shagass C, Schwartz M (2962) Evoked cortical potentials and sensation in man. J Neuropsychiatr 2:262-270
- Shute CC, Lewis PR (1967) The ascending cholinergic reticular system: neocortical olfactory and subcortical projections. Brain 90:497-520
- Skinner JE (1971) Abolition of a conditioned surface-negative, cortical potential during cryogenic blockade of the non-specific thalamocortical system. Electroencephalogr Clin Neurophysiol 31:197-209
- Skinner JE, Lindsley DB (1973) The nonspecific mediothalamic frontocortical system: its influence on electrocortical activity and behavior. In: Pribram K, Luria A (eds) Psychophysiology of the frontal lobes. Academic Press, New York, pp 85-97
- Skinner JE, Yingling CD (1977) Central gating mechanisms that regulate event-related potentials and behavior. A neural model for attention. In: Desmeth JE (ed) Progress in clinical neurophysiology, vol 1. Karger, Basel, pp 30–69
- Small JG, Small IF (1970) Interrelationships of evoked and slow potential responses. Dis Nerv Syst 31:459-464
- Small JG, Small IF (1971) CNV correlations with psychiatric diagnosis. Arch Gen Psychiatry 25:550-554
- Smith DBD, Donchin E, Cohen L, Starr A (1970) Auditory evoked potentials in man during selective binaural listening. Electroencephalogr Clin Neurophysiol 28:146– 152
- Soskis DA, Shagass C (1974) Evoked potential tests of augmenting-reducing. Psychophysiology 11:175-190
- Speckmann EJ, Caspers H (1979) Cortical field potentials in relation to neuronal activities in seizure conditions. In: Speckmann EJ, Caspers H (eds) Origin of cerebral field potentials. Thieme, Stuttgart, pp 205-213
- Speckmann EJ, Caspers H, Janzen RW (1972) Relations between cortical DC shifts and membrane potential changes of cortical neurones associated with seizure activity. In: Petsche H, Brazier MAB (eds) Synchronization of EEG activity in epilepsies. Springer, Wien New York, pp 93-111
- Squires KC, Hillyard SA, Lindsay PH (1973) Vertex potentials evoked during auditory signal detection: relation to decision criteria. Percept Psychophys 13:25-31
- Squires KC, Donchin E, Hering R, McCarthy G (1977) On the influence of task relevance and stimulus probability on event-related potential changes. Electroencephalogr Clin Neurophysiol 42:1–14
- Squires NK, Squires KC, Hillyard SA (1975) Two varities of long-latency positiv waves evoked by unpredictable auditory stimulus. Electroencephalogr Clin Neurophysiol 38:387-401
- Storm von Leeuwen W (1975) Auditory evoked responses (AERs) In: Rémond A (ed) Handbook of electroencephalography and clinical neurophysiology, vol 8/A. Elsevier, Amsterdam, pp 72ff

- Straumanis JJ, Shagass C, Overton DA (1969) Problems associated with application of the contingent negative variation to psychiatric research. J Nerv Ment Dis 148: 170-179
- Sutton S (1979) P 300 Thirteen years later. In: Begleiter H (ed) Evoked brain potentials and behavior. Plenum Press, New York pp 107–126
- Sutton S, Barren M, Zubin J, John ER (1965) Evoked potentials correlates of stimulus uncertainty. Science 150:1187–1188
- Sutton S, Tueting P, Zubin J, John ER (1967) Information delivery and the sensory evoked potential. Science 155:1436-1439
- Symmes D, Eisengart MA (1971) Evoked response correlates of meaningfull visual stimuli in children. Psychophysiology 8:769-778
- Tecce JJ (1972) Contingent negative variation (CNV) and psychological processes in man. Psychol Bull 77:73-108
- Tecce JJ, Cole JO (1974) Amphetamine effects in man: paradoxical drowsiness and lowered electrical brain activity (CNV). Science 185:451-453
- Tecce JJ, Hamilton BT (1973) CNV reduction by sustained cognitive activity (distraction). Electroencephalogr Clin Neurophysiol 33:229-237
- Tecce JJ, Savignano-Bowman J, Meinbresse D (1976) Contingent negative variation and the distraction-arousal hypothesis. Electroencephalogr Clin Neurophysiol 41: 277-286
- Thompson LW, Nowlin JB (1971) Cortical slow potential and cardiovascular correlates of attention during reaction time performance. In: Javik L, Eisdorfer C, Blum J (eds) Aging: psychological and somatic changes. Springer, Berlin Heidelberg New York, pp 113-121
- Thompson JW, Newton P, Pocock PV, Cooper R, Crow H, McCallum WC, Papakostopoulos D (1978) A preliminary study of the pharmacology of the contingent negative variation (CNV) in man. In: Otto D (ed) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-043. Triangle Park Research Center, North Carolina, pp 51-55
- Timsit-Berthier M, Hausman J (1972) Etude de la CNV et du phénomène de préparation motrice chez des enfants de 5-15 ans. Rev EEG 2:141-146
- Timsit-Berthier M, Hausman J (1973) Study of the contingent negative variation (CNV) and the motor preparation phenomenon in children aged 5-15 years. Electroencephalogr Clin Neurophysiol 34:217
- Timsit M, Koninckx N, Dargent J, Fontaine O, Dongier M (1970) Variations contingentes négatives en psychiatrie. Electroencephalogr Clin Neurophysiol 28:41-47
- Timsit-Berthier M, Rousseau JC, Delaunoy J (1971) Reactivite de l'onde d'attente et des ondes negatives post-imperatives. Rev EEG Neurophysiol 1:245-248
- Timsit-Berthier M, Delaunoy J, Koninckx N, Rousseau JC (1973) Slow potential changes in psychiatry. I. Contingent negative variation. Electroencephalogr Clin Neurophysiol 35:355-361
- Timsit-Berthier M, Delaunoy J, Xhenseval B, Timsit M (1978) Reliability of the CNV in psychopathology. In: Otto D (ed) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-043. Triangle Park Research Center, North Carolina, pp 373-375
- Tsubokawa T, Moriyasu N (1978) Motivational slow negative potential shift (CNV). Related to thalamotomy. Appl Neurophysiol 41:202-208
- Vaughan HG Jr (1969) The relationship of brain activity to scalp recordings of eventrelated potentials. In: Donchin E, Lindsley DB (eds) Average evoked potentials, methods, results and evaluations. NASA-SP-191. US Government Printing Office, Washington, DC, pp 45-94
- Vaughan HG (1975) Psychological correlates of the association cortex potentials. In: Rémond A (ed) Handbook of electroencephalography and clinical neurophysiology, vol 8/A. Elsevier, Amsterdam, pp 92-101
- Vaughan HG Jr, Costa LD (1968) Analysis of electroencephalographic correlates of human sensorimotor processes. Electroencephalogr Clin Neurophysiol 24:288-289

- Vaughan HG, Ritter W (1970) The sources of auditory evoked responses recorded from the human scalp. Electroencephalogr Clin Neurophysiol 28:360–367
- Vaughan HG Jr, Costa LD, Ritter W (1968) Topography of the human motor potential. Electroencephalogr Clin Neurophysiol 25:1-10
- Velasco M, Velasco F, Maldonado H, Machado PJ (1975) Differential effect of thalamic and subthalamic lesions on early and late components of the somatic evoked potentials in man. Electroencephalogr Clin Neurophysiol 39:163-171
- Walter WG (1964) Slow potential waves in the human brain associated with expectancy attention and decision. Arch Psychiatry 206:309–322
- Walter WG (1966a) Electrophysiologic contributions to psychiatric therapy. In: Masserman J (ed) Current psychiatric therapies, vol VI. Grune & Stratton, New York, pp 13-25
- Walter WG (1966b) The relations between electrical signs of expectancy in the human brain and autonomic function during operant and classical conditioning. Ab Dtsch Akad Wiss Berlin (Kl Med) 2:47–55
- Walter WG (1967a) Electrical signs of association, expectancy and decision in the human brain. Electroencephalogr Clin Neurophysiol 23:258-263
- Walter WG (1967b) Slow potential changes in the human brain associated with expectancy, decision and intention. Electroencephalogr Clin Neurophysiol (Suppl) 26: 123-130
- Walter WG (1975) Evoked responses general. In: Rémond A (ed) Handbook of electroencephalography and clinical neurophysiology, vol 8/A. Elsevier, Amsterdam, pp 20-32
- Walter WG, Cooper R, Aldridge VJ, McCallum WC, Winter AL (1964) Contingent negative variation: an electric sign of sensorimotr association and expectancy in the human brain. Nature 203:380-384
- Wasman M, Morehead SD, Lee H, Rowland V (1970) Interaction of electro-ocular potentials with the contingent negative variation. Psychophysiology 7:103-111
- Waszak M, Obrist WD (1969) Relationship of slow potential changes to response speed and motivation in man. Electroencephalogr Clin Neurophysiol 27:113-120
- Watanabe S, Konishi M, Creutzfeldt O (1966) Postsynoptic potentials in the cat's visual cortex following electrical stimulation of afferent pathways. Exp Brain Res 1:272-283
- Weinberg H (1973) The contingent negative variation: its relation to feedback and expectant attention. Electroencephalogr Clin Neurophysiol 33:219-228
- Weinberg H, Papakostopoulos D (1975) The frontal CNV: its dissimularity to CNV.s recorded from other sites. Electroencephalogr Clin Neurophysiol 39:21-28
- Weinberg H, Walter WG, Crow HJ (1970) Intracerebral events in humans related to real imaginary stimuli. Electroencephalogr Clin Neurophysiol 29:1-9
- Weinberg H, Walter W, Cooper R, Aldridge V (1974) Emitted cerebral events. Electroencephalogr Clin Neurophysiol 36:449–456
- Wilke JT, Lansing RW (1973) Variations in the motor potential with force exerted during voluntary arm movements in man. Electroencephalogr Clin Neurophysiol 35:259-265
- Wilkinson RT, Haines E (1970) Evoked response correlates of expectancy during vigilance. Acta Psychol (Suppl) (Amst) 33:402-413
- Wilkinson RT, Spence MV (1973) Determinants of the post-stimulus reduction of contingent negative variation (CNV). Electroencephalogr Clin Neurophysiol 35:503-509
- Wood CC, Allison T, Goff WR, Williamson PD, Spencer DB (to be published) On the natural origin of P 300 in man. In: Kornhuber HH, Deecke L (eds) Motivation, motor and sensory processes of the brain. Prog Brain Res
- Wurtz RH, O'Flaherty JJ (1967) Physiological correlates of steady potential shifts during sleep and wakefulness. I. Sensitivity of steady potentials to alternations in carbon dioxide. Electroencephalogr Clin Neurophysiol 22:30–42

- Yingling CD, Skinner JE (1977) Gating of thalamic input to cerebral cortex by nucleus reticularis thalami. In: Desmeth JE (ed) Attention, voluntary contraction and event-related cerebral potentials. Karger, Basel, pp 70–96
- Zappoli R, Papini M, Briani S, Benvenuti P, Pasquinelli A (1976) CNV in patients with frontal-lobe lesions and mental disturbances. In: McCallum WC, Knott JR (eds) The responsive brain. Wright, Bristol, pp 158–163
- Zappoli R, Papini M, Denoth F, Pasquinelli A, Rossi L, Martinetti MG, Guerri S (1978a) CNV in patients treated by different psychosurgical procedures. In: Otto D (ed) Multidisciplinary perspectives in event-related brain potential research. EPA 600/9-77-043. Triangle Park Research Center, North Carolina, pp 376-382
- Zappoli R, Giuliano G, Rossi L, Pappini M, Ronchi O (1978b) CNV and SEP in shoeindustry workers with clinical EEG, EMG and CNV abnormalities due to toxic effects of adhesive solvents. In: Otto D (ed) Multidisciplinary perspectives in eventrelated brain potential research. EPA 600/9-77-043. Triangle Park Reearch Center, North Carolina, pp 481-484
- Zimmermann GN, Knott JR (1974) Slow potentials of the brain related to speech processing in normal speakers and stutterer. Electroencephalogr Clin Neurophysiol 37:599-607
- Zappoli R, Denoth F, Navona C (to be published) Changes of vertex-CNV, parietooccipital alpha activity and reaction time under closed- and open-eyed conditions.
   In: Kornhuber H, Deecke L (eds) Motivation, motor and sensory processes of the brain. Prog Brain Res

## $\alpha$ -Adrenoceptor Subclassification

#### KLAUS STARKE \*

#### Contents

1	Introduction.	199
2	Functional Studies: Pre- and Postsynaptic α-Adrenoceptors at Sympathetic Nerve-Muscle Junctions	201
3	Functional Studies: α <sub>2</sub> -Adrenoceptors Outside Noradrenergic Axons         3.1       Cholinergic Neurones         3.2       Noradrenergic Cell Bodies         3.3       Central Nervous System         3.4       Blood Platelets         3.5       Smooth Muscle	209 210 212 212 212 214 215
4	Radioligand-Binding Studies4.1 <sup>3</sup> H-Clonidine, <sup>3</sup> H-WB 4101 and <sup>3</sup> H-Prazosin.4.2 <sup>3</sup> H-Dihydroergokryptine.	216 216 221
5	Synthesis and Outlook5.1Agonist Potencies versus Affinities5.2Some Open Questions5.3Therapeutic Implications	221 221 225 227
Re	eferences	228

#### 1 Introduction

In this review the classification of  $\alpha$ -adrenoceptors into two major subgroups is discussed, an idea which derived primarily from studies on the pre- and postsynaptic  $\alpha$ -receptors at junctions between postganglionic sympathetic neurones and smooth muscle cells. The terms  $\alpha_1$  and  $\alpha_2$ , analogous to  $\beta_1$  and  $\beta_2$ , were loosely suggested (*Delbarre* and *Schmitt* 1973), subsequently used to refer to pre- and postsynaptic  $\alpha$ -receptors (*Langer* 1974), and finally generalized (*Berthelsen* and *Pettinger* 1977).

Different names are given to receptors when the sites at which they recognize and bind agonists and competitive antagonists differ structurally

<sup>\*</sup> Pharmakologisches Institut, Universität Freiburg, Hermann-Herder-Straße 5, D-7800 Freiburg i.B.

and when, hence, for a given drug the drug—receptor dissociation constants differ. We must then examine the methods used to determine the drug—receptor dissociation constant or, in other words, the *affinity* of a drug for the receptor, which is *proportional to the reciprocal of the dissociation constant*. One way is to measure the binding of radioactive drugs to receptors and the competitive inhibition of this binding by nonradioactive congeners. This yields dissociaton constants for the radioligand and for the competitor. Receptor properties may change, however, during the homogenization and incubation involved in such assays.

Affinities can also be determined in functional experiments in which the biological response of the tissue indicates the drug-receptor interaction. The interpretation is straightforward for antagonists. Receptor theory (see Ariens 1964; Furchgott 1964, 1972; Waud 1968) shows that the antagonist concentration which shifts the log concentration-response curve of an agonist to the right by the factor 2 is equal to the antagonistreceptor dissociation constant. The interpretation is less straightforward for agonists. In most studies only the  $EC_{50}$  is determined, i.e., the concentration that produces 50% of the maximal response obtainable with the agonist; only the potency of the agonists is thus evaluated, which is proportional to the reciprocals of the  $EC_{50}$  values. In general, the function relating response or potency to receptor occupation or affinity is not known. The  $EC_{50}$  of an agonist is not necessarily the concentration at which 50% of the receptors are occupied. Moreover, the relative potencies of a series of agonists at a receptor are not necessarily equal to the relative affinities for that receptor. The latter is only the case when the response is proportional to the 'stimulus' generated by the drug-receptor interaction, or when the agonists have equal efficacies (see Stephenson 1956; Furchgott 1972). Procedures have been worked out for functional determination of agonist-receptor dissociation constants, but have seldom been applied to  $\alpha$ -adrenoceptors (e.g., Besse and Furchgott 1976).

The present review is not the final presentation of a universally accepted theory, but a provisional account of a young and rapidly growing field of research. Contradictions are not only inevitable but welcome. In particular, the definition of  $\alpha_1$  - and  $\alpha_2$ -adrenoceptors given in Table 1 is open to correction and improvement. It incorporates most of the available data from the radioligand binding as well as from the functional studies that will be discussed at length. The drugs were chosen because they have been used in both kinds of studies. Agents that are more selective at  $\alpha_1$  - or  $\alpha_2$ receptors may eventually lead to a better definition. It can be seen that the relative *potencies* of agonists in general agree with their relative *affinities*. An exception that will be discussed in Chap. 5 is the high affinity but low potency of clonidine relative to phenylephrine at  $\alpha_1$ -adrenoceptors. Table 1. Proposed definition of  $\alpha_1$  - and  $\alpha_2$  -adrenoceptors

$\alpha_1$ -Adrenoceptor	
Agonist affinities and potencies	
Affinity and potency of ()-noradrenaline	$3-10$ times affinity and potency of $\alpha$ -methyl- noradrenaline <sup>a</sup>
Affinity and potency of (-)-noradrenaline	2-8 times affinity and potency of (-)-phenyl ephrine
Affinity of clonidine	4-15 times affinity of (-)-phenylephrine
Potency of clonidine	0.1-1 times potency of (-)-phenylephrine
Antagonist affinities	
$Prazosin \ge corynanthine, yohimb$	sine > rauwolscine
$\alpha_2$ -Adrenoceptor	
Agonist affinities and potencies	
-	0.1- 0.6 times affinity and potency of α-me- thylnoradrenaline <sup>a</sup>
Affinity and potency of (-)-noradrenaline	thylnoradrenaline a
Affinity and potency of	thylnoradrenaline a 15 – 150 times affinity and potency of
Affinity and potency of (-)-noradrenaline Affinity and potency of	thylnoradrenaline a
Affinity and potency of (-)-noradrenaline Affinity and potency of (-)-noradrenaline Affinity and potency of	thylnoradrenaline <sup>a</sup> 15 – 150 times affinity and potency of (-)-phenylephrine 50 –1000 times affinity and potency of

<sup>a</sup> The stereoisomers of  $\alpha$ -methylnoradrenaline are not always indicated unequivocally in the literature. It can be assumed that the racemate or the levorotatory form of the *ery thro*-isomer was used

Some aspects of the  $\alpha_1/\alpha_2$  subclassification have already been reviewed by *Berthelsen* and *Pettinger* (1977), *Starke* (1977), *Wood* et al. (1979) and, most extensively, *Wikberg* (1979b).

# 2 Functional Studies: Pre- and Postsynaptic α-Adrenoceptors at Sympathetic Nerve–Muscle Junctions

Presynaptic  $\alpha$ -adrenoceptors mediate the autoinhibition of transmitter release from noradrenaline neurones (see *Starke* 1977; *Westfall* 1977; *Vizi* 1979). That they differ from postsynaptic  $\alpha$ -adrenoceptors was first suggested by observations on rabbit hearts and cat spleens. In the rabbit heart, the relative potencies of phenylephrine, oxymetazoline, and naphazoline

Agonist	EC20 pre	EC50 post	EC20 pre	Antagonist	EC30 pre	K <sub>B post</sub>	EC30 pre
	(Mn)	(Wu)	EC20 post		(Wu)	$(M_{\rm m})$	KB post
(±)-Methoxamine	24 000	2 700	32.5	Prazosin	> 100	2	> 50
(-)-Phenylephrine	1 700	250	30.9	Corynanthine	10 000	250	40
(±)-Isoprenaline	12000	$10\ 000$	4	Clozapine	006	37	27
(-)-Noradrenaline	12	56	1.6	Azapetine	1 300	110	12
(±)-Metanephrine	$> 10\ 000$	26 000	> 1.4	Phentolamine	85	32	2.7
(-)-Adrenaline	1.9	13	0.58	Mianserin	550	250	2.2
Naphazoline	16	120	0.41	Piperoxan	600	870	0.69
Oxymetazoline	3.1	87	0.17	Dihydroergokristine	15	32	0.47
Clonidine	10	380	0.15	Tolazoline	700	3 900	0.18
$(-)$ -ery thro- $\alpha$ -				Dihydroergotamine	4	28	0.14
methylnoradrenaline	8.1	300	0.13	Yohimbol	115	1 000	0.12
Tramazoline	3.7	230	0.066	Yohimbine	×	400	0.020
				β-Yohimbine	15	760	0.020
				Rauwolscine	5	1 300	0.0039

sponse curves of agonists and the displacement of the curves to the right by antagonists were determined. For the investigation of presynaptic effects, the strips were preincubated with <sup>3</sup>H-noradrenaline (in the absence of propranolol, cocaine and corticosterone); the sympathetic fibres were stimulated transmurally at 2 Hz; the evoked overflow of tritium under these conditions reflects the EC30 pre, antagonist concentration that increased the evoked overflow of tritium by 30%. KB post, dissociation constant of the antagonist-postsynaptic receptor complex, calculated according to Arunlakshana and Schild (1959) or a similar method. From Starke neuronal and extraneuronal uptake of noradrenaline, respectively. For the investigation of postsynaptic effects concentration-rerelease of noradrenaline. EC20 pre, agonist concentration that reduced the evoked overflow of tritium by 20%. EC50 post and EC20 post, concentrations that produced 50% and 20%, respectively, of the maximal contractions obtainable with the agonist. et al. 1974, 1975b; Borowski et al. 1977; Endo et al. 1977; Weitzell et al. 1979; and unpublished results

Table 2. Comparison of effects of agonists and antagonists at pre- and postsynaptic α-adrenoceptors in the pulmonary artery of the

at the presynaptic, release-inhibiting receptors did not agree with their relative potencies at the postsynaptic, contractility-enhancing receptors (*Starke* 1972). In the cat spleen, phenoxybenzamine was more potent in blocking the postsynaptic than in blocking the presynaptic receptors (*Langer* 1973). In both studies it was explicitly concluded that the neuronal receptors differed from the classic postsynaptic receptors of the muscle cells.

Pre- and postsynaptic effects of agonists and antagonists in the pulmonary artery of the rabbit were compared in detail. Table 2 shows that the presynaptic receptors fulfil  $\alpha_2$  criteria and the postsynaptic receptors  $\alpha_1$  criteria. From the effective concentrations postsynaptic/presynaptic potency ratios were calculated (Table 2; note that a presynaptic/postsynaptic concentration ratio such as EC<sub>20 pre</sub>/EC<sub>20 post</sub> yields a postsynaptic/ presynaptic potency ratio). Given the same kind of receptors pre- and postsynaptically (and, for agonists, linear stimulus-response functions or the same efficacies), the ratios should be equal. In fact, however, they vary 500-fold for agonists and more than 10 000-fold for antagonists. The ratios also demonstrate that some drugs are quite selective. Clonidine,  $\alpha$ -methylnoradrenaline, and tramazoline preferentially activate the presynaptic  $\alpha_2$ -receptor, whereas phenylephrine and methoxamine preferentially activate the postsynaptic  $\alpha_1$ -receptor. After it had been classified as selectively presynaptic in the rabbit pulmonary artery (Starke et al. 1974), clonidine was widely used not only for the activation of  $\alpha_2$ -adrenoceptors in functional experiments but also for their labelling in radioligand-binding studies. Yohimbine was the first preferentially presynaptic antagonist to be recognized (Starke et al. 1975a). One of its diastereomers, namely rauwolscine, is even more selective. Prazosin (Cambridge et al. 1977; Cavero et al. 1977; Doxey et al. 1977), the yohimbine diastereomer corynanthine, clozapine, and azapetine preferentially block the postsynaptic  $\alpha_1$ -receptor. Phenoxybenzamine also primarily blocks postsynaptic  $\alpha$ receptors in the pulmonary artery but, being an irreversible antagonist, is not listed in Table 2.

The postsynaptic/presynaptic potency ratio is an essential feature of a drug, deciding how the drug affects the transmission of information through neuroeffector junctions (with postsynaptic receptors of the  $\alpha$  type).  $\alpha_1$ -Selective agonists such as phenylephrine act mainly postsynaptically, where their effect *is additive to* that of sympathetic nerve stimulation. In contrast,  $\alpha_2$ -selective agonists such as clonidine at low concentrations mainly diminish the release of noradrenaline and thereby *reduce* the response to sympathetic nerve stimulation. Furthermore,  $\alpha_1$ -selective antagonists like prazosin always *inhibit* the transmission of information. In contrast, low concentrations of  $\alpha_2$ -selective antagonists such as yohimbine primarily interrupt presynaptic autoinhibition, facilitate the release

Table 3. Comparison of effects of	ts of $\alpha$ -adrenergic agonists at pre- and postsynaptic $\alpha$ -adrenoceptors	aptic a-adrenoceptors	
Presynaptic effect <sup>a</sup>	Postsynaptic effect	Postsynaptic/presynaptic potency ratio	Reference
In vitro			
Rabbit heart, inhibition of noradrenaline overflow	Rabbit heart, increase in force of contraction	Phenylephrine > naphazoline, oxymetazoline	Starke (1972)
Rabbit pulmonary artery, inhibition of noradrenaline overflow	Rabbit pulmonary artery, contraction	Methoxamine, phenylephrine ≫ noradrenaline > naphazoline > oxymetazoline, clonidine, α-methyl- noradrenaline > tramazoline	<i>Starke</i> et al. (1975b) b
Rabbit ear artery, inhibition of contraction	Rabbit ear artery, contraction	Phenylephrine > clonidine > xylazine	Drew (1979) b
Rat vas deferens, inhibition of contraction	Rat vas deferens, contraction	Noradrenaline ≫ clonidine	<i>Doxey</i> et al. (1977) b
Rat vas deferens, inhibition of contraction	Rat vas deferens, contraction	Methoxamine, phenylephrine ≫ xylazine, naphazoline, clonidine, oxymetazoline	Drew (1977b)
Rat anococcygeus muscle, inhibition of noradrenaline overflow	Rat anococcygeus muscle, contraction	Methoxamine, phenylephrine, naphazoline > oxymetazoline ≫ clonidine	<i>Leighton</i> et al. (1979)
Mouse vas deferens, inhibition of contraction	Mouse vas deferens, contraction	Phenylephrine > noradrenaline > clonidine	<i>Marshall</i> et al. (1978)
In vivo			
Rabbit autoperfused hindlimb, inhibition of vasoconstriction	Rabbit autoperfused hindlimb, vasoconstriction	Phenylephrine > tramazoline	Steppeler et al. (1978)
Pithed rat, inhibition of tachycardia <sup>c</sup>	Pithed rat, increase in blood pressure	Phenylephrine, methoxamine, naphazoline > oxymetazoline > clonidine > xylazine	Drew (1976)

204

Pithed rat, inhibition of tachycardia c	Spinal rat, increase in blood pressure	Methoxamine > oxymetazoline > clonidine	Pichler and Kobinger (1978)
Pithed rat anococcygeus muscle, inhibition of contraction <sup>c</sup>	Pithed rat anococcygeus muscle, contraction	Naphazoline, oxymetazoline ≫ clonidine ≫ guanfacine	<i>Doxey</i> (1979) d
Cat, inhibition of reflex increase in blood pressure	Cat, increase in blood pressure	Phenylephrine > methoxamine > noradrenaline > oxymetazoline > tramazoline	Walland (1978)
Cat autoperfused hindlimb, inhibition of vasoconstriction	Cat autoperfused hindlimb, vasoconstriction	Methoxamine > oxymetazoline, clonidine	Pichler and Kobinger (1978)
Dog, inhibition of tachycardia	Dog, increase in blood pressure	Phenylephrine, noradrenaline $> \alpha$ -methylnoradrenaline	<i>Lokhandwala</i> et al. (1979) b

A comma between two drugs indicates that they had similar postsynaptic/presynaptic potency ratios or that their ratios were not differentiated.

- <sup>a</sup> The response inhibited by activation of presynaptic  $\alpha$ -receptors was evoked by electrical stimulation of the sympathetic pathway except in the experiments of *Walland* (1978) where a reflex increase in blood pressure was elicited by afferent stimulation.
  - b Noradrenaline uptake mechanisms and/or  $\beta$ -adrenoceptors blocked.
- c Preganglionic stimulation of the sympathetic pathway.
- d Similar results were obtained for the anococcygeus muscle in vitro.

#### $\alpha$ -Adrenoceptor Subclassification

of noradrenaline and thereby may *enhance* the transmission of information. This seemingly paradoxical effect of yohimbine was observed long ago (*Bacq* 1935).

Postsynaptic/presynaptic potency ratios found in other tissues are shown in Table 3. The rank orders agree well. In all studies phenylephrine has a high ratio and is rather selective for postsynaptic  $\alpha$ -receptors, and clonidine has a low ratio and is rather selective for presynaptic  $\alpha$ -receptors. Tramazoline, xylazine and guanfacine may surpass clonidine in selectivity (see Table 3; but see *Wikberg* 1978b), as may 2-(3,4-dihydroxyphenylimino)imidazolidine (*Hieble* and *Pendleton* 1979), 2-amino-6-allyl-5,6,7,8tetrahydro-4H-thiazolo-(5,4-d)-azepin (B-HT 920; *Kobinger* and *Pichler* 1980), and azepexole (B-HT 933; *Pichler* et al. 1980). Some dopamine derivatives also are preferentially presynaptic  $\alpha$ -adrenergic agonists in certain preparations (*Hicks* and *Cannon* 1979; *Steinsland* and *Hieble* 1979).

The findings with antagonists in cat spleen (Langer 1973; see above) and rabbit pulmonary artery (Table 2) have also been substantiated. Phenoxybenzamine preferentially blocks postsynaptic a-receptors in many tissues (Cubeddu et al. 1974; Dubocovich and Langer 1974; Drew 1976; Doxey et al. 1977), as does prazosin (Constantine et al. 1978; Lefèvre-Borg et al. 1978), whereas yohimbine preferentially blocks presynaptic  $\alpha$ -receptors (Doxey et al. 1977; Constantine et al. 1978; Kapur and Mottram 1978; Marshall et al. 1978; Walland 1978; Drew 1979; Leighton et al. 1979). Other mainly postsynaptic antagonists are thymoxamine (Drew 1976, 1977b; Marshall et al. 1978; Rhodes and Waterfall 1978), labetalol (Blakeley and Summers 1977), some butyrophenones (Göthert et al. 1977), indoramin (Algate and Waterfall 1978; Rhodes and Waterfall 1978), 2-[(2',6'-dimethoxyphenoxyethyl)-aminomethyl]1,4-benzodioxane (WB 4101: Butler and Jenkinson 1978; Kapur and Mottram 1978), and 2-[2-(4-o-methoxyphenyl-piperazine-1-yl)ethyl]-4,4-dimethyl-1,3(2H,4H)-isoquinolinedione (AR-C 239; Mouille et al. 1979, 1980). Slight presynaptic or slight postsynaptic or no preference has been reported for phentolamine (Cubeddu et al. 1974; Borowski et al. 1977; Doxey et al. 1977; Kapur and Mottram 1978; Rhodes and Waterfall 1978), piperoxan (Borowski et al. 1977; Blakeley and Summers 1978) and mianserin (Borowski et al. 1977; Doxey et al. 1978; Robson et al. 1978; Cavero et al. 1979b), indicating perhaps that these drugs occupy an intermediate position with no marked selectivity. In the rabbit pulmonary artery, tolazoline and dihydroergotamine have lower presynaptic than postsynaptic threshold concentrations, but have very flat presynaptic log concentration-response curves; high presynaptic affinity may be coupled with a presynaptic partial agonist effect that curtails the  $\alpha$ -adrenolytic facilitation of noradrenaline release (Borowski et al. 1977).

Before a general conclusion is drawn, some critical comments seem appropriate.

1) Only some of the studies reviewed here were carried out in vitro under optimal conditions for receptor characterization (*Furchgott* 1972), for instance in the presence of  $\beta$ -adrenolytic drugs and inhibitors of neuronal and extraneuronal noradrenaline uptake. Other studies were aimed at clarifying the consequences of selective receptor activation under near-normal conditions; these were of course performed in vivo and without auxiliary drugs. Some relevant details are specified in Table 3.

2) Presynaptic receptors modify the release of noradrenaline. In only a few studies was release measured as directly as possible, namely as 'overflow' into the medium (see Table 3). More frequently, the postsynaptic response was taken to reflect noradrenaline release. The postsynaptic response, however, may be modified by the postsynaptic as well as by the presynaptic action of a drug (see *Starke* 1977). For instance, a marked discrepancy between the effect on noradrenaline release and the effect on the response of the effector cells has been demonstrated for the  $\alpha$ -adrenergic imidazoline derivative cirazoline (*Dubocovich* et al. 1980).

3) Whenever drug effects on the depolarization-evoked release of noradrenaline are investigated, the transmitter itself is inevitably present in the biophase at rather high concentrations. This leads to underestimation of the presynaptic potency of  $\alpha$ -receptor agonists and antagonists (*Starke* et al. 1974), whether release is determined as overflow or by the postsynaptic response.

4) The term  $EC_x$  is normally used for the concentration that produces x% of the maximal response obtainable with the same drug; such  $EC_x$  values are suitable for calculation of relative potencies. The  $EC_{20}$  pre and  $EC_{30}$  pre values of Table 2 do not conform to this definition, since they do not refer to the individual maxima (see also, e.g., *Drew* 1976). Maximal presynaptic  $\alpha$ -receptor effects could not be determined for many drugs because at high concentrations they exerted marked side effects, e.g., displacement of stored noradrenaline. Of course these side effects also occur, although unnoticed, when postsynaptic responses are used to detect presynaptic effects, and are a possible source of error. It seems unlikely that the use, in Table 2, of presynaptic concentrations unrelated to the individual maxima greatly distorts the potency ratios.

5) The sympathetic pathway was sometimes stimulated preganglionically, and ganglionic actions of drugs cannot be excluded (see some papers in Table 3). For studies on the vas deferens one has to recall that the motor transmitter may not, or not only, be noradrenaline, in which case the presynaptic receptors may be located on non-noradrenergic neurones.

6) Potency ratios of agonists are the same as their affinity ratios only when their efficacies are equal (or in the rare event that the responses are proportional to the stimulus). The agonists in Tables 2 and 3, however, include some with high postsynaptic efficacy, e.g., noradrenaline, and some with low postsynaptic efficacy, e.g., clonidine (see Chap. 5). Clonidine and oxymetazoline may also have a lower presynaptic efficacy than noradrenaline (*Starke* et al. 1974; *Rand* et al. 1975; *Stjärne* 1975; *Medgett* et al. 1978). Part of the variability in potency ratios may thus reflect differences in agonist efficacies rather than affinities, although the size of the variability makes it unlikely that efficacy differences are the only reason. Moreover, the postsynaptic/presynaptic ratios of antagonists vary even further, and this can hardly be explained by anything but differences in affinity.

7) Although, on the whole, all studies agree, there are discrepancies in details. This is illustrated by the postsynaptic/presynaptic potency ratio of clonidine. Drew (1976) calculated the ratio of the dose which in pithed rats reduced the tachycardia evoked by sympathetic nerve stimulation by 50 beats/min, and the dose which increased blood pressure by 50 mmHg; the ratio was 1.4 and close to the ratio 0.5 that Pichler and Kobinger (1978) found in similar experiments. Starke et al. (1975b) calculated the ratio of the concentration that decreased the evoked overflow of noradrenaline from strips of the rabbit pulmonary artery by 20%, and the concentration that elicited 20% of the maximal contraction obtainable with clonidine; the ratio was 0.15. From experiments by Leighton et al. (1979) on the rat anococcygeus muscle in vitro two ratios can be calculated: one, namely 0.3, from the  $EC_{50}$  for inhibition of the evoked overflow of noradrenaline and the  $EC_{50}$  for the direct postsynaptic effect; the other, namely 0.03, from the EC<sub>50</sub> for inhibition of stimulation-evoked contractions and the postsynaptic EC<sub>50</sub>. Finally, Medgett et al. (1978) compared the EC<sub>50</sub> for inhibition of the evoked overflow of noradrenaline from guinea-pig atria with the EC<sub>50</sub> for contraction of rabbit aorta; the ratio was 0.0025. The ratios differ enormously. Nevertheless, all studies show that clonidine has a far lower postsynaptic/presynaptic potency ratio than other agonists such as naphazoline and phenylephrine. Much of the discrepancy is probably attributable to the arbitrary choice of potency parameters (see the two values of *Leighton* et al. 1979) and to the factors discussed under points 1-6. For instance, the presynaptic potency reported by *Medgett* et al. (1978) may be particularly high because these authors stimulated the atrial sympathetic nerves with only 5 pulses at 1 Hz. This vields low perineuronal concentrations of noradrenaline, little reduction in the apparent potency of clonidine (point 3), and a good approximation to the 'true' potency.

These comments question certain aspects of the work summarized here. On the whole, however, the evidence for differences between the pre- and the postsynaptic  $\alpha$ -adrenoceptors at sympathetic nerve-muscle junctions is overwhelming. Moreover, in all tissues examined so far, the presynaptic receptors appear to be similar to those of the rabbit pulmonary artery and, hence, at least predominantly  $\alpha_2$ , whereas the postsynaptic receptors appear to be at least predominantly  $\alpha_1$ . This conclusion does not imply that all noradrenergic axons possess only  $\alpha_2$ -receptors and all smooth and cardiac muscle cells only  $\alpha_1$ -receptors. There is now evidence for heterogeneity. Contraction-eliciting,  $\alpha_2$ -like adrenoceptors seem to occur on some smooth muscle cells such as those of rat blood vessels (Chap. 3). One reason, perhaps, for the high postsynaptic/presynaptic potency ratio of clonidine found by Drew (1976) and Pichler and Kobinger (1978) is that in rats clonidine causes vasoconstriction in part via smooth muscle  $\alpha_2$ -receptors. Presynaptic  $\alpha$ -receptors may be heterogeneous as well (*Doxey* and Everitt 1977; Constantine et al. 1978; Roach et al. 1978; Dubocovich 1979). For instance, the clonidine-induced inhibition of the tachycardia evoked by sympathetic nerve stimulation is antagonized by prazosin in dogs, cats and, according to some but not all investigators, rats (Constantine et al. 1978; Lefèvre-Borg et al. 1978; Roach et al. 1978; Cavero et al. 1979a; Mouillé et al. 1979; Timmermans et al. 1979b), suggesting that the neurones may possess both presynaptic  $\alpha_2$ - and prazosin-sensitive  $\alpha_1$ -receptors (Cavero et al. 1979a; see also Kobinger and Pichler 1980). On the other hand, there is evidence that prazosin at high doses blocks  $\alpha_2$ -receptors, and that at least in rats, the cardiac sympathetic nerves contain only the  $\alpha_2$  type (Docherty and McGrath 1980).

Presynaptic receptors of noradrenergic neurones other than those supplying muscle have rarely been investigated. In slices of rat brain cortex, yohimbine and rauwolscine increase the stimulation-evoked release of noradrenaline at much lower concentrations than does corynanthine (*Starke* and *Stamm*, unpublished results), and prazosin has no effect (*Dubocovich* 1979). Thus, the receptors satisfy  $\alpha_2$ -criteria. On the other hand, in brain slices phentolamine is more potent than yohimbine, and piperoxan is almost equipotent with yohimbine (*Taube* et al. 1977; *Delini-Stula* et al. 1979), whereas in the rabbit pulmonary artery yohimbine is much more potent than its congeners (Table 2), casting some doubt on the similarity between the two kinds of presynaptic receptors.

#### 3 Functional Studies: $\alpha_2$ -Adrenoceptors Outside Noradrenergic Axons

Observations on pre- and postsynaptic  $\alpha$ -receptors at noradrenergic synapses gained a new dimension when receptors resembling the presynaptic ones were found on structures other than noradrenergic terminal axons.

The terms  $\alpha_1$ - and  $\alpha_2$ -receptors had been proposed as synonyms for postand presynaptic  $\alpha$ -receptors (*Langer* 1974). The existence of the  $\alpha_2$  type elsewhere made it necessary to use these prefixes solely for receptors with different affinities for drugs regardless of location or function (*Berthelsen* and *Pettinger* 1977; *Cedarbaum* and *Aghajanian* 1977; *Drew* 1977a, 1978; *Wikberg* 1978a, b; *Starke* and *Langer* 1979). The following examples are just a selection, to illustrate the wide range of potential  $\alpha_2$ -adrenoceptor locations.

#### 3.1 Cholinergic Neurones

 $\alpha$ -Adrenergic agonists inhibit the release of acetylcholine from central, preganglionic autonomic, and postganglionic parasympathetic neurones; they facilitate the release of acetylcholine from motor neurones to skeletal muscle (see Vizi 1979). The poperties of the  $\alpha$ -receptors in the intestine have been the subject of most frequent study.  $\alpha$ -Receptor agonists can inhibit intestinal cholinergic activity at three sites at least, namely preganglionic terminals, the cell bodies of the cholinergic neurones of the enteric plexuses, and the axon terminals of these neurones. Much of the inhibition is probably exerted at the latter two sites (Paton and Vizi 1969; Kosterlitz et al. 1970; Drew 1978). An early, unexplained observation was that methoxamine failed to inhibit acetylcholine release (Paton and Vizi 1969), whereas naphazoline, xylazine, and clonidine were potent inhibitors (Oberdorf and Kroneberg 1970; Deck et al. 1971). Retrospectively, this is easily understood from the character of the receptors, which differ from classic smooth muscle  $\alpha$ -adrenoceptors (Wikberg et al. 1975), and were eventually shown to be of the  $\alpha_2$  type (Drew 1977a, 1978; Wikberg 1978a.b).

Table 4 summarizes potencies of agonists, relative to noradrenaline, in inhibition of acetylcholine outflow or cholinergic transmission in the guinea-pig ileum. The receptors fulfil  $\alpha_2$  criteria. In the terminal ileum phenylephrine fails to inhibit cholinergic transmission (Table 4, column c), but competitively antagonizes the effect of noradrenaline. Its affinity can, therefore, be estimated; it is low, the dissociation constant being 63  $\mu M$  (Wikberg 1978a).

Similar experiments with agonists suggest that  $\alpha_2$ -receptors occur on cholinergic neurones of the rabbit jejunum (*Wikberg* 1979a), the chicken stomach (*Seno* et al. 1978), the guinea-pig gallbladder (*Lee* and *Fujiwara* 1977), the cat submaxillary gland (*Green* et al. 1979), and perhaps the cerebral cortex of the rat (*Vizi* 1979).

The  $\alpha_2$  character is confirmed by experiments with blocking drugs. Yohimbine is a potent antagonist at the  $\alpha$ -receptors of the cholinergic neu-

	Potencies	Potencies relative to noradrenaline				
	а	b	c	d	e	
Clonidine	9.1	10	20	10	18	
(–)-ery thro-	3.2					
$\alpha$ -methylnoradrenaline	e					
Tramazoline	2.9					
Naphazoline	2.2					
Oxymetazoline	1.7	2.2	3			
(-)-Noradrenaline	1	(1)	1	1	1	
(-)-Phenylephrine	0.0089	0.0004	0	0.001	0.018	
(±)-Methoxamine	0.0004	0.0002				

Table 4. Relative potencies of agonists at  $\alpha$ -adrenoceptors of the cholinergic neurones of the guinea-pig ileum

The effect measured was the inhibition of contractions of the ileum evoked by electrical stimulation (columns a-d) or the inhibition of the stimulation-evoked outflow of acetylcholine (column e). a) From Wikberg (1978b). Ilea from reserpine-pretreated guinea pigs were mounted in medium containing sotalol, cocaine and corticosterone. With the author's consent, attention is drawn to the fact that this paper contains some printing errors that confuse the meaning, especially in Fig. 1 where the drugs are coded. For instance,  $\alpha$ -methylnoradrenaline is 41, not 4k. b) From Drew (1978). Ilea were mounted in medium containing propranolol. Since the paper contains no data for noradrenaline, the potency of clonidine was taken to be 10. Effect of phenylephrine and methoxamine not reversed by piperoxan. c) From Wikberg (1978a). Terminal ilea were incubated in medium containing sotalol. d) From Ennis et al. (1979). Enantiomers not indicated. e) From Vizi (1979). Stereoisomers not indicated.

rones of the guinea-pig ileum (*Drew* 1978; *Andréjak* et al. 1980) and the cat submaxillary gland (*Green* et al. 1979). Tolazoline and piperoxan are also potent antagonists in the ileum, whereas for phentolamine divergent results have been reported (*Paton* and *Vizi* 1969; *Drew* 1978; *Wikberg* 1978a). The  $\alpha_1$ -selective antagonists labetalol (*Drew* 1978), prazosin and AR-C 239 (*Andréjak* et al. 1980) fail to block the receptors. Interestingly, the facilitatory  $\alpha$ -receptors at skeletal muscle motor nerve endings are more akin to  $\alpha_1$ - than to  $\alpha_2$ -receptors (*Malta* et al. 1979).

Some of the critical comments of Chap. 2 also pertain here. For instance, not only acetylcholine but also noradrenaline is released by field stimulation of the guinea-pig ileum. Noradrenaline then inhibits the release of acetylcholine as shown by the fact that yohimbine and tolazoline increase the evoked release of acetylcholine (*Kilbinger* and *Wessler* 1979). The presence of noradrenaline may lead to underestimation of the potency of exogenous  $\alpha$ -receptor agonists and antagonists as discussed earlier. Moreover, drugs may affect the release of acetylcholine indirectly by changing primarily the release of noradreanline.

Wikberg (1978b) avoided this by depletion of the noradrenaline stores by reserpine. Moreover, in his experiments  $\beta$ -adrenoceptors as well as the neuronal and extraneuronal uptake of noradrenaline were blocked. Wikberg (1979b) calculated the correlation of the relative potencies of agonists at inhibiting cholinergic transmission in the guinea-pig ileum (Table 4, column a) and at inhibiting noradrenaline release in the rabbit pulmonary artery (Table 2). The very close correlation (r = 0.96) is good evidence for the similarity of these receptors.

### 3.2 Noradrenergic Cell Bodies

 $\alpha$ -Receptor agonists inhibit the firing of the noradrenergic cells of the locus ceruleus and hyperpolarize sympathetic ganglion cells. The receptors involved appear to be  $\alpha_2$  (*Cedarbaum* and *Aghajanian* 1977; *Brown* and *Caulfield* 1979). For instance, the effect of agonists on sympathetic ganglia was reduced by yohimbine but not by up to 10  $\mu$ M prazosin (*Brown* and *Caulfield* 1979). Phenylephrine was only slightly less potent in hyperpolarizing the ganglia than was noradrenaline (*Brown* and *Caulfield* 1979), and this might be taken as evidence against a pure  $\alpha_2$ -receptor population. The experimental conditions of the authors did not, however, exclude release of noradrenaline by phenylephrine and, hence, an indirect mode of action. From these observations and those discussed in Chap. 2 the general picture emerges that central and peripheral noradrenaline neurones may be endowed with inhibitory  $\alpha_2$ -adrenoceptors, in other words inhibitory  $\alpha_2$ -autoreceptors, in both their soma-dendritic and their terminal region.

### 3.3 Central Nervous System

Interest in central  $\alpha$ -adrenoceptors rose steeply when it became clear that these receptors were important sites of action of clonidine and related drugs as well as of  $\alpha$ -methylnoradrenaline, the active metabolite of  $\alpha$ -methyldopa (*Delbarre* and *Schmitt* 1969; *Andén* et al. 1970; *Heise* and *Kroneberg* 1970, 1973; *Hoyer* and *van Zwieten* 1971; *Kobinger* and *Walland* 1971; *Schmitt* et al. 1971). As has been mentioned, electrophysiological and brain slice experiments indicate that cerebral noradrenergic neurones possess soma-dendritic and presynaptic  $\alpha_2$ -adrenoceptors. These studies concerned relatively isolated structures and simple responses. The involvement of  $\alpha$ -receptor subtypes in more complex functions of the central nervous system is, of course, very difficult to elucidate. The blood-brain barrier impedes drug access, and drugs may act simultaneously at many sites. All conclusions are tentative, and only selected aspects will be discussed here (see also *Berthelsen* and *Pettinger* 1977; *Schmitt* 1977; *Starke* 1977). Clonidine and related compounds reduce the synthesis and utilization of noradrenaline in rat brain and spinal cord (Andén et al. 1970; Scholtysik et al. 1975; Fuller et al. 1977). Low doses of clonidine are required. Its effect is strongly counteracted by yohimbine, piperoxan and tolazoline, but only weakly by phenoxybenzamine and not at all by prazosin. In the case of another  $\alpha$ -adrenergic effect upon the central nervous system, namely the increase in strength of the flexor reflex of acutely spinalized rats, the pattern of drug effects is quite different. High doses of clonidine are needed, and its effect is strongly counteracted by phenoxybenzamine and prazosin, but only weakly by yohimbine, piperoxan and tolazoline (Andén et al. 1976, 1978). Since both the biochemical and the motor effect occur in the spinal cord, the contrasting patterns are probably not due to differing access of the drugs to the receptors. Rather, the receptors differ. Those mediating the inhibition of noradrenaline neurones appear to be  $\alpha_2$ , whereas those mediating the increase of the reflex appear to be  $\alpha_1$ .

The reflex-enhancing receptors are probably located on non-catecholamine neurones (Andén et al. 1976). The turnover-decreasing receptors might be identical with the  $\alpha_2$ -autoreceptors of noradrenergic neurones. However, this is by no means certain. The turnover-decreasing receptors may well be located on non-noradrenergic neurones and may depress the noradrenergic neurones via a neurone chain (Andén et al. 1976). The question of identity with the  $\alpha_2$ -autoreceptors always arises when a central drug effect is supposed to be  $\alpha_2$ -adrenergic. It is stressed that the  $\alpha_2$  properties of a central receptor do not imply that it is a soma-dendritic or presynaptic receptor of noradrenergic neurones or that the effect it mediates involves central noradrenergic pathways.

The central receptors responsible for the cardiovascular depression caused by clonidine-like drugs and  $\alpha$ -methyldopa appear to differ from smooth muscle  $\alpha$ -receptors (see *Schmitt* 1977; *Kobinger* 1978) and may be of the  $\alpha_2$  type (*Berthelsen* and *Pettinger* 1977; *Starke* 1977). The recently reported antagonism by prazosin does not exclude this possibility, since high doses of prazosin were given (*Timmermans* et al. 1979b). Whatever the subtype, most but not all of the receptors are probably located on non-catecholamine neurones because clonidine and related compounds retain much of their central autonomic effect after near-complete catecholamine depletion (*Haeusler* 1974; *Kobinger* and *Pichler* 1974; *Reynoldson* et al. 1979; *Pichler* et al. 1980).

Sedation is a prominent side effect of clonidine-like drugs and of  $\alpha$ methyldopa and is probably mediated by central  $\alpha$ -adrenoceptors. The following findings suggest that these receptors may be  $\alpha_2$ .

1) In chicks with incomplete blood-brain barrier the potency of agonists in inducing sleep declines in the order clonidine  $\gg \alpha$ -methylnoradrenaline, naphazoline > noradrenaline (*Fügner* and *Hoefke* 1971). 2) Sleep caused in chicks by clonidine is antagonized by phentolamine, tolazoline, piperoxan, and yohimbine, but not by azapetine and phenoxybenzamine (*Delbarre* and *Schmitt* 1969, 1971, 1973; *Fügner* 1971). Similar results were obtained in mice where the agonists prolong the chloral hydrate sleeping time (*Delbarre* and *Schmitt* 1971, 1973).

3) In rats the sedative potencies of intracerebro-ventricularly injected agonists decline in the order clonidie  $\gg$  xylazine, naphazoline > methoxamine; phenylephrine is inactive. The sedation induced by clonidine is antagonized by intraventricular injection of phentolamine, piperoxan, yohimbine and tolazoline, but not by injection of labetalol, prazosin and thymoxamine (*Drew* et al. 1979). Essentially similar results were obtained in rats by other authors (*Clineschmidt* et al. 1979, 1980; *Delini-Stula* et al. 1979; *Nomura* et al. 1980).

The question again arises as to whether the sedation-mediating sites, if  $\alpha_2$ , are autoreceptors. Since sedation and inhibition of noradrenaline turnover are produced by similarly low doses, this does seem possible. In further support of this view, clonidine no longer causes sedation after catecholamine depletion, but, on the contrary, causes locomotor stimulation (Zebrowska-Lupina et al. 1977; see also Strömbom and Svensson 1980).

Effects of clonidine related to sedation include suppression of conditioned avoidance responses, of self-stimulation, and of the fear-potentiated startle reflex. These effects also seem to be mediated by  $\alpha_2$ -receptors and, moreover, by inhibition of central noradrenaline neurones (*Franklin* and *Herberg* 1977; *Hunt* et al. 1978; *Robson* et al. 1978; *Davis* et al. 1979; *Hawkins* and *Monti* 1979).

Like noradrenaline neurones, central *adrenaline* pathways can be modulated via  $\alpha$ -receptors belonging to the  $\alpha_2$  group (*Scatton* et al. 1979).

#### 3.4 Blood Platelets

Adrenaline, noradrenaline, and  $\alpha$ -methylnoradrenaline induce an  $\alpha$ -adrenergic aggregation of human platelets. Non-catecholamine agonists do not share this effect with the three catecholamines but may enhance the aggregation produced by other stimuli (see *Grant* and *Scrutton* 1979; *Hsu* et al. 1979). The catecholamines also inhibit platelet adenylate cyclase. Non-catecholamine agonists fail to do so or are only partial agonists (*Jakobs* et al. 1978; *Tsai* and *Lefkowitz* 1978). All  $\alpha_2$ -receptors discussed hitherto were probably constituents of the cell membrane of neurones. The platelet  $\alpha$ -receptors may be an example of non-neuronal  $\alpha_2$ -receptors.

 $\alpha$ -Methylnoradrenaline is twice and clonidine 15 times as potent as noradrenaline in enhancing ADP-induced aggregation, whereas phenyl-ephrine and methoxamine have little effect (*Hsu* et al. 1979). Yohimbine

and phentolamine, but not up to 100  $\mu M$  prazosin, counteract adrenalineinduced aggregation (Glusa et al. 1979; Hsu et al. 1979; Lasch and Jakobs 1979). Non-catecholamines that have little or no effect of their own do antagonize adrenaline-induced aggregation as well as the inhibition of adenylate cyclase (Hsu et al. 1979; Lasch and Jakobs 1979; Rossi et al. 1979); oxymetazoline, naphazoline, and clonidine are much more potent in this respect than is phenylephrine (Lasch and Jakobs 1979). These findings are compatible with the view that platelets possess an  $\alpha_2$ -receptor through which only adrenaline, noradrenaline, and  $\alpha$ -methylnoradrenaline act with full efficacy (Glusa et al. 1979; Hsu et al. 1979; Lasch and Jakobs 1979). They may, however, in addition contain an  $\alpha_1$ -receptor which does not contribute much to the response to adrenaline but which may mediate the aggregation-enhancing effect of phenylephrine (Grant and Scrutton 1979). The observation that  $\alpha$ -methylnoradrenaline is only half as potent as noradrenaline in eliciting aggregation and inhibiting adenylate cyclase would be unusual for a pure  $\alpha_2$  population.

#### 3.5 Smooth Muscle

Perhaps the most unforeseen  $\alpha_2$ -receptors, or  $\alpha_2$ -like receptors, have recently been detected in smooth muscle cells, the  $\alpha$ -receptors of which had served as  $\alpha_1$  prototypes. The initial observation was that prazosin antagonized the contractile effect of noradrenaline strongly in human visceral arteries, but at best only slightly in digital arteries. There seemed to be a prazosin-sensitive and a prazosin-resistant group of contraction-mediating vascular  $\alpha$ -adrenoceptors (Moulds and Jauernig 1977; Jauernig et al. 1978). Following up the idea that the two groups might be  $\alpha_1$ - and  $\alpha_2$ -receptors, Drew and Whiting (1979) showed that in cats and rats prazosin strongly counteracted the hypertensive effect of phenylephrine, but much less markedly counteracted the effect of noradrenaline. Conversely, yohimbine was slightly more potent against noradrenaline than against phenylephrine. The effect of noradrenaline was also relatively prazosin-resistant in the isolated cat hindlimb and mesenteric vascular beds but not in the renal vascular bed. Drew and Whiting (1979) concluded that phenylephrine constricts blood vessels solely via the prazosin-sensitive  $\alpha_1$ -receptor, whereas noradrenaline, except in cat kidney, possesses an additional, prazosin-insensitive site of action. Although they hesitated to classify the latter site as  $\alpha_2$ , other experiments support this possibility. For instance, in pithed rats prazosin, at a dose which abolishes the pressor effect of high doses of phenylephrine, only reduces the pressor effect of clonidine and does not change that of the highly  $\alpha_2$ -selective agonist xylazine; yohimbine antagonizes the effect of all three agonists, but antagonizes xylazine more than
phenylephrine (*Docherty* et al. 1979; *Docherty* and *McGrath* 1980). There are, by now, numerous analogous observations, the most clear-cut and consistent findings being that contractions of smooth muscle caused by agonists with a strong  $\alpha_2$ -component are at least partly resistant to  $\alpha_1$ -selective antagonists, and that the resistant contractions can be blocked by  $\alpha_2$ -selective antagonists and, hence, are probably mediated by  $\alpha_2$ -adrenoceptors (*Timmermans* et al. 1979a; *Constantine* et al. 1980; *Drew* 1980; *Flavahan* and *McGrath* 1980; *Kobinger* and *Pichler* 1980; *Madjar* et al. 1980; *Timmermans* and *van Zwieten* 1980).

A cautionary remark must be added. None of the studies summarized above was carried out under optimal conditions for receptor characterization (see *Furchgott* 1972). Only in one published abstract has differentiation of smooth muscle  $\alpha_1$ - and  $\alpha_2$ -receptors under strictly controlled in vitro conditions been reported;  $\alpha_2$ -receptors were found in dog veins but not arteries (*De Mey* and *Vanhoutte* 1980). Further such in vitro experiments are necessary before the occurrence of smooth muscle  $\alpha_2$ -receptors can be considered as established.

#### 4 Radioligand-Binding Studies

 $\alpha$ -Adrenoceptors have been labelled with several radioactive ligands. In the present context, <sup>3</sup> H-dihydroergokryptine, <sup>3</sup> H-clonidine, <sup>3</sup> H-WB 4101, and <sup>3</sup> H-prazosin are especially important.

### 4.1 <sup>3</sup> H-Clonidine, <sup>3</sup> H-WB 4101 and <sup>3</sup> H-Prazosin

In the studies summarized in Tables 5 and 6, membrane fractions of tissue homogenates were incubated with low concentrations of the radioligands in 25–30 mM Tris-HCl buffer pH 7.4–8.0. Nonspecific binding was measured in the presence of high concentrations of unlabelled ligands and was subtracted from total binding. The remaining specific binding was saturable and of high affinity, as shown by the dissociation constants  $K_D$  of the radioligand-receptor complexes in the Table legends. The Tables contain dissociation constants  $K_i$  of nonradioactive drugs that competed with the radioligands for specific binding.

The rank orders of affinities of agonists and antagonists for the <sup>3</sup> Hclonidine-binding site agree well in the various tissues and studies (Table 5). The rank orders of affinities for the <sup>3</sup> H-WB 4101 site resemble the rank orders for the <sup>3</sup> H-prazosin site and are therefore presented together

Table 5. $K_i$ values $(nM)$ for the inhibition of the binding of <sup>3</sup> H-clonidine to $\alpha$ -adrenoceptors	for the inhibiti	on of the bindi	ng of <sup>3</sup> H-clon	idine to α-a	drenoceptor	S		
	Rat brain minus cereballium	Rat brain minus cerebelium	Rat cerebral	Rat cerebral	Calf cerebral	Rabbit duodenum	Guinea-pig ileum	Guinea-pig kidney
	a	b	C	d	e e	يىغ	ත	Ч
Agonists		A NAME AND A DAMAGE AND A	A second s					
(-)-Phenylephrine	270			390	260		2300	4200
()-Noradrenaline	17	10.2		25.2	16	27	58	170
$\alpha$ -Methylnoradrenaline	7.7	1.5		8.7			18	
Clonidine	5.7	0.9		4.8	1.2	8.5	5.7	27
Naphazoline	5.7			5.4	0.9			30
Tramazoline	4.2			10.4			6.9	
Oxymetazoline	1.9			4.6	1.6			780
Antagonists								
Phentolamine	22	5.2	2.1	78	1.2	300	12	24
Piperoxan	95	325						430
Rauwolscine			41				57	
Yohimbine	150	253	47	360			82	220
I olazoline WP A 101	180	025	110	7560			730	
Prazosin	007	6960 6960	5000	3430			5400	1400
Corvnanthine			6200				5800	
Indoramin	6500		6500					
Tissues were homogenized and incubated with $0.4-20 \text{ nM}^3$ H-clonidine. KD for $^3$ H-clonidine was $2-11 \text{ nM}$ . <i>a) UPrichard</i> et al. (1977). <i>b) Miach</i> et al. (1980). <i>c)</i> Phentolamine, WB 4101, prazosin and indoramin from <i>UPrichard</i> et al. (1978); rauwolscine, yo-himbine and corynanthine from <i>Tanaka</i> and <i>Starke</i> (1980). <i>d) Hornung</i> et al. (1979). <i>e) UPrichard</i> et al. (1977). <i>f) UPrichard</i> and <i>Snyder</i> (1977). <i>f</i> and <i>f</i> an	zed and incub: (1980). c) Phe ine from Tanak anaka and Star	ated with $0.4-$ ntolamine, WB ia and Starke (1 ke (1979). h) Ji	-20 nM <sup>3</sup> H-cl 4101, prazo 980). <i>d</i> ) <i>Horr</i> <i>arrott</i> et al. (	lonidine. KD sin and indo <i>nung</i> et al. (1 1979); pheny	) for <sup>3</sup> H-clo tramin from (979). e) U'I	nidine was 2- U'Prichard et Prichard and S antiomer not	-11 nM. a) U'I t al. (1978); rau Snyder (1977). specified.	'richard et al. Iwolscine, yo- f) U'Prichard

 $\alpha$ -Adrenoceptor Subclassification

Table 6. $K_i$ values $(nM)$ for the inhibition of the binding of <sup>3</sup> H-WB 4101 and <sup>3</sup> H-prazosin to $\alpha$ -adrenoceptors	nM) for the	inhibition o	of the bindi	ng of <sup>3</sup> H-W	/B 4101 and <sup>3</sup> 1	H-prazosin to $\alpha$	-adrenoceptors		
	Rat brain a	Rat cerebral cortex b	Rat cerebral cortex c	Calf cerebral cortex d	Rat vas deferens e	Rat brain minus cerebellum f	Rat brain minus cerebellum	Rat cerebral cortex h	Guinea-pig lung
Agonists						4	o	1	
(-)-Phenylephrine ()-Noradrenaline	2 600 1 000		4 200	1 300	470	1 400 900	5 615	16 000 10 200	110 000 1 400
a-meunyi- noradrenaline Clonidine Naphazoline Tramazoline Oxymetazoline	2 800 430 110 24		1 162 226	450	850	340 43 290 23	36 474 2 315	56 000 950 320 1 000 630	2 200
Antagonists									
Phentolamine Piperoxan	3.6 180	3.6	74.8	1.2	5.3 150	360	304 5 775	30	6.6 770
Kauwoiscine Yohimbine	480	419 419			480	1 000	8 906	800	3 100
WB 4101 Prazosin	0.6	0.3 0.5	2.7	0.3		2 000 1.0 0.1	3.8 0.3	8 0.6	0.8 0.1
Corynanthine Indoramin	5.9	170 5.9				5.0	1	2	4.0
Tissues were homogenized and incubated with 0.2–8 nM <sup>3</sup> H-WB 4101 (a–e) or 0.1–2 nM <sup>3</sup> H-prazosin (f–i). KD values were 0.3–2.7 nM for <sup>3</sup> H-WB 4101 and 0.2–0.5 nM for <sup>3</sup> H-prazosin. a) U'Prichard et al. (1977). b) Phentolamine, WB 4101, prazosin and indoramin from U'Prichard et al. (1978). d) U'Prichard et al. (1978). d) U'Prichard et al. (1979). f) Greengrass and Starke (1980). c) Davis et al. (1978). d) U'Prichard and Snyder (1977). e) U'Prichard and Snyder (1979). f) Greengrass and Bremner (1979); phenylephrine enantiomer not specified. g) Miach et al. (1980). h) Hornung et al. (1979). i) Barnes et al. (1979). f) Greengrass and Bremner (1979); phenylephrine enantiomer not specified. g) Miach et al. (1980). h) Hornung et al. (1979). i) Barnes et al. (1979).	genized and b 4101 and Prichard et rd and Sny 1 g) Miach e	incubated 0.2-0.5 nM al. (1978); der (1977). t al. (1980)	with 0.2–8 M for <sup>3</sup> H-p rauwolscine e) U'Prichu . h) Hornun	$nM^{3}H-W$ razosin. a) e, yohimbi ard and Sn ng et al. (1)	B 4101 (a-e) U'Prichard et ne and coryna yder (1979). f) 979). i) Barnes	d and incubated with 0.2–8 nM <sup>3</sup> H-WB 4101 (a–e) or 0.1–2 nM <sup>3</sup> H-prazosin (f–i). KD values were 0.3– and 0.2–0.5 nM for <sup>3</sup> H-prazosin. a) U'Prichard et al. (1977). b) Phentolamine, WB 4101, prazosin and rd et al. (1978); rauwolscine, yohimbine and corynanthine from Tanaka and Starke (1980). c) Davis et al. d Snyder (1977). e) U'Prichard and Snyder (1979). f) Greengrass and Bremner (1979); phenylephrine enan- iach et al. (1980). h) Hornung et al. (1979). i) Barnes et al. (1979)	'H-prazosin (f- Phentolamine, <i>maka</i> and <i>Starl</i> I <i>Bremner</i> (197	-i). KD valu WB 4101, <i>ke</i> (1980). d (9); phenyle	es were 0.3- prazosin and <i>J Davis</i> et al. sphrine enan-

218

K. Starke

in Table 6, but they differ from the rank orders for the <sup>3</sup>H-clonidine site. There are, however, many discrepancies between absolute  $K_i$  values. For instance, the  $K_i$  values of agonists for inhibition of <sup>3</sup>H-prazosin binding to rat brain membranes found by *Hornung* et al. (1979; Table 6, column h) exceed the corresponding values for inhibition of <sup>3</sup>H-prazosin binding found by *Greengrass* and *Bremner* (1979; column f) and for inhibition of <sup>3</sup>H-WB 4101 binding found by *U'Prichard* et al. (1977; column a) by factors of between 2 and 27. The  $K_i$  values of agonists for inhibition of <sup>3</sup>H-clonidine binding reported by *Jarrott* et al. (1979; Table 5, column h) also are higher than corresponding values (in columns a–g); the discrepancy for oxymetazoline is particularly large, as pointed out by the authors. Surprisingly different constants have been published for phentolamine. Its  $K_i$  for <sup>3</sup>H-clonidine binding varied between 1.2 and 300 nM in one laboratory (Table 5, columns e and f) and between 2.1 and 22 nM even in one tissue, namely rat brain (columns a and c).

The  $K_i$  values reveal two characteristic properties of the binding sites. Firstly, agonists tend to have much higher affinities for the <sup>3</sup> H-clonidine site than for the <sup>3</sup> H-WB 4101 and <sup>3</sup> H-prazosin sites, whereas the reverse holds true for antagonists. This was noted even in the first experiments by *Greenberg* et al. (1976), *U'Prichard* and *Snyder* (1977), and *U'Prichard* et al. (1977). These authors concluded that either there was a single  $\alpha$ -adrenoceptor that could take agonist- and antagonist-preferring conformations, or, more probably, that there were two distinct, non-interconvertible  $\alpha$ adrenoceptors, one with high affinity for agonists (the 'agonist receptor site'), the other with high affinity for antagonists (the 'antagonist receptor site', see also *Greenberg* and *Snyder* 1978; *Greenberg* et al. 1978; *Peroutka* et al. 1978).

With growing functional evidence for two major  $\alpha$ -receptor subtypes, the second property of the binding sites became obvious, namely that the <sup>3</sup>H-clonidine site satisfies  $\alpha_2$ -, whereas the <sup>3</sup>H-WB 4101 site and the <sup>3</sup>Hprazosin site satisfy  $\alpha_1$ -receptor criteria (*U'Prichard* et al. 1978; *Greengrass* and *Bremner* 1979; *Hornung* et al. 1979; *U'Prichard* and *Snyder* 1979). Comparison of Tables 1, 5, and 6 makes this plain.

How are the two properties related? Is the <sup>3</sup>H-clonidine site an  $\alpha_2$ -receptor, and is this receptor simultaneously an agonist-selective receptor? Is the <sup>3</sup>H-WB 4101 and <sup>3</sup>H-prazosin site (if both drugs label the same site) an  $\alpha_1$ -receptor, and is this receptor simultaneously an antagonist-preferring receptor?

The predilection of antagonists for the <sup>3</sup>H-WB 4101 and <sup>3</sup>H-prazosin site is not without exception. In the experiments of *U'Prichard* et al. (1977) tolazoline, piperoxan, and yohimbine had higher affinity to the <sup>3</sup>H-clonidine site (the presumed agonist site) than to the <sup>3</sup>H-WB 4101 site (the presumed antagonist site). Similar results were obtained by *Hornung* et al. (1979) and *Miach* et al. (1980; see Tables 5 and 6). *U'Prichard* et al.

(1977) explained this by a partial agonist character of the three antagonists. Although the explanation may hold good for tolazoline, there is no evidence for a partial agonist effect of yohimbine and piperoxan at  $\alpha$ -adrenoceptors. Even more striking are recent observations with the yohimbine diastereomer rauwolscine. Rauwolscine had 40–54 times higher affinity for the <sup>3</sup>H-clonidine site in guinea-pig ileum and rat cerebral cortex (Table 5, columns c and g) than for the cerebrocortical <sup>3</sup>H-WB 4101 site (Table 6, column b). Since rauwolscine, yohimbine, tolazoline, and (not consistently) piperoxan preferentially block  $\alpha_2$ -adrenoceptors, all these findings fit in with the <sup>3</sup>H-clonidine site being an  $\alpha_2$ -receptor but not an agonist receptor.

There is as yet no exception to the predilection of agonists for the <sup>3</sup>Hclonidine site. Yet, Tables 5 and 6 show that the K<sub>i</sub> values of phenylephrine for <sup>3</sup>H-clonidine binding (260–4 200 nM) and for <sup>3</sup>H-WB 4101 and <sup>3</sup>H-prazosin binding (1 400–110 000 nM) overlap, indicating that the preference of this drug for the presumed agonist site is small. On the whole, the available data suggest that the <sup>3</sup>H-clonidine site is identical with the  $\alpha_2$ -receptor found in functional studies, whereas the <sup>3</sup>H-WB 4101 and <sup>3</sup>H-prazosin sites are identical with the  $\alpha_1$ -receptor. There is a marked tendency for agonists to bind mainly to the <sup>3</sup>H-clonidine site and for antagonists to bind mainly to the <sup>3</sup>H-WB 4101 and <sup>3</sup>H-prazosin sites *under the conditions of the binding assays*. This, however, is not generally valid, and the sites should not be called the agonist and antagonist site, respectively.

Given this conclusion, a major difficulty remains. In the binding studies all agonists except phenylephrine have much higher affinity for the  $\alpha_2$ than for the  $\alpha_1$ -receptor. From the data of *UPrichard* et al. (1977) the following  $\alpha_1/\alpha_2$  affinity ratios can be calculated (columns a of Tables 5 and 6): phenylephrine 0.10; noradrenaline 0.017;  $\alpha$ -methylnoradrenaline 0.0028; clonidine 0.013; naphazoline 0.052; tramazoline 0.038; oxymetazoline 0.079). Functionally, however, the agonists are not so much more potent at  $\alpha_2$ - than at  $\alpha_1$ -receptors; some are equipotent (like noradrenaline) and some actually more potent at  $\alpha_1$ -receptors (like phenylephrine). For instance, comparison of the affinity ratios with the postsynaptic/presynaptic  $(\alpha_1/\alpha_2)$  potency ratios of Table 2 shows that the latter are about 10 times higher than the affinity ratios for the imidazolines, and about 100 times higher than the affinity ratios for the phenylethylamines. Comparison of  $\alpha_1/\alpha_2$  affinity ratios with the  $\alpha_1/\alpha_2$  potency ratios of agonists in guinea-pig aorta ( $\alpha_1$ ) and in cholinergic neurones ( $\alpha_2$ ; Wikberg 1978b) yields essentially similar values. As will be discussed in detail in Chap. 5, two factors may account for this discrepancy. Firstly, in physiological media the affinity of agonists to the  $\alpha_2$ -receptor may be lower than in the Tris buffer media used in the binding assays; secondly, in intact tissues a moderate affinity to  $\alpha_1$ -receptors may be translated into a high potency.

### 4.2 <sup>3</sup>H-Dihydroergokryptine

In several tissues the number of <sup>3</sup>H-dihydroergokryptine-binding sites is equal to the sum of the numbers of <sup>3</sup>H-clonidine and <sup>3</sup>H-WB 4101 binding sites. <sup>3</sup>H-Dihydroergokryptine binds to the <sup>3</sup>H-clonidine site and to the <sup>3</sup>H-WB 4101 site with similar affinity. These and other observations were first interpreted in terms of agonist and antagonist  $\alpha$ -receptors and a partial agonist character of dihydroergokryptine (Greenberg and Snyder 1978; Peroutka et al. 1978). It now seems more likely that <sup>3</sup>H-dihydroergokryptine binds with about equal affinity to  $\alpha_1$ - and  $\alpha_2$ -adrenoceptors (Miach et al. 1978; U'Prichard et al. 1978; Hoffman et al. 1979; U'Prichard and Snyder 1979; Wood et al. 1979). The binding of <sup>3</sup>H-dihydroergokryptine to  $\alpha_1$ - and  $\alpha_2$ -receptors in tissues in which both occur is inhibited by other ligands in a typical manner. If the unlabelled ligand has similar affinity to either receptor, the log concentration-inhibition curve is monophasic and steep, as one would expect from the competition of two drugs for one site. If, however, the ligand is selective for one receptor, the curve becomes shallow or overtly biphasic. In experiments on brain membranes, steep monophasic curves are obtained with phentolamine (Miach et al. 1978), but biphasic curves with clonidine, WB 4101 and indoramin (Peroutka et al. 1978) as well as with yohimbine and prazosin (Miach et al. 1978). The differential effects of phentolamine, prazosin and yohimbine on the binding of <sup>3</sup>H-dihydroergokryptine have been used to study quantitatively  $\alpha$ receptor subtypes in various tissues. Only  $\alpha_1$ -adrenoceptors were found in rat liver, only  $\alpha_2$ -adrenoceptors in human platelets, and a mixture in rabbit uterus. Surprisingly, the affinity of a given antagonist for one receptor type varied greatly. For instance, the K<sub>i</sub> of yohimbine at the  $\alpha_2$ -receptor of rabbit uterus was 14 nM, but at the  $\alpha_2$ -receptor of human platelets 0.8 nM; its K<sub>i</sub> at the  $\alpha_1$ -receptor of rabbit uterus was 3000 nM, but at the  $\alpha_1$ receptor of rat liver 64 nM (Hoffman et al. 1979). Species or tissue differences may be responsible, as the authors propose. Nevertheless, such large differences are disquieting when one looks for generally valid properties of  $\alpha_1$ -adrenoceptors and  $\alpha_2$ -adrenoceptors.

### 5 Synthesis and Outlook

### 5.1 Agonist Potencies Versus Affinities

Functional studies on sympathetic nerve-muscle junctions, functional studies in other tissues, and radioligand binding studies have converged to the hypothesis that  $\alpha$ -adrenoceptors fall into two subclasses. A major

discrepancy between the functional and the binding experiments was pointed out earlier. For all agonists,  $\alpha_1/\alpha_2$  potency ratios greatly exceed  $\alpha_1/\alpha_2$  affinity ratios. The difference is about 10-fold for imidazoline agonists and about 100-fold for phenylethylamine agonists. It is suggested that two factors may explain the discrepancy: firstly, the use of nonphysiological media in binding experiments, and secondly, a particular kind of stimulus-response relationship in the case of smooth muscle  $\alpha_1$ -adrenoceptors.

In the first place, all incubations for radioligand binding that yielded the data shown in Tables 5 and 6 were carried out in Tris buffer, sometimes supplemented with ascorbic acid, EDTA, or magnesium, but without the monovalent cations that abound in physiological media. Sodium and, less effectively, potassium ions, however, reduce the binding of <sup>3</sup>H-clonidine to brain membranes; the binding of <sup>3</sup>H-WB 4101 and <sup>3</sup>H-prazosin, on the other hand, is not changed (Greenberg et al. 1978; Glossmann and Presek 1979; Hornung et al. 1979; Rouot et al. 1980). In platelets, sodium ions diminish the affinity of agonists but hardly affect that of antagonists to the <sup>3</sup>H-dihydroergokryptine sites which, in these cells, are mainly  $\alpha_2$ receptors (Tsai and Lefkowitz 1978). These studies suggest that sodium reduces the affinity of agonists to  $\alpha_2$ - but not  $\alpha_1$ -adrenoceptors. Sodium may even increase the affinity of clonidine for  $\alpha_1$ -adrenoceptors (Glossmann and Hornung 1980). If so, the use of sodium-free media will lead to a systematic overestimation of agonist affinities for the  $\alpha_2$ -receptor and a systematic understimation of their  $\alpha_1/\alpha_2$  affinity ratios (in comparison with more physiological conditions).

Secondly, in several blood vessels the contractile effect of  $\alpha$ -adrenergic agonists seems to be an upwardly convex hyperbolic function of the stimulus generated by the agonist-receptor interaction (or of the fraction of receptors occupied, which is proportional to the stimulus; see Fig. 3 of Johansson et al. 1972; Fig. 3 of Besse and Furchgott 1976; and Fig. 2 of Ruffolo et al. 1979). Classic receptor theory states that, in such a case, an agonist with high efficacy should produce a large proportion of its maximal effect when only a small fraction of the receptors is occupied; in other words, its EC<sub>50</sub> should be much lower than its dissociation constant, and the potency of the agonist should exceed its affinity. Conversely, an agonist with low efficacy even at full receptor occupation should produce only stimuli in the lower, near-linear part of the stimulus-response hyperbola; its EC<sub>50</sub> should be similar to its dissociation constant, and potency similar to affinity. Indeed, noradrenaline, phenylephrine and  $\alpha$ -methylnoradrenaline, which all have high and equal efficacies in rabbit aorta, produce 50% of their maximal effects when only 6% of the receptors are occupied (Besse and Furchgott 1976). Conversely, clonidine and naphazoline, which in rat aorta possess only 2% of the efficacy of phenylephrine, have to occupy approximately 50% of the receptors in order to produce 50% of their maximal effect (*Ruffolo* et al. 1979). Thus, in the smooth muscle of blood vessels with classic  $\alpha_1$ -adrenoceptors the modest affinity of phenylethylamines is translated into high potency because of the upwardly convex hyperbolic stimulus-response curve. This mechanism does not work for low-efficacy drugs such as many imidazolines.

A brief recapitulation of receptor theory may illustrate this point further (see *Ariens* 1964; *Furchgott* 1964, 1972; *Waud* 1968). The equilibrium between the agonist A, the receptor R and the agonist—receptor complex AR is assumed to obey the law of mass action, from which the familiar equation

$$\frac{[RA]}{[R_t]} = \frac{[A]}{[A] + K_A}$$
(1)

is derived, where [RA], [R<sub>t</sub>] and [A] are the concentrations of agonist-receptor complex, total receptors, and free agonist, respectively, and K<sub>A</sub> is the dissociation constant. *Stephenson* (1956) suggested that formation of the agonist-receptor complex generates a stimulus S which is proportional to the fraction of receptors occupied:

$$S = e \frac{[RA]}{[R_t]}$$

where e is the efficacy of the agonist. The stimulus, in turn, triggers the response of the tissue, so that the effect is some function (not necessarily linear) of S:

$$\frac{E}{E_{max}} = f(S) = f\left(\frac{[RA]}{[R_t]}\right)$$

where E is the response at a certain concentration of a given agonist, and  $E_{max}$  the maximal response obtainable with an agonist of maximal efficacy. Let us now consider the case, exemplified by the effect of  $\alpha$ -adrenergic agonists on vascular smooth muscle, in which  $E/E_{max}$  is a rectangular hyperbolic function of S according to

$$\frac{E}{E_{max}} = \frac{S}{S+1}$$

In Fig. 1,  $E/E_{max}$  is plotted against S, which is indicated on the abscissa in arbitrary units. Below the abscissa S is shown, for an agonist with high efficacy (e = 100) and for an agonist with low efficacy (e = 1), which fractional receptor occupation generates a certain stimulus. When the high efficacy drug occupies all receptors ([RA]/[Rt] = 1), it produces the stimulus S = 100 and, hence, the effect  $E/E_{max} = 0.99$  of the theoretical maximal effect obtainable with an agonist of maximal efficacy. The upper dashed horizontal line is drawn at the response  $E/E_{max} = 0.99 / 2 = 0.495$ , and the corresponding dashed vertical line indicates that this effect is triggered by the stimulus S = 0.98 and a fractional receptor occupation of only 0.0098. Thus, at the EC<sub>50</sub> of the agonist with e = 100, only about 1% of the receptors are occupied, and from equation (1) it follows that  $EC_{50} = 0.01 \text{ K}_A$ . This is the type of relationship found when noradrenaline or phenylephrine acts on  $\alpha_1$ -adrenoceptors. On the other hand, when the low-efficacy drug occupies all receptors, it produces the stimulus S = 1 and, hence the effect  $E/E_{max} = 0.5$ . The lower dashed horizontal line is drawn at the response  $E/E_{max} = 0.5 / 2 = 0.25$ , and the corresponding vertical line indicates that this effect is triggered by the stimulus S = 0.33 and a fractional receptor occupation of 0.33. Thus, at the EC<sub>50</sub> of the agonist with e = 1, about 33% of the receptors are occupied, and from equation (1) it follows that EC<sub>50</sub> = 0.5 K<sub>A</sub>. This is the type of relationship found when clonidine or naphazoline acts on  $\alpha_1$ -adrenoceptors.

One can also express the difference between high- and low-efficacy agonists using the concept of spare receptors. A calculation analogous to that described above shows that the e = 100 agonist produces 95% of its own maximal effect when only 16% of the receptors are occupied; in other words, there are unoccupied spare receptors even when the agonist produces essentially its maximal effect, and essentially the whole concentration—response curve is covered when receptor occupation is increased from 0 to only about 20%; hence the high potency in comparison with affinity. Conversely, the e = 1 agonist produces 95% of its own maximal effect when 90% of the receptors are occupied; there is no appreciable receptor reserve, and in order to cover the whole concentration—response curve, receptor occupation has to be increased from 0 to about 100%; hence the low potency in comparison with affinity.



Fig. 1. Relationship between receptor occupation  $[RA]/[R_t]$ , stimulus S and response  $E/E_{max}$  for an agonist with high efficacy, e = 100, and an agonist with low efficacy, e = 1

In the definition proposed in Table 1 it was necessary to distinguish between the high affinity and the low potency of clonidine, relative to phenylephrine, at  $\alpha_1$ -adrenoceptors. We can now explain this by the low efficacy of clonidine coupled with a hyperbolic stimulus—response curve at the  $\alpha_1$ -receptors so far studied. Interestingly, published data show no analogous difference between the potency and the affinity of clonidine, relative to phenylephrine, at  $\alpha_2$ -adrenoceptors (see Table 1). Agreement between relative potencies and relative affinities at  $\alpha_2$ -receptors is also found when other imidazolines and phenylethylamines are compared. Since the agonists do have different efficacies in eliciting  $\alpha_2$ -adrenergic effects (see Chap. 2 and Wikberg 1978b), the agreement suggests that for the  $\alpha_2$ -adrenergic mechanisms studied so far the stimulus—response function is approximately linear. In conclusion, factor 1 (the monovalent cation effect) holds good for all agonists. Factor 2 (the hyperbolic stimulus-response function) introduces a difference between the effects of high and the effects of low efficacy agonists at  $\alpha_1$ - but not at  $\alpha_2$ -adrenoceptors. Taken together, the factors may explain why the  $\alpha_1/\alpha_2$  potency ratio is about 10 times higher than the  $\alpha_1/\alpha_2$  affinity ratio for the imidazolines, and about 100 times higher for the phenylethylamines, for the  $\alpha$ -adrenergic mechanisms studied so far.  $\alpha_1/\alpha_2$  Affinity ratios of phenylephrine, noradrenaline and  $\alpha$ -methylnoradrenaline are 0.10, 0.017 and 0.0028, respectively (*U'Prichard* et al. 1977). The 100-fold increase results in phenylephrine becoming  $\alpha_1$ -selective, noradrenaline being equipotent, and  $\alpha$ -methylnoradrenaline remaining  $\alpha_2$ -selective in functional experiments.

Wikberg (1978b) proposed the following definition of  $\alpha$ -receptor subtypes: "When the relative affinities of phenylephrine and noradrenaline are greater than those of tramazoline, xylazine and clonidine,  $\alpha_1$  receptors are present, and when these orders are reversed, the receptors are of the  $\alpha_2$  type". Ruffolo et al. (1979) criticized this definition since "ED50 values . . . may be poor estimates of agonist affinity". They pointed out that in rat aorta clonidine had 14 times higher affinity than phenylephrine for the smooth muscle  $\alpha$ -receptor, so that, according to *Wikberg* (1978b), "the postsynaptic *alpha* receptors of the rat aorta would have to be termed presynaptic which is clearly incorrect". The somewhat misleading terminology in these statements (presynaptic/postsynaptic; affinity/potency) should not detract from the fact that the scientific messages of these authors can be reconciled when affinity and potency are distinguished as in Table 1 and when possible reasons for the apparent discrepancy are recognized. Wikberg (1978b) determined relative potencies only, and the potency of phenylephrine does exceed, or is about as high as, that of clonidine at all  $\alpha_1$ -receptors studied including those of rat aorta (Ruffolo et al. 1979).

# 5.2 Some Open Questions

Questions, not least those concerning discrepant findings, have been raised throughout this review. Some of a more general nature will be added here.

Receptor classes, once established, tend to divide, and it is stressed that the  $\alpha_1/\alpha_2$  subclasses may not be homogeneous. Some divergent results may be due to the existence of sub-subclasses. A given receptor may also change its behaviour when its environment changes. Therefore, descriptions and distinctions should refer to a defined, preferably the physiological, environment. Although differences between receptors were defined here as differences in their recognition sites, receptors may differ elsewhere in their molecules as well. We know too little, however, about components outside the recognition site to make them the basis for classification.

There may be many more  $\alpha_2$ -receptors than those discussed in Chaps. 2 and 3, such as in frog skin melanocytes (*Berthelsen* and *Pettinger* 1977), fat cells (*Berthelsen* and *Pettinger* 1977; *Aktories* et al. 1980), and the kidney, where they may inhibit renin release (*Berthelsen* and *Pettinger* 1977; see however, *Morris* et al. 1979). These various possibilities deserve further study.

The occurrence of  $\alpha_2$ -like receptors in smooth muscle makes reconsideration of previous investigations necessary.  $\alpha_2$ -Receptors should be sought in tissues which have been used for the definition of  $\alpha_1$ -receptors, such as the rabbit pulmonary artery. It has been reported that high concentrations of oxymetazoline desensitize the smooth muscle of rat vas deferens to oxymetazoline and other imidazoline agonists but not to noradrenaline, methoxamine, and phenylephrine (*Ruffolo* et al. 1977). Is this because the imidazolines, in contrast to the phenylethylamines, act mainly via smooth muscle  $\alpha_2$ -receptors [in spite of the fact that under different experimental conditions Docherty et al. (1979) did not find postsynaptic  $\alpha_2$ -receptors in this organ]? In some tissues the response to sympathetic nerve stimulation is reduced only slightly or not at all by clonidine (e.g. in the cat hindlimb: Haeusler 1976). Postsynaptic  $\alpha_2$ -receptors might account for such findings. It may be that some reports on tissue or species differences in smooth muscle  $\alpha$ -receptors (see Harper et al. 1979) which have not been discussed here can also be partly explained in the light of the existence of smooth muscle  $\alpha_2$ -receptors. Generally speaking, the possibility that both  $\alpha_1$ - and  $\alpha_2$ -receptors mediate smooth muscle contraction, and in particular vasoconstriction, opens up a wide field for research (e.g., Constantine and Lebel 1980; Langer et al. 1980).

Studies of radioligand binding to  $\alpha$ -adrenoceptors have led to many discrepancies, such as the wide variation in K<sub>i</sub> values of antagonists for inhibiton of <sup>3</sup>H-dihydroergokryptine binding to what one would assume to be the same receptor. It is important to discover the reason. The binding of ligands to  $\alpha$ -receptors is influenced not only by monovalent but also by divalent cations and nucleotides (*U'Prichard* and *Snyder* 1978; *Glossmann* and *Presek* 1979; *Tsai* and *Lefkowitz* 1979; *Rouot* et al. 1980), and this may be relevant for the physiological implications of binding studies. The binding of selective radioligands has been used to visualize  $\alpha_1$ - and  $\alpha_2$ -receptors in rat brain autoradiographically (*Young* and *Kuhar* 1979). This method may help to localize the receptors not only in terms of brain nuclei but also in terms of cell regions, such as the pre- and postsynaptic sides of synapses.

<sup>3</sup>H-Dihydroergokryptine is assumed to bind with about equal affinity to  $\alpha_1$ - and  $\alpha_2$ -adrenoceptors. Surprisingly, the effects of dihydroergokryptine on the subtypes have not been compared functionally.

A final question that can only be mentioned briefly is whether part of the binding of radioligands is to presynaptic or soma-dendritic  $\alpha$ -receptors of noradrenergic neurones. So far, such autoreceptor binding has been demonstrated only in rat heart (*Story* et al. 1979), and has been sought in vain in several other tissues (e.g., *U'Prichard* et al. 1977; *Tanaka* and *Starke* 1979; for possible reasons, see *Taube* et al. 1977).

## 5.3 Therapeutic Implications

The existence of two  $\alpha$ -adrenoceptor subclasses has therapeutic significance. As mentioned in Chap. 3.3 clonidine-like drugs and  $\alpha$ -methylnoradrenaline probably owe their antihypertensive effect to their  $\alpha_2$ -adrenergic properties. The same holds good for some side effects such as sedation and dry mouth (inhibition of cholinergic transmission). Imidazolines used as nasal decongestants may cause sedation; this may also be an  $\alpha_2$ adrenergic effect. Clonidine suppresses symptoms of opiate withdrawal (Gold et al. 1978) possibly by acting at  $\alpha_2$ -autoreceptors of the noradrenaline neurones of the locus ceruleus, which are also endowed with opiate receptors (Montel et al. 1974). If depression involves diminished transmission through central noradrenergic synapses,  $\alpha_2$ -selective antagonists might have a salutary effect by facilitating transmission. This has in fact been reported (Puech et al. 1979). On the other hand, the view that mianserin acts as an antidepressant by blockade of central  $\alpha_2$ -autoreceptors is questionable since the drug is not selective (see Chap. 2).

Selectivity for  $\alpha_1$ -adrenoceptors may also be important. Sympathomimetic drugs used as vasoconstrictors should have a marked  $\alpha_1$  component. From this point of view, phenylephrine and methoxamine would have an advantage over the imidazolines.  $\alpha_1$ -Selective antagonists such as prazosin appear to be useful, peripherally acting antihypertensive drugs. At low doses they should not interrupt the  $\alpha$ -adrenergic feedback inhibition of noradrenaline release and hence should cause less tachycardia and hyperreninaemia than nonspecific  $\alpha$ -adrenolytic drugs like phentolamine. Other factors, however, must contribute to the lack of reflex tachycardia after hypotensive doses of prazosin (*Hardey* and *Lokhandwala* 1979).  $\alpha_1$ -Selective antagonists also cause less gastrointestinal stimulation than does phentolamine, possibly because they do not interfere with the noradrenergic inhibition of intestinal cholinergic neurones.

#### References

- Aktories K, Schultz G, Jakobs KH (1980) Regulation of adenylate cyclase activity in hamster adipocytes. Inhibition by prostaglandins, α-adrenergic agonists and nicotinic acid. Naunyn Schmiedebergs Arch Pharmacol 312:167-173
- Algate DR, Waterfall JF (1978) Action of indoramin on pre- and postsynaptic α-adrenoceptors in pithed rats. J Pharm Pharmacol 30:651-652
- Andén NE, Corrodi H, Fuxe K, Hökfelt B, Hökfelt T, Rydin C, Svensson T (1970) Evidence for a central noradrenaline receptor stimulation by clonidine. Life Sci 9/I: 513-523
- Andén NE, Grabowska M, Strömbom U (1976) Different alpha-adrenoreceptors in the central nervous system mediating biochemical and functional effects of clonidine and receptor blocking agents. Naunyn Schmiedebergs Arch Pharmacol 292:43-52
- Andén NE, Gomes C, Persson B, Trolin G (1978) R 28935 and prazosin: effects on central and peripheral alpha-adrenoreceptor activity and on blood pressure. Naunyn Schmiedebergs Arch Pharmacol 302:299–306
- Andréjak M, Pommier Y, Mouillé P, Schmitt H (in press) Effects of some α-adrenoceptor agonists and antagonists on the guinea-pig ileum. Naunyn Schmiedebergs Arch Pharmacol
- Ariens EJ (ed) (1964) Molecular pharmacology, vol 1. Academic Press, New York
- Arunlakshana O, Schild HO (1959) Some quantitative uses of drug antagonists. Br J Pharmacol 14:48-58
- Bacq ZM (1935) Recherches sur la physiologie et la pharmacologie du système nerveux autonome. Arch Int Pharmacodyn Thér 52:471–492
- Barnes P, Karliner J, Hamilton Č, Dollery C (1979) Demonstration of  $\alpha_1$ -adrenoceptors in guinea pig lung using <sup>3</sup>H-prazosin. Life Sci 25:1207–1214
- Berthelsen S, Pettinger WA (1977) A functional basis for classification of α-adrenergic receptors. Life Sci 21:595-606
- Besse JC, Furchgott RF (1976) Dissociation constants and relative efficacies of agonists acting on *alpha* adrenergic receptors in rabbit aorta. J Pharmacol Exp Ther 197: 66-78
- Blakeley AGH, Summers RJ (1977) The effects of labetalol (AH 5158) on adrenergic transmission in the cat spleen. Br J Pharmacol 59:643-650
- Blakeley AGH, Summers RJ (1978) The effects of piperoxan on uptake of noradrenaline and overflow of transmitter in the isolated blood perfused spleen of the cat. Br J Pharmacol 63:683-687
- Borowski E, Starke K, Ehrl H, Endo T (1977) A comparison of pre- and postsynaptic effects of  $\alpha$ -adrenolytic drugs in the pulmonary artery of the rabbit. Neuroscience 2:285-296
- Brown DA, Caulfield MP (1979) Hyperpolarizing 'α<sub>2</sub>'-adrenoceptors in rat sympathetic ganglia. Br J Pharmacol 65:435-445
- Butler M, Jenkinson DH (1978) Blockade by WB 4101 of α-adrenoceptors in the rat vas deferens and guinea-pig taenia caeci. Eur J Pharmacol 52:303-311
- Cambridge D, Davey MJ, Massingham R (1977) Prazosin, a selective antagonist of postsynaptic α-adrenoceptors. Br J Pharmacol 59:514P-515P
- Cavero I, Lefèvre F, Roach A (1977) Further studies on cardiovascular effects of prazosin (P). Fed Proc 36:955
- Cavero I, Dennis T, Lefèvre-Borg F, Perrot P, Roach AG, Scatton B (1979a) Effects of clonidine, prazosin and phentolamine on heart rate and coronary sinus catecholamine concentration during cardioaccelerator nerve stimulation in spinal dogs. Br J Pharmacol 67:283-292
- Cavero I, Gomeni R, Lefèvre-Borg F, Roach AG (1979b) Effects of mianserin on noradrenaline uptake, cardiac presynaptic and vascular postsynaptic α-adrenoceptors in rats. Br J Pharmacol 66:132P-133P

- Cedarbaum JM, Aghajanian GK (1977) Catecholamine receptors on locus coeruleus neurons: pharmacological characterization. Eur J Pharmacol 44:375–385
- Clineschmidt BV, Flataker LM, Faison E, Holmes R (1979) An in vivo model for investigating  $\alpha_1$  and  $\alpha_2$ -receptors in the CNS: studies with mianserin. Arch Int Pharmacodyn Thér 242:59-76
- Clineschmidt BV, Flataker LM, Faison EP, Haubrich DR (1980) α-Methyldopa reduces locomotor activity in rats via its metabolite, α-methylnorepinephrine, acting on α<sub>2</sub>adrenoceptors. Arch Int Pharmacodyn Thér 244:231--243
- Constantine JW, Lebel W (in press) Irreversible blockade by phenoxybenzamine of  $\alpha_1$ -but not of  $\alpha_2$ -vascular receptors in dogs and the effects of propranolol. Naunyn Schmiedebergs Arch Pharmacol
- Constantine JW, Weeks RA, McShane WK (1978) Prazosin and presynaptic  $\alpha$ -receptors in the cardioaccelerator nerve of the dog. Eur J Pharmacol 50:51–60
- Constantine JW, Gunnell D, Weeks RA (1980)  $\alpha_1$  and  $\alpha_2$ -vascular adrenoceptors in the dog. Eur J Pharmacol 66:281–286
- Cubeddu L, Barnes EM, Langer SZ, Weiner N (1974) Release of norepinephrine and dopamine-β-hydroxylase by nerve stimulation. I. Role of neuronal and extraneuronal uptake and of *alpha* presynaptic receptors. J Pharmacol Exp Ther 190:431-450
- Davis JN, Arnett CD, Hoyler E, Stalvey LP, Daly JW, Skolnick P (1978) Brain α-adrenergic receptors: comparison of (<sup>3</sup>H)WB 4101 binding with norepinephrine-stimulated cyclic AMP accumulation in rat cerebral cortex. Brain Res 159:125–135
- Davis M, Redmond DE, Baraban JM (1979) Noradrenergic agonists and antagonists: effects on conditioned fear as measured by the potentiated startle paradigm. Psychopharmacology (Berlin) 65:111-118
- Deck R, Oberdorf A, Kroneberg G (1971) Die Wirkung von 2-(2,6-Dichlorphenylamino)-2-imidazolin-Hydrochlorid (Clonidin) auf die Kontraktion und die Acetylcholin-Freisetzung am isolierten, koaxial elektrisch gereizten Meerschweinchenileum. Arzneim Forsch 21:1580-1584
- Delbarre B, Schmitt H (1969) Influences pharmacologiques exercées sur la durée du sommeil provoqué chez le poussin par la 2-(2,6-dichlorophénylamino)-2-imidazoline (St 155, Catapresan). CR Soc Biol (Paris) 163:1922-1926
- Delbarre B, Schmitt H (1971) Sedative effects of α-sympathomimetic drugs and their antagonism by adrenergic and cholinergic blocking drugs. Eur J Pharmacol 13: 356-363
- Delbarre B, Schmitt H (1973) A further attempt to characterize sedative receptors activated by clonidine in chickens and mice. Eur J Pharmacol 22:355-359
- Delini-Stula A, Baumann P, Büch O (1979) Depression of exploratory activity by clonidine in rats as a model for the detection of relative pre- and postsynaptic central noradrenergic receptor selectivity of  $\alpha$ -adrenolytic drugs. Naunyn Schmiedebergs Arch Pharmacol 307:115–122
- De Mey JG, Vanhoutte PM (1980) Differences in pharmacological properties of postjunctional alpha-adrenergic receptors among arteries and veins. Arch Int Pharmacodyn Thér 244:328-329
- Docherty JR, McGrath JC (1980) A comparison of pre- and post-junctional potencies of several alpha-adrenoceptor agonists in the cardiovascular system and anococcygeus of the rat. Evidence for two types of post-junctional alpha-adrenoceptor. Naunyn Schmiedebergs Arch Pharmacol 312:107-116
- Docherty JR, MacDonald A, McGrath JC (1979) Further sub-classification of  $\alpha$ -adrenoceptors in the cardiovascular system, vas deferens and anococcygeus of the rat. Br J Pharmacol 67:421P-422P
- Doxey JC (1979) Pre- and postsynaptic effects of  $\alpha$ -agonists in the anococcygeus muscle of the pithed rat. Eur J Pharmacol 54:185–189
- Doxey JC, Everitt J (1977) Inhibitory effects of clonidine on responses to sympathetic nerve stimulation in the pithed rat. Br J Pharmacol 61:559-566

- Doxey JC, Everitt J, Metcalf G (1978) Mianserin an analysis of its peripheral autonomic actions. Eur J Pharmacol 51:1–10
- Doxey JC, Smith CFC, Walker JM (1977) Selectivity of blocking agents for pre- and postsynaptic  $\alpha$ -adrenoceptors. Br J Pharmacol 60:91–96
- Drew GM (1976) Effects of  $\alpha$ -adrenoceptor agonists and antagonists on pre- and postsynaptically located  $\alpha$ -adrenoceptors. Eur J Pharmacol 36:313–320
- Drew GM (1977a) Pharmacological characterization of presynaptic  $\alpha$ -adrenoceptors which regulate cholinergic activity in the guinea-pig ileum. Br J Pharmacol 59:513P
- Drew GM (1977b) Pharmacological characterisation of the presynaptic  $\alpha$ -adrenoceptors in the rat vas deferens. Eur J Pharmacol 42:123–130
- Drew GM (1978) Pharmacological characterization of the presynaptic α-adrenoceptors regulating cholinergic activity in the guinea-pig ileum. Br J Pharmacol 64:293–300
- Drew GM (1979) Presynaptic α-adrenoceptors: their pharmacological characterisation and functional significance. In: Langer SZ, Starke K, Dubocovich ML (eds) Presynaptic receptors. Pergamon Press, Oxford, pp 59–65
- Drew GM (1980) Postsynaptic  $\alpha_2$ -adrenoceptors mediate pressor responses to 2-N,N-dimethylamino-5,6-dihydroxy-1,2,3,4-tetrahydronaphthalene (M-7). Eur J Pharma-col 65:85–87
- Drew GM, Whiting SB (1979) Evidence for two distinct types of postsynaptic αadrenceptor in vascular smooth muscle in vivo. Br J Pharmacol 67:207-215
- Drew GM, Gower AJ, Marriott AS (1979) α<sub>2</sub>-Adrenoceptors mediate clonidine-induced sedation in the rat. Br J Pharmacol 67:133-141
- Dubocovich ML (1979) Pharmacological differences between the alpha-presynaptic adrenoceptors in the peripheral and the central nervous systems. In: Langer SZ, Starke K, Dubocovich ML (eds) Presynaptic receptors. Pergamon Press, Oxford, pp 29-36
- Dubocovich ML, Langer SZ (1974) Negative feed-back regulation of noradrenaline release by nerve stimulation in the perfused cat's spleen: differences in potency of phenoxybenzamine in blocking the pre- and post-synaptic adrenergic receptors. J Physiol (Lond) 237:505-519
- Dubocovich ML, Langer SZ, Massingham R (1980) Lack of correlation between presynaptic inhibition of noradrenaline release and end organ responses during nerve stimulation. Br J Pharmacol 69:81-90
- Endo T, Starke K, Bangerter A, Taube HD (1977) Presynaptic receptor systems on the noradrenergic neurones of the rabbit pulmonary artery. Naunyn Schmiedebergs Arch Pharmacol 296:229-247
- Ennis C, Janssen PAJ, Schnieden H, Cox B (1979) Characterization of receptors on postganglionic cholinergic neurons in the guinea-pig isolated ileum. J Pharm Pharmacol 31:217-221
- Flavahan NA, McGrath JC (1980) Blockade by yohimbine of prazosin-resistant pressor effects of adrenaline in the pithed rat. Br J Pharmacol 69:355-357
- Franklin KBJ, Herberg LJ (1977) Presynaptic α-adrenoceptors: the depression of selfstimulation by clonidine and its restoration by piperoxane but not by phentolamine or phenoxybenzamine. Eur J Pharmacol 43:33–38
- Fügner A (1971) Antagonism of the drug induced behavioural sleep in chicks. Arzneim Forsch 21:1350-1352
- Fügner A, Hoefke W (1971) A sleep-like state in chicks caused by biogenic amines and other compounds; quantitative evaluation. Arzneim Forsch 21:1243-1247
- Fuller RW, Snoddy HD, Marshall WS (1977) Lowering of rat brain 3-methoxy-4hydroxyphenylethylene glycol sulphate (MOPEG sulphate) concentration by 2,6dichlorobenzylidene aminoguanidine. J Pharm Pharmacol 29:375-376
- Furchgott RF (1964) Receptor mechanisms. Annu Rev Pharmacol 4:21-50
- Furchgott RF (1972) The classification of adrenoceptors (adrenergic receptors). An evaluation from the standpoint of receptor theory. In: Blaschko H, Muscholl E (eds) Catecholamines. Springer, Berlin Heidelberg New York, pp 283-335
- Glossmann H, Hornung R (1980) Sodium ions increase the affinity of clonidine for  $\alpha_1$ -adrenoceptors in rat brain. Naunyn Schmiedebergs Arch Pharmacol 312:105–106

- Glossmann H, Presek P (1979) Alpha noradrenergic receptors in brain membranes: sodium, magnesium and guanyl nucleotides modulate agonist binding. Naunyn Schmiedebergs Arch Pharmacol 306:67-73
- Glusa E, Markwardt F, Barthel W (1979) Studies on the inhibition of adrenaline-induced aggregation of blood platelets. Pharmacology 19:196-201
- Göthert M, Lox HJ, Rieckesmann JM (1977) Effects of butyrophenones on the sympathetic nerves of the isolated rabbit heart and on the postsynaptic  $\alpha$ -adrenoceptors of the isolated rabbit aorta. Naunyn Schmiedebergs Arch Pharmacol 300:255–265
- Gold MS, Redmond DE, Kleber HD (1978) Clonidine blocks acute opiate-withdrawal symptoms. Lancet II:599-602
- Grant JA, Scrutton MC (1979) Novel  $\alpha_2$ -adrenoreceptors primarily responsible for inducing human platelet aggregation. Nature 277:659–661
- Green GJ, Wilson H, Yates MS (1979) The mechanism of the clonidine-induced reduction in peripheral parasympathetic submaxillary salivation. Eur J Pharmacol 56: 331-336
- Greenberg DA, Snyder SH (1978) Pharmacological properties of (<sup>3</sup>H)dihydroergokryptine binding sites associated with *alpha* noradrenergic receptors in rat brain membranes. Mol Pharmacol 14:38-49
- Greenberg DA, U'Prichard DC, Snyder SH (1976) Alpha-noradrenergic receptor binding in mammalian brain: differential labeling of agonist and antagonist states. Life Sci 19:69-76
- Greenberg DA, U'Prichard DC, Sheehan P, Snyder SH (1978) α-Noradrenergic receptors in the brain: differential effects of sodium on binding of (<sup>3</sup>H)agonists and (<sup>3</sup>H)antagonists. Brain Res 140:378-384
- Greengrass P, Bremner R (1979) Binding characteristics of <sup>3</sup>H-prazosin to rat brain αadrenergic receptors. Eur J Pharmacol 55:323-326
- Haeusler G (1974) Clonidine-induced inhibition of sympathetic nerve activity: no indication for a central presynaptic or an indirect sympathomimetic mode of action. Naunyn Schmiedebergs Arch Pharmacol 286:97–111
- Haeusler G (1976) Studies on the possible contribution of a peripheral presynaptic action of clonidine and dopamine to their vascular effects under in vivo conditions. Naunyn Schmiedebergs Arch Pharmacol 295:191–202
- Hardey DW, Lokhandwala MF (1979) Influence of prazosin on cardiac reflex function. Eur J Pharmacol 57:251-254
- Harper B, Hughes IE, Stott J (1979) Possible differences in α-adrenoceptors in rat, guinea-pig and rabbit spleen. J Pharm Pharmacol 31:105-107
- Hawkins M, Monti JM (1979) Effects of pretreatment with 6-hydroxydopamine or noradrenergic receptor blockers on the clonidine-induced disruption of conditioned avoidance responding. Eur J Pharmacol 58:53-58
- Heise A, Kroneberg G (1970) Periphere und zentrale Kreislaufwirkung des α-Sympathicomimeticums 2-(2,6-Xylidino)-5,6-dihydro-4H-1,3-thiazinhydrochlorid (BAY 1470). Naunyn Schmiedebergs Arch Pharmakol 266:350-351
- Heise A, Kroneberg G (1973) Central nervous α-adrenergic receptors and the mode of action of α-methyldopa. Naunyn Schmiedebergs Arch Pharmacol 279:285-300
- Hicks PE, Cannon JG (1979) NN-Dialkyl derivatives of 2-amino-5,6-dihydroxy-1,2,3, 4-tetrahydronaphthalene as selective agonists at presynaptic  $\alpha$ -adrenoceptors in the rat. J Pharm Pharmacol 31:494-496
- Hieble JP, Pendleton RG (1979) Effects of ring substitution on the pre- and postjunctional alpha-adrenergic activity of aryliminoimidazolidines. Naunyn Schmiedebergs Arch Pharmacol 309:217-224
- Hoffman BB, de Lean A, Wood CL, Schocken DD, Lefkowitz RJ (1979) Alpha-adrenergic receptor subtypes: quantitative assessment by ligand binding. Life Sci 24: 1739-1746
- Hornung R, Presek P, Glossmann H (1979) Alpha adrenoceptors in rat brain: direct identification with prazosin. Naunyn Schmiedebergs Arch Pharmacol 308:223-230

- Hoyer I, van Zwieten PA (1971) The centrally induced fall in blood pressure after the infusion of amphetamine and related drugs into the vertebral artery of the cat. J Pharm Pharmacol 23:892–893
- Hsu CY, Knapp DR, Halushka PV (1979) The effects of *alpha* adrenergic agents on human platelet aggregation. J Pharmacol Exp Ther 208:366–370
- Hunt GE, Atrens DM, Becker FT, Paxinos G (1978)  $\alpha$ -Adrenergic modulation of hypothalamic self-stimulation: effects of phenoxybenzamine, yohimbine, dexamphetamine and their interactions with clonidine. Eur J Pharmacol 53:1–8
- Jakobs KH, Saur W, Schultz G (1978) Characterization of α- and β-adrenergic receptors linked to human platelet adenylate cyclase. Naunyn Schmiedebergs Arch Pharmacol 302:285-291
- Jarrott B, Louis WJ, Summers RJ (1979) The characteristics of  $({}^{3}$ H)-clonidine binding to an  $\alpha$ -adrenoceptor in membranes from guinea-pig kidney. Br J Pharmacol 65: 663-670
- Jauernig RA, Moulds RFW, Shaw J (1978) The action of prazosin in human vascular preparations. Arch Int Pharmacodyn 231:81–89
- Johansson B, Johansson SR, Ljung B, Stage L (1972) A receptor kinetic model of a vascular neuroeffector. J Pharmacol Exp Ther 180:636–646
- Kapur H, Mottram DR (1978) A comparative study on the pre- and post-synaptic alpha blocking activity of a series of benzodioxanes. Biochem Pharmacol 27: 1879-1880
- Kilbinger H, Wessler I (1979) Increase by α-adrenolytic drugs of acetylcholine release evoked by field stimulation of the guinea-pig ileum. Naunyn Schmiedebergs Arch Pharmacol 309:255-257
- Kobinger W (1978) Central  $\alpha$ -adrenergic systems as targets for hypotensive drugs. Rev Physiol Biochem Pharmacol 81:39–100
- Kobinger W, Pichler L (1974) Evidence for direct α-adrenoceptor stimulation of effector neurons in cardiovascular centers by clonidine. Eur J Pharmacol 27:151-154
- Kobinger W, Pichler L (1980) Investigation into different types of post- and presynaptic α-adrenoceptors at cardiovascular sites in rats. Eur J Pharmacol 65:393-402
- Kobinger W, Walland A (1971) Involvement of adrenergic receptors in central vagus activity. Eur J Pharmacol 16:120-122
- Kosterlitz HW, Lydon RJ, Watt AJ (1970) The effects of adrenaline, noradrenaline and isoprenaline on inhibitory  $\alpha$ - and  $\beta$ -adrenoceptors in the longitudinal muscle of the guinea-pig ileum. Br J Pharmacol 39:398-413
- Langer SZ (1973) The regulation of transmitter release elicited by nerve stimulation through a presynaptic feed-back mechanism. In: Usdin E, Snyder SH (eds) Frontiers in catecholamine research. Pergamon Press, New York, pp 543-549
- Langer SZ (1974) Presynaptic regulation of catecholamine release. Biochem Pharmacol 23:1793-1800
- Langer SZ, Massingham R, Shepperson NB (in press) Differential sensitivity to prazosin blockade of endogenously released and exogenously administered noradrenaline: possible relationship to the synaptic location of  $\alpha_1$ - and the extrasynaptic location of  $\alpha_2$ -adrenoceptors in dog vascular smooth muscle. Br J Pharmacol
- Lasch P, Jakobs KH (1979) Agonistic and antagonistic effects of various &-adrenergic agonists in human platelets. Naunyn Schmiedebergs Arch Pharmacol 306:119-125
- Lee WH, Fujiwara M (1977) Untersuchungen zum Wirkungsmechanismus von Katecholaminen an der elektrisch gereizten isolierten Meerschweinchen-Gallenblase. Arzneim Forsch 27:1149-1152
- Lefèvre-Borg F, Roach AG, Cavero I (1978) Comparison of cardiovascular actions of dihydralazine, phentolamine, and prazosin in spontaneously hypertensive rats. J Cardiovasc Pharmacol 1:19-29
- Leighton J, Butz KR, Parmeter LL (1979) Effect of  $\alpha$ -adrenergic agonists and antagonists on neurotransmission in the rat anacoccygeus muscle. Eur J Pharmacol 58:27-38

- Lokhandwala MF, Coats JT, Buckley JP (1977) Effects of several catecholamines on sympathetic transmission to the myocardium: role of presynaptic  $\alpha$ -adrenoceptors. Eur J Pharmacol 42:257–265
- Madjar H, Docherty JR, Starke K (1980) An examination of pre- and postsynaptic  $\alpha$ adrenoceptors in the autoperfused rabbit hindlimb. J Cardiovasc Pharmacol 2: 619-627
- Malta E, McPherson GA, Raper C (1979) Comparison of pre-junctional  $\alpha$ -adrenoceptors at the neuromuscular junction with vascular post-junctional  $\alpha$ -receptors in cat skeletal muscle. Br J Pharmacol 65:249–256
- Marshall I, Nasmyth PA, Nicholl CG, Shepperson NB (1978) α-Adrenoceptors in the mouse vas deferens and their effects on its response to electrical stimulation. Br J Pharmacol 62:147-151
- Medgett IC, McCulloch MW, Rand MJ (1978) Partial agonist action of clonidine on prejunctional and postjunctional α-adrenoceptors. Naunyn Schmiedebergs Arch Pharmacol 304:215-221
- Miach PJ, Dausse JP, Meyer P (1978) Direct biochemical demonstration of two types of  $\alpha$ -adrenoreceptor in rat brain. Nature 274:492-494
- Miach PJ, Dausse JP, Cardot A, Meyer P (1980) <sup>3</sup>H-Prazosin binds specifically to 'α<sub>1</sub>'adrenoceptors in rat brain. Naunyn Schmiedebergs Arch Pharmacol 312:23-26
- Montel H, Starke K, Weber F (1974) Influence of morphine and naloxone on the release of noradrenaline from rat brain cortex slices. Naunyn Schmiedebergs Arch Pharmacol 283:257-269
- Morris BJ, Reid IA, Ganong WF (1979) Inhibition by  $\alpha$ -adrenoceptor agonists of renin release in vitro. Eur J Pharmacol 59:37-45
- Mouillé P, Huchet AM, Lucet B, Chelly J, Schmitt H (1979) Interactions between clonidine and α-adrenoceptor blocking drugs in the tachycardiac response to stimulation of the cardiac nerve in dogs. J Cardiovasc Pharmacol 1:515-528
- Mouillé P, Huchet AM, Chelly J, Lucet B, Doursout MF, Schmitt H (1980) Pharmacological properties of AR-C239, 2-[2-[4(o-methoxyphenyl)-piperazine-1-yl]-ethyl]-4,4-dimethyl-1,3(2H-4H)isoquinolinedione, a new α-adrenoceptor blocking drug. J Cardiovasc Pharmacol 2:175-191
- Moulds RFW, Jauernig RA (1977) Mechanism of prazosin collapse. Lancet I:200-201
- Nomura Y, Oki K, Segawa T (1980) Pharmacological characterization of central αadrenoceptors which mediate clonidine-induced locomotor hypoactivity in the developing rat. Naunyn Schmiedebergs Arch Pharmacol 311:41-44
- Oberdorf A, Kroneberg G (1970) Alpha- und beta-sympathicomimetische Wirkungen am elektrisch gereizten isolierten Meerschweinchenileum. Naunyn Schmiedebergs Arch Pharmakol 267:195-207
- Paton WDM, Vizi ES (1969) The inhibitory action of noradrenaline and adrenaline on acetylcholine output by guinea-pig ileum longitudinal muscle strip. Br J Pharmacol 35:10-28
- Peroutka SJ, Greenberg DA, U'Prichard DC, Snyder SH (1978) Regional variations in *alpha* adrenergic receptor interactions of (<sup>3</sup>H)-dihydroergokryptine in calf brain: implications for a two-site model of *alpha* receptor function. Mol Pharmacol 14: 403-412
- Pichler L, Kobinger W (1978) Presynaptic activity at peripheral adrenergic sites and blood pressure effect of  $\alpha$ -adrenoceptor stimulating drugs. Eur J Pharmacol 52: 287-295
- Pichler L, Placheta P, Kobinger W (1980) Effect of azepexole (B-HT 933) on pre- and postsynaptic α-adrenoceptors at peripheral and central nervous sites. Eur J Pharma- col 65:233-241
- Puech AJ, Lecrubier Y, Simon P (1979) Are alpha and beta presynaptic receptors involved in mood regulation? Pharmacological and clinical data. In: Langer SZ, Starke K, Dubocovich ML (eds) Presynaptic receptors. Pergamon Press, Oxford, pp 359-362

- Rand MJ, McCulloch MW, Story DF (1975) Pre-junctional modulation of noradrenergic transmission by noradrenaline, dopamine and acetylcholine. In: Davies DS, Reid JL (eds) Central action of drugs in blood pressure regulation. Pitman, Tunbridge Wells, pp 94–132
- Reynoldson JA, Head GA, Korner PI (1979) Effect of 6-hydroxydopamine on blood pressure and heart rate responses to intracisternal clonidine in conscious rabbits. Eur J Pharmacol 55:257-262
- Rhodes KF, Waterfall JF (1978) Pre- and postsynaptic α-adrenoceptor antagonism by indoramin in isolated tissues of the rat. J Pharm Pharmacol 30:516-517
- Roach AG, Lefèvre F, Cavero I (1978) Effects of prazosin and phentolamine on cardiac presynaptic  $\alpha$ -adrenoceptors in the cat, dog and rat. Clin Exp Hypertension 1: 87-101
- Robson RD, Antonaccio MJ, Saelens JK, Liebman J (1978) Antagonism by mianserin and classical α-adrenoceptor blocking drugs of some cardiovascular and behavioral effects of clonidine. Eur J Pharmacol 47:431-442
- Rossi EC, Louis G, Zeller EA (1979) Structure activity relationships between catecholamines and the α-adrenergic receptor responsible for the aggregation of human platelets by epinephrine. J Lab Clin Med 93:286-294
- Rouot BM, U'Prichard DC, Snyder SH (1980) Multiple  $\alpha_2$ -noradrenergic receptor sites in rat brain: selective regulation of high-affinity (<sup>3</sup>H)clonidine binding by guanine nucleotides and divalent cations. J Neurochem 34:374–384
- Ruffolo RR, Turowski BS, Patil PN (1977) Lack of cross-desensitization between structurally dissimilar  $\alpha$ -adrenoceptor agonists. J Pharm Pharmacol 29:378–380
- Ruffolo RR, Rosing EL, Waddell JE (1979) Receptor interactions of imidazolines. I. Affinity and efficacy for *alpha* adrenergic receptors in rat aorta. J Pharmacol Exp Ther 209:429-436
- Scatton B, Pelayo F, Dubocovich ML, Langer SZ, Bartholini G (1979) Effect of clonidine on utilization and potassium-evoked release of adrenaline in rat brain areas. Brain Res 176:197-201
- Schmitt H (1977) The pharmacology of clonidine and related products. In: Gross F (ed) Antihypertensive agents. Springer, Berlin Heidelberg New York, pp 299–396
- Schmitt H, Schmitt H, Fenard S (1971) Evidence for an α-sympathomimetic component in the effects of catapresan on vasomotor centres: antagonism by piperoxane. Eur J Pharmacol 14:98-100
- Scholtysik G, Lauener H, Eichenberger E, Bürki H, Salzmann R, Müller-Schweinitzer E, Waite R (1975) Pharmacological actions of the antihypertensive agent N-amidino-2-(2,6-dichlorophenyl)acetamide hydrochloride (BS 100-141). Arzneim Forsch 25:1483-1491
- Seno N, Nakazato Y, Ohga A (1978) Presynaptic inhibitory effects of catecholamines on cholinergic transmission in the smooth muscle of the chick stomach. Eur J Pharmacol 51:229-237
- Starke K (1972) Alpha sympathomimetic inhibition of adrenergic and cholinergic transmission in the rabbit heart. Naunyn Schmiedebergs Arch Pharmacol 274:18-45
- Starke K (1977) Regulation of noradrenaline release by presynaptic receptor systems. Rev Physiol Biochem Pharmacol 77:1-124
- Starke K, Langer SZ (1979) A note on terminology for presynaptic receptors. In: Langer SZ, Starke K, Dubocovich ML (eds) Presynaptic receptors. Pergamon Press, Oxford, pp 1-3
- Starke K, Montel H, Gayk W, Merker R (1974) Comparison of the effects of clonidine on pre- and postsynaptic adrenoceptors in the rabbit pulmonary artery. Naunyn Schmiedebergs Arch Pharmacol 285:133-150
- Starke K, Borowski E, Endo T (1975a) Preferential blockade of presynaptic α-adrenoceptors by yohimbine. Eur J Pharmacol 34:385-388

- Starke K, Endo T, Taube HD (1975b) Relative pre- and postsynaptic potencies of αadrenoceptor agonists in the rabbit pulmonary artery. Naunyn Schmiedebergs Arch Pharmacol 291:55-78
- Starke K, Taube HD, Borowski E (1977) Presynaptic receptor systems in catecholaminergic transmission. Biochem Pharmacol 26:259–268
- Steinsland OS, Hieble JP (1979) Activities of structural analogues of dopamine on prejunctional dopaminergic and alpha adrenergic receptors. In: Langer SZ, Starke K, Dubocovich ML (eds) Presynaptic receptors. Pergamon Press, Oxford, pp 93-98
- Stephenson RP (1956) A modification of receptor theory. Br J Pharmacol 11:379-393
- Steppeler A, Tanaka T, Starke K (1978) A comparison of pre- and postsynaptic αadrenergic effects of phenylephrine and tramazoline on blood vessels of the rabbit in vivo. Naunyn Schmiedebergs Arch Pharmacol 304:223-230
- Stjärne L (1975) Clonidine enhances the secretion of sympathetic neurotransmitter from isolated guinea-pig tissues. Acta Physiol Scand 93:142–144
- Story DD, Briley MS, Langer SZ (1979) The effects of chemical sympathectomy with 6-hydroxydopamine on α-adrenoceptor and muscarinic cholinoceptor binding in rat heart ventricle. Eur J Pharmacol 57:423-426
- Strömbom U, Svensson TH (1980) Clonidine: attenuation of sedative action by facilitated central noradrenergic neurotransmission. J Neural Transmission 47:29–39
- Tanaka T, Starke K (1979) Binding of <sup>3</sup>H-clonidine to an α-adrenoceptor in membranes of guinea-pig ileum. Naunyn Schmiedebergs Arch Pharmacol 309:207-215
- Tanaka T, Starke K (1980) Antagonist-/agonist-preferring  $\alpha$ -adrenoceptors or  $\alpha_1/\alpha_2$  adrenoceptors? Eur J Pharmacol 63:191–194
- Taube HD, Starke K, Borowski E (1977) Presynaptic receptor systems on the noradrenergic neurones of rat brain. Naunyn Schmiedebergs Arch Pharmacol 299: 123-141
- Timmermans PBMWM, van Zwieten PA (1980) Postsynaptic  $\alpha_1$  and  $\alpha_2$ -adrenoceptors in the circulatory system of the pithed rat: selective stimulation of the  $\alpha_2$ -type by B-HT 933. Eur J Pharmacol 63:199–202
- Timmermans PBMWM, Kwa HY, van Zwieten PA (1979a) Possible subdivision of postsynaptic α-adrenoceptors mediating pressor responses in the pithed rat. Naunyn Schmiedebergs Arch Pharmacol 310:189–193
- Timmermans PBMWM, Lam E, van Zwieten PA (1979b) The interaction between prazosin and clonidine at  $\alpha$ -adrenoceptors in rats and cats. Eur J Pharmacol 55:57–66
- Tsai BS, Lefkowitz RJ (1978) Agonist-specific effects of monovalent and divalent cations on adenylate cyclase-coupled *alpha* adrenergic receptors in rabbit platelets. Mol Pharmacol 14:540-548
- Tsai BS, Lefkowitz RJ (1979) Agonist-specific effects of guanine nucleotides on alpha-adrenergic receptors in human platelets. Mol Pharmacol 16:61-68
- U'Prichard DC, Snyder SH (1977) Binding of <sup>3</sup>H-catecholamines to α-noradrenergic receptor sites in calf brain. J Biol Chem 252:6450-6463
- U'Prichard DC, Snyder SH (1978) Guanyl nucleotide influences on <sup>3</sup>H-ligand binding to  $\alpha$ -noradrenergic receptors in calf brain membranes. J Biol Chem 253:3444-3452
- U'Prichard DC, Snyder SH (1979) Distinct  $\alpha$ -noradrenergic receptors differentiated by binding and physiological relationships. Life Sci 24:79–88
- U'Prichard DC, Greenberg DA, Snyder SH (1977) Binding characteristics of a radiolabeled agonist and antagonist at central nervous system *alpha* noradrenergic receptors. Mol Pharmacol 13:454-473
- U'Prichard DC, Charness ME, Robertson D, Snyder SH (1978) Prazosin: differential affinities for two populations of  $\alpha$ -noradrenergic receptor binding sites. Eur J Pharmacol 50:87-89
- Vizi ES (1979) Presynaptic modulation of neurochemical transmission. Prog Neurobiol 12:181-290
- Walland A (1978) Inhibition of a somato-sympathetic reflex via peripheral presynaptic  $\alpha$ -adrenoceptors. Eur J Pharmacol 47:211-221

Waud DR (1968) Pharmacological receptors. Pharmacol Rev 20:49-88

- Weitzell R, Tanaka T, Starke K (1979) Pre- and postsynaptic effects of yohimbine stereoisomers on noradrenergic transmission in the pulmonary artery of the rabbit. Naunyn Schmiedebergs Arch Pharmacol 308:127–136
- Westfall TC (1977) Local regulation of adrenergic neurotransmission. Physiol Rev 57: 659-728
- Wikberg J (1978a) Differentiation between pre- and postjunctional  $\alpha$ -receptors in guinea pig ileum and rabbit aorta. Acta Physiol Scand 103:225–239
- Wikberg JES (1978b) Pharmacological classification of adrenergic  $\alpha$  receptors in the guinea pig. Nature 273:164–166
- Wikberg JES (1979a) Pre- and postjunctional α-receptors. In: Langer SZ, Starke K, Dubocovich ML (eds) Presynaptic receptors. Pergamon Press, Oxford, pp 117-123
- Wikberg JES (1976b) The pharmacological classification of adrenergic  $\alpha_1$  and  $\alpha_2$  receptors and their mechanisms of action. Acta Physiol Scand Suppl 468
- Wikberg J, Andersson R, Lundholm L (1975) Differentiation of adrenergic  $\alpha$ -receptors in guinea pig intestine and rabbit aorta. Blood Vessels 12:383–384
- Wood CL, Arnett CD, Clarke WR, Tsai BS, Lefkowitz RJ (1979) Subclassification of alpha-adrenergic receptors by direct binding studies. Biochem Pharmacol 28:1277– 1282
- Young WS, Kuhar MJ (1979) Noradrenergic  $\alpha_1$  and  $\alpha_2$  receptors: autoradiographic visualization. Eur J Pharmacol 59:317–319
- Zebrowska-Lupina I, Przegalinski E, Sloniec M, Kleinrok Z (1977) Clonidine-induced locomotor hyperactivity in rats. Naunyn Schmiedebergs Arch Pharmacol 297: 227-231

# **Author Index**

Page numbers in *italics* refer to the bibliography.

Abraham P, McCallum WC, Docherty T, Newton P, Fox A 177, 183 Abraham P, McCallum W, Gourlay I 176, 177, 183 Abraham P, see McCallum WC 176, 192 Abrahams VC, Hilton SM, Zbrozyna A 52, 65, 97, 104 Achari NK, Al-Ubaidy SS. Downman CBB 65, 90, 91, 104 Achari NK, Downman CBB 65,90,91,104 Adair JR, Manning JW 37, 99,104 Adrian ED, Bronk DW, Phillips G 53, 58, 104 Adrian 141 Aghajanian GK, see Cedarbaum JM 34, 107, 210, 212, 229 Agostoni E, Chinnock JE, Daly M de B, Murray JG 75,104 Akert K, see Hess WR 21 Akimoto H, Saito Y 53, 104 Aktories K, Schultz G, Jakobs KH 226,228 Alanis J, Mascher D, Miyamoto J 64,104 Albefessard D, see Besson JM 146,184 Aldridge VJ, see Walter WG 127, 132, 159, 171, 181, 196 Aleonard P, see Besson JM 146, 184 Alexander RS 24, 29, 40, 43, 50, 53, 54, 58, 60, 62, 63, 66,104 Alexander S, see Kerr FWL 115 Algate DR, Waterfall JF 206, 228 Allen SL, see Marczynski TJ 146, 191 Allison T, see Wood CC 159, 196

Altansberger RA, see Kumada M 89, 98, 116 Al-Ubaidy SS, see Achari NK 65,90,91,104 Amendt K, Czachurski J, Dembowsky K, Seller H 32, 33,104 Amendt K, see Dembowsky K 69,71,109 Ancri D, see Gahery Y 36, 38, 42, 110 Andén NE, Corrodi H, Fuxe K, Hökfelt B, Hökfelt T, Rydin C, Svensson T 212, 213, 228 Andén NE, Gomes C, Persson B, Trolin G 213, 228 Andén NE, Grabowska M, Strömbom U 213, 228 Anderson FD, Berry OM 35, 104 Andersson B 18 Andersson R, see Wikberg J 210.236 Andréjak M, Pommier Y, Mouillé P, Schmitt H 211, 228 Angell-James JE 84, 105 Angell-James JE, Daly M de B 84,87,89,105 Angell-James JE, Daly M de B, Elsner R 87, 105 Angell-James JE, see Daly M de B 87, 108, 109 Angell-James JE, see Elsner N 87,110 Anrep GV, Pascual W, Rössler R 92, 93, 105 Antonaccio MJ, see Robson RD 206, 214, 234 Aranda Coddou L, see Baumgarten R von 28, 106 Aranibar A, see Pfurtscheller G 183, 193 Ariens EJ 200, 223, 228 Arndt JO, see Samodelov LF 84,121 Arnett CD, see Davis JN 218, 229

Arnett CD, see Wood CL 201. 221. 236 Arunlakshana O, Schild HO 202, 228 Ashton H, Millman IE, Rawlins MD, Telford R, Thompson IW 178, 183 Ashton H, Millman IE, Telford R, Thompson IW 178, 183 Aström KE 28, 105 Atrens DM, see Hunt GE 214,232 Attinger F, see Trzebski A 44.123 Auerswald W, see Hess WR 21 Aunon JI, Cantor F 138, 184 Bach LMN 50, 105 Bacq ZM 206, 228 Baertschi AJ, Munzer RF, Ward DG, Johnson RN, Gann DS 86, 105 Baertschi AJ, see Ward DG 86,123 Bagchi BK, see Kooi KA 137, 190 Bagshaw RJ, Iizuka M, Peterson LH 89, 98, 105 Bailey see Schaltenbrand 155 Bancaud J 150, 184 Bangerter A, see Endo T 202, 230 Baraban JM, see Davis M 214, 229 Barbas H, Dubrovsky B, Solyom L 176, 184 Barker JL, Crayton JW, Nicoll RA 52,105 Barlow JS 169, 184 Barman SM, Gebber GL 49, 50, 55, 59, 61, 67, 105 Barman SM, McCaffrey TV, Wurster RD 65, 105 Barman SM, Wurster RD 65,105 Barman SM, see Gebber GL 54, 55, 59, 61, 62, 111

Bouchard G, see Fukushima T

Barman SM, see McCall RS 42, 55, 58, 60, 61, 67, 68, 73,118 Barnes EM, see Cubeddu L 206, 229 Barnes P, Karliner J, Hamilton C, Dollery C 218, 228 Barnett U, see Katona P 74, 75, 80, 81, 93, 114 Barratt G, see Geis GS 55, 65,66,70,*111* Barren M, see Sutton S 127, 128, 131, 167, 195 Barthel W, see Glusa E 215, 231 Bartholini G, see Scatton B 214, 234 Bartorelli C, Bizzi E, Libretti A, Zanchetti A 53, 106 Bartorelli C, Weisschedel E 18 Bauer H, Lauber W 171, 184 Baumann P, see Delini-Stula A 209, 214, 229 Baumgarten R von, Aranda Coddou L 28, 106 Baumgarten R von, Kanzow E 33,95,106 Baumgarten R von, see Hellner K 41,85,112 Baust W, Heinemann H 53, 106 Baust W, Niemczyk H 44, 106 Baust W, Niemczyk H, Schaeffer H, Vieth J 44, 106 Bayliss WM 24, 106 Beattie J, Brow GR, Long CNH 65,106 Bechtereva NP 126, 181, 184 Beck EC, see Dustman RE 162,186 Becker FT, see Hunt GE 214, 232 Becker W, Hoehne O, Iwase K, Kornhuber HH 160, 184 Becker W, Kristeva R 174, 184 Becker W, see Deecke L 160, 186 Becker W, see Kornhuber HH 173.190 Bennett JA, Kidd C, Lafit AB, McWilliams PN 76, 77, 79, 106 Benvenuti P, see Zappoli R 177, 197 Berger 9, 12 Berger AJ 28, 106 Berthelsen S, Pettinger WA 199, 201, 210, 212, 226, 228

Berry OM, see Anderson FD 35.104 Besse JC, Furchgott RF 200, 222, 228 Besson JM, Woody CD, Aleonard P, Thompson HK, Albefessard D, Diarshall WH 146, 184 Bianchi AL 64, 106 Bickford RG, see Cant BR 159, 171, 184 Bignall KE, see Schramm LP 65, 121 Birbaumer N, Elbert Th, Lutzenberger W, Rockstroh B 171, 184 Biscoe JJ, Sampson SR 29, 106 Biscoe TJ, Sampson SR 36. 37, 38, 39, 40, 44, 45, 47, 85,106 Bizzi E, see Bartorelli C 53, 106 Blakeley AGH, Summers RJ 206, 228 Blatt S, see Cohen J 165, 185 Bohus B, see Jong W de 48, 109 Bok T 61, 106 Bolme P, Ngai SH, Rosen S 98,106 Bonvallet M, Dell P, Hiebel G 53,106 Bonvallet M, Sigg B 28, 106 Bonvallet M, see Dell P 53, 109 Borda RP 149, 159, 184 Borda RP, Hablitz JJ 141, 184 Borda RP, see Hablitz JJ 178, 188 Borda RP, see Low MD 140, 147, 149, 159, 164, 171, 191 Borda RP, see McSherry JW 145, 149, 173, 192 Borison HL, Domjan D 50, 77,87,106 Borison HL, see Wang SC 84, 85, 86, 87, 123 Borowski E, Starke K, Ehrl H, Endo T 202, 206, 228 Borowski E, see Starke K 203, 234 Borowski E, see Taube HD 209, 227, 235 Bostem F, Rousseau JC, Degossely M, Dongier M 175,184 Bostem F, see Dongier M 175,186

152, 187 Bowman JT, see Miller FR 77,118 Bragdon HR, see Chapman RM 167, 185 Brattström A, see Warzel H 84,123 Bremner R, see Greengrass P 218, 219, *231* Briani S, see Zappoli R 177, 197 Brickman AL, Kaufman MP, Petrik GK, Schneiderman N 38, 52, 90, *106* Brickman AL, see Gimpl MP 88,90,*111* Briley MS, see Story DD 227, 235 Brodal A 106 Brodal A, see Jansen J 91, 114 Bronk DW, Tower SS, Solandt DY 106 Bronk DW, Pitts RF, Larrabee MG 53,65,107 Bronk DW, see Adrian ED 53, 58, 104 Bronk DW, see Pitts RF 58, 59, 62, 97, 120 Brooks CMcC, see Koizumi K 53-55, 57, 115 Brow GR, see Beattie J 65, 106 Brown DA, Caulfield MP 212, 228 Brown GL, Eccles JC 75, 107 Brügger M, see Hess WR 20 Bucher V, see Hess WR 21 Buchsbaum MS, Henkin RI, Christiansen RL 162, 184 Buckley JP, see Lokhandwala MF 205, 233 Büch O, see Delini-Stula A 209, 214, 229 Bürgi S, see Hess WR 21 Bürgi Sandro 17 Bürki H, see Scholtysik G 213, 234 Burkhart SM Furnell L, Ledsome JR 48, 107 Burkhart SM, Ledsome JR 48,107 Burton H, see Loewy AD 32 - 34, 117Burton H, see Middleton S 41, 42, 118 Busch G, see Schulte FJ 53, 121 Butler M, Jenkinson DH 206, 228

Butz KR, see Leighton J 204, 206, 208, 232 Cahi CY, see Chen HI 77, 107 Cajal S, Ramon Y 27, 30, 35, 76,77,107 Calaresu FR, Cottle MK 77, 78,107 Calaresu FR, Henry JL 64, 107 Calaresu FR, Pearce JW 77, 78,107 Calaresu FR, see Chess GF 107 Calaresu FR, see Ciriello J 107 Calaresu FR, see Cottle MKW 27, 32, 33, 39, 108 Calaresu FR, see Henry JL 58,64,65,67,68,112 Calaresu FR, see Thomas MR 52, 68, 77, 78, 89, *123* Callaway E, Jones RT, Layne RS 137, 184 Callaway E, see Conchin E 139, 186 Cambridge D, Davey MJ, Massingham R 203, 228 Camerer H, Stroh-Werz M, Krienke B, Langhorst P 54,107 Camerer H, see Langhorst P 42-44,116 Camerer H, see Stroh-Werz M 41, 42, 94, 122 Cannon JG, see Hicks PE 206, 231 Cannon W 3, 9 Cannon WB 55, 107 Cant BR, Bickford RG 171, 184 Cant BR, Pearson JE, Bickford RG 159, 184 Cantor F, see Aunon JI 138, 184 Cardot A, see Miach PJ 217, 218, 219, 233 Carmeliet J, see Debecker J 141,*185* Caspers H 126, 128, 181, 184, 185 Caspers H, Speckmann EJ 144, 145, 185 Caspers H, see Hess WR 21 Caspers H, see Speckmann EJ 145, 181, 183, 194 Castro F de 26, 109 Caton R 126, 185 Caulfield MP, see Brown DA 212, 228

Cavero I, Dennis T, Lefèvre-Borg F, Perrot P, Roach AG, Scatton B 209, 228 Cavero I, Gomeni R, Lefèvre-Borg F, Roach AG 206, 228 Cavero I, Lefèvre F, Roach A 203, 228 Cavero I, see Lefèvre-Borg F 206, 209, 232 Cavero I, see Roach AG 209, 234 Cedarbaum JM, Aghajanian GK 34, 107, 210, 212, 229 Chai CY, Wang SC 49, 107 Chai CY, see Lee TM 50, 77, 116 Chai CY, see Wang SC 49, 123 Chapman RM, Bragdon HR 167, 185 Charness ME, see U'Prichard DC 217-219, 221, 235 Chelly J, see Mouillé P 206, 209, *233* Chen HI, Cahi CY 77, 107 Chess GF, Tam RMK, Calaresu FR 107 Chinnock JE, see Agostoni E 75,104Chiurugi E, Mollica A 77, 78, 107 Christiansen RL, see Buchsbaum MS 162, 184 Chung JM, Chung K, Wurster RD 58,107 Chung K, see Chung JM 58, 107 Ciriello J, Calaresu FR 107 Clarke WR, see Wood CL 201, 221, *236* Clausing KW, see De Vito JL 76,109 Clement DL, see Pelletier CL 84,119 Clineschmidt BV, Flataker LM, Faison EP, Haubrich DR 214, 229 Clineschmidt BV, Flataker LM, Faison E, Holmes R 214, 229 Coats AC, see Low MD 172, 175,191 Coats JT, see Lokhandwala MF 205, 233 Cohen DH, Schnall AM, MacDonald RL, Pitts LH 53,54,107 Cohen DH, see Schwaber JS 79, *121* Cohen J 137, 147, 159, 164, 165, 172, 173, 185

Cohen J, Offner F, Blatt S 165, 185 Cohen J, Offner F, Palmer C 165, 185 Cohen J, Walter WG 168, 185 Cohen L, see Smith DBD 167,194 Cohen MI, Gootman PM 53, 54,107 Cohen MI, see Gootman PM 43,53-55,66,67,111 Cohen S, see Peters JF 171, 193 Cole JO, see Tecce JJ 172, 178, 195 Coleridge JCG, see Coleridge HM 53,108 Coleridge HM, Coleridge JCG, Rosenthal F 53, 108 Constantine JW, Gunnell D, Weeks RA 216, 229 Constantine JW, Lebel W 216, 226, 229 Constantine JW, Weeks RA, McShane WK 206, 209, 229 Cooper 149 Cooper R, Crow HJ, Walter WG, Winter AL 147, 185 Cooper R, Osselton JW, Shaw JC 133, 138, 140, 185 Cooper R, see Donchin E 139,186 Cooper R, see Thompson JW 179, 195 Cooper R, see Walter WG 127, 132, 159, 171, 181, 196 Cooper R, see Weinberg H 131, 169, 196 Coote JH 56, 58, 89, 101, 108Coote JH, Downman CBB 55,56,67,108 Coote JH, Downman CBB, Weber WV 55-57, 108 Coote JH, Hilton SM, Perez-Gonzalez JF 89, 98, 99, 108 Coote JH, Hilton SM, Zbrozyna AW 98, 108 Coote JH, Macleod VH 51, 56, 57, 63, 65, 67-71, 108 Coote JH, Macleod VH, Martin TL 68, 108 Coote JH, Perez-Gonzalez JF 89,108 Coote JH, Sato A 56, 66-69, 108 Coote JH, Westbury DR 55, 58-60, 73, 89, 108

Coote JH, see Lipski J 47, 48, 54, 58, 66, 96, 117 Coquery JM, see Demaire C 168, 186 Corrodi H, see Andén NE 212, 213, 228 Costa LD, see Gilden L 127, 130, 147, 187 Costa LD, see Ritter W 127, 128, 131, 132, 168, 193 Costa LD, see Vaughan HG Jr 127, 130, 135, 148, 160, 161, 195, 196 Cottle MK 27, 31, 108 Cottle MKW, Calaresu FR 27, 32, 33, 39, 108 Cottle MK, see Calaresu FR 77,78,107 Courchesne E 162, 185 Courchesne E, Hillyard SA, Courchesne RY 162, 185 Courchesne E, Hillyard SA, Galambos R 131, 132, 157, 168, 185 Courchesne RY, see Courchesne E 162, 185 Cowan WN, see Saper C 65, 83, 101, 121 Cox A, see Warner HR 74, 123 Cox B, see Ennis C 211, 230 Crayton JW, see Barker JL 52,105 Creutzfeldt O, Kuhnt U 145, 185 Creutzfeldt OD, Kuhnt U 145, 185 Creutzfeldt O, see Watanabe S 145,196 Crill WE, Reis DJ 29-31, 108 Crill WE, see Weiss GK 99, 123 Crow HJ, see Cooper R 147, 185 Crow HJ, see Papakostopoulos D 146, 148, 149, 173, 192 Crow H, see Thompson JW 179, 195 Crow HJ, see Weinberg H 169, 196 Cubeddu L, Barnes EM, Langer SZ, Weiner N 206, 229 Cuenod M, see Reis DJ 51, 120 Culberson JL, Kimmel DL 27, 28, 108 Cummings JF, see Petras JM 58, 61, 120 Cummins B, see McCallum WC 177,192

Czachurski J, see Amendt K 32, 33, 104 Czacnurski J, see Dembowsky K 69, 71, 109 Dahlström A, Fuxe K 34, 51, 68.108 Daly JW, see Davis JN 219, 229 Daly M de B, Elsner R, Angell-James JE 108 Daly M de B, Korner PI, Angell-James JE, Oliver JR 87,109 Daly M de B, see Agostoni E 75.104 Daly M de B, see Angel-James JE 84, 87, 89, 105 Daly M de B, see Elsner N 87,110 Danahy T, see Ellingson RJ 137, 186 Dargent JS, Timsit M 175, 176, 195 Dauchot PJ, see Katona PG 74,75,80,81,95,*114* Dausse JP, see Miach PJ 217–219, 221, 233 Davey MJ, see Cambridge D 203, 228 Davidson NS, Goldner S, McCloskey DI 74, 80, 81, 89,93,109 Davies RO, Edwards MW 40, 109 Davis AL, McCloskey DI, Potter EK 96, 109 Davis H, Davis PA, Loomis AL, Harvey EN, Hobart G 126,185 Davis JN, Arnett CD, Hoyler E, Stalvey LP, Daly JW, Skolnik P 218, 229 Davis M, Redmond DE, Baraban JM 214, 229 Davis PA, see Davis H 126, 185 Dawkins EA, see Ingram WR 27,113 Day MD, Roach AG 25, 109 Debecker J, Carmeliet J 141, 185 Debecker J, see Desmeth JE 146, 181, 186 Deck R, Oberdorf A, Kroneberg G 210, 229 Deecke L, Becker W, Grözinger G, Scheid P, Kornhuber H 160,186 Deecke L, Englitz HG, Schmitt G 166, 167, 186

Deecke L, Grözinger B, Kornhuber HH 135, 186 Deecke L, Grözinger B, Kornhuber HH, Kriebel I, Kristeva R 160, 167, 186 Deecke L, Kornhuber HH 134-136, 160, 185 Deecke L, Kornhuber HH, Schmidt G 177, 186 Deecke L, Scheid P, Kornhuber HH 130, 160, 162, 185 Deecke L, see Kornhuber HH 127, 130, 135, 147, 148, 173,190 Degeest H, see Levy MN 86, 116 Degossely M, see Bostem F 175,184 Delanoy J, see Dongier M 175,176,186 Delaunoy J, see Timsit-Berthier M 137, 175, 176, 195 Delbarre B, Schmitt H 199, 212, 214, 229 Deliege M, see Faidherbe J 172,187 Delini-Stula A, Baumann P, Büch O 209, 214, 229 Dell P, Bonvallet M 53, 109 Dell P, see Bonvallet M 53, 106 Demaire C, Coquery JM 168, 186 Dembowsky K, Czacnurski J, Amendt K, Seller H 69, 71, 109 Dembowsky K, see Amendt K 32, 33, 104 Dennis T, see Cavero I 209, 228 Denoth F, see Zappoli R 177, 179, 197 Desmeth JE, Debecker J 146, 181,186 Desmet JE, see Donchin E 139,186 Devaux-Pasque N, see Faidherbe J 172, 187 Diarshall WH, see Besson JM 146,184 Dinko SJ, see Ford JM 187 Dittmar K, see Langhorst P 42-44,116 Djojosugito A, Folkow B, Lisander B, Sparks H 101, 109 Djojosugito AM, Folkow B, Kvlstra PH. Lisander B. Tuttle RS 89, 98, 109 Doba N, Reis DJ 48-50, 109

Doba N, see Reis DJ 48, 49, 120 Docherty JR, MacDonald A, McGrath JC 216, 226, 229 Docherty JR, McGrath JC 209, 216, 229 Docherty JR, see Madjar H 216, 233 Docherty T, see Abraham P 177, 183 Dollery C, see Barnes P 218, 228Domjan D, see Borison HL 50, 77, 87, 106 Donchin E 169, 186 Donchin E, Callaway E, Cooper R, Desmet JE, Goff WR, Hillyard SA, Sutton S 139, 186 Donchin E, Otto D, Gerbrandt LK, Pribram K 149, 186 Donchin E, Tueting P, Ritter Q, Kutes M, Hoffley E 157, 186 Donchin E, see Kutas M 174, 190 Donchin E, see Smith DBD 167, 194 Donchin E, see Squires KC 167, 168, 194 Dongier M, Bostem F 175, 186 Dongier M, Koninckx N 176, 186 Dongier M, Timsit-Berthier M, Koninckx N, Delanoy J 175, 176, 186 Dongier M, see Bostem F 175,184 Dongier M, see Dubrovsky B 177,186 Dongier M, see Timsit M 175, 176,195 Donoghue S 31, 38, 86, 109 Donoghue S, Fox RE, Kidd C 29-31,109 Donoghue S, Kidd C, McWilliam PN 38, 109 Douglas WW, Ritchie JM 26, 110 Douglas WW, Ritchie JM. Schaumann W 26, 110 Douglas WW, Schaumann W 26, 110Doursout MF, see Mouillé P 206.233 Downman CBB 25, 110 Downman CBB, see Achari NK 65,90,91,104 Downman CBB, see Coote JH 55-57,67,108

Doxey JC 205, 229 Doxey JC, Everitt J 209, 229 Doxey JC, Everitt J, Metcalf G 206, 230 Doxey JC, Smith CFC, Walker JM 203, 204, 206, 230 Drane JW, see Kent BB 51, 115 Drew GM 204, 206-211, 216, 230 Drew GM, Gower AJ, Marriott AS 214, 230 Drew GM, Whitting SB 214, 215,230 Dubocovich ML 209, 230 Dubocovich ML, Langer SZ 206,230 Dubocovich ML, Langer SZ, Massingham R 207, 230 Dubocovich ML, see Scatton B 214, *234* Dubois FS, see Foley JO 27, 110 Dubrovsky B, Garcia-Rill E, Dongier M 177, 186 Dubrovsky B, see Barbas H 176,184 Dugin SF, Zakharow ST, Samonina GE, Udelnov MG 77,78,110 Dustman RE, Beck EC 162, 186 Eccles JC, see Brown GL 75, 107 Edwards MW, see Davies RO 40,109 Ehrl H, see Borowski E 202, 206, 228 Eichenberger E, see Scholtysik G 213, 234 Eisengart MA, see Symmes D 168,195 Elbert Th, see Birbaumer N 171, 184 Elliason S, Folkow B, Lindgren P, Üvnas B 97, 110 Ellingson RJ, Lathrop GH, Danahy T, Nelson B 137, 186 Elsner N, Angell-James JE, Daly M de B 87, 110 Elsner R, see Angell-James JE 87,105 Elsner R, see Daly M de B 108 Endo T, Starke K, Bangerter A, Taube HD 202, 230 Endo T, see Borowski E 202, 206, 228

Endo T, see Starke K 202-204, 208, 234, 235 Englitz HG, see Deecke L 166, 167, 186 Ennis C, Janssen PAJ, Schnieden H, Cox B 211, 230Ervin FR, Mark VH 152, 187 Euler C von, Hayward JN, Mattila I, Wyman R 33, 64, 110 Evans MH 88, 110 Everitt J, see Doxey JC 206, 209, 229, 230 Faidherbe J, Deliege M, Devaux-Pasque N 172, 187 Faison E, see Clineschmidt BV 214,229 Faison EP, see Clineschmidt BV 214, 229 Feigl E, Johansson B, Lofving B 89, 99, 110 Fenelon B 177, 187 Fenelon B, see Papakostopoulos D 159, 171, 192 Ferguson JH, Jasper HM 146, 187 Ferguson JP, see Rhoton AL 28,120 Fernard S, see Schmitt H 212, 234 Fernandez de Molina A, Kuno M, Perl E 55, 58-60, 110 Fidone SJ, Sato A 26, 29, 110 Finger H, see Otto D 163, 178, 192 Finkenzeller P, see Klinke R 131, 169, 190 Fischer H 15 Flataker LM, see Clineschmidt BV 214, 229 Flavahan NA, McGrath JC 216, 230 Fleisch A 17 Fleisch Alfred 2 Foley JO, Dubois FS 27, 110 Folkow B, Lisander B, Tuttle RS, Wang SC 89, 110 Folkow B, Öberg B, Rubinstein EH 89, 99, 110 Folkow B, see Djojosugito A 101.109 Folkow B, see Djojosugito AM 89, 98, 109 Folkow B, see Elliason S 97, 110 Fontaine O, see Timsit M 175, 176, 195

Geis GS, Wurster RD 76, 79,

Ford JM, Roth WT, Dinko SJ, Kopell BS 187 Ford JM, see Roth WT 168, 193 Foreman RD, Wurster RD 55, 65, 70, 110 Fox A, see Abraham P 177, 183 Fox RE, see Donoghue S 29-31,109 Franklin KBJ, Herberg LJ 214,230 Franz GN, see Wilson MF 89, 98, 124 Friedman J, Goldberg H, Horvath TB, Meares RA 178,187 Frost JD Jr, see Low MD 140, 147, 159, 164, 171, 191 Fruhstorfer H, see Järvilehto T 159, 172, 173, 189 Fruhstorfer H, see Klinke R 131, 169, 190 Fügner A 214, 230 Fügner A, Hoefke W 213, 230 Fujiwara M, see Lee WH 210, 232 Fukushima T, Mayanagi Y, Bouchard G 152, 187 Fuller RW, Snoddy HD, Marshall WS 213, 230 Furchgott RF 200, 207, 216, 223, *230* Furchgott FR, see Besse JC 200, 222, 228 Furnell L, see Burkhart SM 48,107 Fussey IF, Kidd C, Whitwam JG 71, 86, 110 Fuxe K, see Andén NE 212, 213, 228 Fuxe K, see Dahlström A 34, 51,68,108 Gabriel M, Seller H 29, 30, 36, 37, 48, 85, 110 Gabriel M, see Illert M 65, 68, 69, 113 Gabriel M, see Richter DW 64,120 Gahery Y, Ancri D 36, 38, 42,110 Galambos R, see Courchese E 131,132,157,168,175 Galambos R, see Hillyard SA 141, 171, 189 Gandevia SC, McCloskey DI, Potter EK 89, 110 Ganglberger JA 148, 187

Ganglberger JA, Gestring GF, Groll-Knapp E, Haider M 152, 159, 187 Ganglberger JA, Haider M, Knapp E, Schmid H 136, 148, 156, 182, 187 Ganglberger JA, see Groll-Knapp E 145, 146, 148, 149, 151, 152, 154, 159, 161, 173, 182, 187 Ganglberger JA, see Haider M 145, 146, 149, 152, 154, 173, 182, 188, 189 Ganglberger JA, see Knapp E 174,190 Gann DS, see Baertschi AJ 86,105 Gann DS, see Ward DG 86, 123 Ganong WF, see Morris BJ 226, 233 Garcia M, Jordan D, Schneiderman N, Spyer KM 28, 76, 77, 95, 111 Garcia M, Jordan D, Spyer KM 28, 31, 34, 111 Garcia-Rill E, see Dubrovsky B 177, 186 Gaule 2, 3 Gayk W, see Starke K 202, 203, 207, 208, 234 Gebber GL 54,55,66,67,111 Gebber GL, Barman SM 54, 55, 59, 61, 62, 111 Gebber GL, Barman SM, McCall RB 59, 61, 62, 111 Gebber GL, Klevans LR 87, 88,111 Gebber LG, McCall RS 55, 58-63,65-68,111 Gebber GL, Snyder DW 89, 111 Gebber GL, Taylor DG, Weaver LC 63, 65, 66, 67,111 Gebber GL, see Barman SM 49, 50, 55, 59, 61, 67, 105 Gebber GL, see Klevans LR 87, 88, 115 Gebber GL, see McCall RS 42, 55, 58, 60, 61, 67, 68, 73,118 Gebber GL, see Quest JA 89, 120 Gebber GL, see Snyder DW 54, 55, 59, 63, 68, 122 Gebber GL, see Taylor DG 53-55, 58-60, 63, 66-68, 123 Geddes LA 138, 187 Geis GS, Baratt G, Wurster RD 55,65,66,70,111

111 Geller E, see Kezdi P 56, 115 Gellhorn E 51, 52, 87, 99, 111 Gellhorn E, Nakao H, Redgate ES 51, 52, 87, 99, 111 Gerbrandt LK, Groff WR, Smith DB 130, 135, 136, 160, 161, 187 Gerbrandt LK, see Donchin E 149,186 Gestring GF, see Ganglberger JA 152, 159, 187 Gestring GF, see Haider M 149,188 Ghelarducci B, Pompeiano O, Spyer KM 91, 111 Gilden L, Vaughan HG Jr, Costa LD 127, 130, 147, 187 Gimpl MP, Brickman AL, Kaufman MP, Schneiderman N 88,90,*111* Girton DG, Kamiya J 141, 187 Giuliano G, see Zappoli R 179,197 Glees P, Nauta WJH 27, 111 Glötzner F, Grüsser OJ 145, 187 Glossmann H, Hornung R 222, 230 Glossmann H, Presek P 222, 226, 231 Glossmann H, see Hornung R 217, 218, 219, 222, 231 Glusa E, Markwardt F, Barthel W 215, 231 Godehard E, see Samodelov LF 84,121 Göthert M, Lox HJ, Rieckesmann JM 206, 231 Goff WR, see Donchin E 139,186 Goff WR, see Gerbrandt LK 130, 135, 136, 160, 161, 187 Goff WR, see Wood CC 159, 196 Gold MS, Redmond DE, Kleberg HD 227, 231 Goldberg H, see Friedman J 178, 187 Goldner S, see Davidson NS 74,80,81,89,93,109 Goldring S 145, 187 Goldring S, see Karahashi Y 145,189 Goldring S, see O'Leary JL 126, 181, 192

Gomeni R, see Cavero I 206, 228 Gomes C, see Andén NE 213, 228 Gootman PM, Cohen MI 53-55,66,67,111 Gootman PM, Cohen MI, Piercey MP, Wolotsky P 43,111 Gootman PM, see Cohen MI 53, 54, 107 Gourlay I, see Abraham P 176, 177, 183 Gower AJ, see Drew GM 214, 230 Grabowska M, see Andén NE 213, 228 Grant JA, Scrutton MC 214, 215, 231 Green GJ, Wilson H, Yates MS 210, 211, 231 Green JH 26, 74, 80, 112 Green JH, Heffron PF 54, 56,112 Greenberg DA, Snyder SH 219, 221, 231 Greenberg DA, U'Prichard DC, Sheehan P, Snyder SH 219, 222, 231 Greenberg DA, U'Prichard DC, Snyder SH 219, 231 Greenberg DA, see Peroutka SJ 219, 221, 233 Greenberg DA, see U'Prichard DC 217-220, 225, 227, 235 Greengrass P, Bremner P 218, 219.231 Gregor M, Janig W, Wiprich L 53, 54, 112 Griffith HB, see McCallum WC 130, 145, 146, 153, 154, 160, 182, 192 Groat WC de, Lalley PM 26, 29, 36, 109 Grösse M, Janig W 53, 54, 112 Grözinger B, see Deecke L 135, 160, 186 Grözinger G, see Deecke L 160, 167, 186 Groll-Knapp E, Ganglberger JA, Haider M 145, 146, 148, 149, 151, 152, 154, 159, 161, 173, 182, 187 Groll-Knapp E, Haider M, Höller H, Jenkner G, Neuberger M, Stidl H 179, 187 Groll-Knapp E, Wagner H, Hauck H, Haider M 179, 187

Groll-Knapp E, see Ganglberger JA 152, 159, 187 Groll-Knapp E, see Haider M 127, 130-132, 145-149, 152, 154, 168, 170, 173, 182, 188, 189 Grossman RG, Hampton T 145,188 Grundfest H 75, 112 Grundfest H, see Middleton S 75,118 Grünewald G, Grünewald-Zuberbier E, Hömberg V, Netz J 174, 188 Grünewald G, Grünewald-Zuberbier E, Netz J 130, 177,188 Grünewald G, Grünewald-Zuberbier E, Netz J, Hömberg V, Sander G 171, 174, 188 Grünewald G, see Grünewald-Zuberbier E 130, 136, 137, 177,*188* Grünewald-Zuberbier E, Grünewald G 130, 136, 137,188 Grünewald-Zuberbier E, Grünewald G, Jung R 136, 174, 188 Grünewald-Zuberbier E, Grünewald G, Rasche A Netz J 136, 174, 177, 188 Grünewald-Zuberbier E, see Grünewald G 130, 171, 177, 188 Grüsser OJ, see Glötzner F 145,187 Guazzi M, Zanchetti A 53, 112 Guerri S, see Zappoli R 179, 197 Gullickson GR 159, 164, 166, 167, 171, 188 Gumnit RJ 126, 188 Gunn CG, Sevelinus G, Puiggari MS, Myers FK 77, 112 Gunnell D, see Constantine JW 216, 229 Gurevitch MI, Vyshatina AI 90,112 Haab 2 Hablitz JJ, Borda RP 178, 188 Hablitz JJ, see Borda RP 141. 184 Hablitz JS 159, 188 Hackett CJ, see Marczynski TJ 146, 191

Haeusler G 25, 71, 112, 213, 226, *231* Hagbarth RE, Vallbo AB 53, 112 Haider M 151, 159, 169, 172, 181, 188 Haider M, Ganglberger JA, Gestring GF, Groll-Knapp E 149, 188 Haider M, Ganglberger JA, Groll-Knapp E 145, 146, 152, 182, 188, 189 Haider M, Ganglberger JA, Groll-Knapp E, Schmid H 149, 152, 154, 173, 182, 189 Haider M, Groll-Knapp E 170, 188 Haider M, Groll-Knapp E, Studynka G 127, 130–132, 147, 168, 188 Haider M, see Ganglberger JA 136, 148, 152, 156, 159, 182,187 Haider M, see Groll-Knapp E 145, 146, 148, 149, 151, 152, 154, 159, 161, 173, 179, 182, 187 Haider M, see Knapp E 174, 190 Haines E, see Wilkinson RT 172, 196 Halushka PV, see Hsu CY 214, 215, 232 Hamilton BT, see Tecce JJ 172.195 Hamilton C, see Barnes P 218, 228 Hamilton RB, Wallach JH, Petrik GK, Schneiderman N 112Hamilton RB, see Kaufman MP 52, 79, 88, 114 Hampton T, see Grossman RG 145,188 Hardey DW, Lokhandwala MF 227, 231 Harper B, Hughes IE, Scott J 226, 231 Hart S, see Otto D 163, 178, 192 Harter MR, Salomon LE 131, 189 Harvey EN, see Davis H 126, 185 Hassler R 155, 182, 189 Hassler R, see Hess WR 21 Haubrich DR, see Clineschmidt BV 214, 229 Hauck H, see Groll-Knapp E 179,187

Hauck H, see Groll-Knapp E 179, 187 Hausman J, see Timsit-Berthier M 164, 176, 195 Hawkins M, Monti JM 214, 231 Haymett BT, McCloskey DI 89,93,112 Hayward JN, see Euler C von 33, 64, 110 Hazemann PS, Metral S, Lille F 174, 189 Head GA, see Reynoldson JA 213, 234 Heffron PF, see Green JH 54, 56, 112 Heinbecker P 75, 112 Heinbecker P, O'Leary J 75, 112 Heinemann H, see Baust W 53,106 Heise A, Kroneberg G 212, 231 Held R, see Köhler W 126, 190 Hellner K, Baumgarten R von 41,85,112 Henatsch HD, see Schulte FJ 53, 121 Henkin RI, see Buchsbaum MS 162, 184 Henry JL, Calaresu FR 58, 64, 65, 67, 68, 112 Henry JL, see Calaresu FR 64,107 Herberg LJ, see Franklin KBJ 214,230 Hering R, see Squires KC 167, 168, 194 Hess WR 1-19, 20 Hess WR, Akert K 21 Hess WR, Akert K, McDonald DA 21 Hess WR, Auerswald W 21 Hess WR, Brügger M 20 Hess WR, Bürgi S, Bucher V 21 Hess WR, Caspers H, Hess R, Kugler J, Petsche O, Struppler A 21 Hess WR, Hassler R 21 Hess WR, Weisschedel E 21 Hess WR, Wyss OAM 20 Hess R, see Hess WR 21 Hetzel R, see Weidenger H 74,80,81,123 Heyde F, see Richter DW 64, 120Heymans C, Neil E 24, 26, 53, 73, 74, 86, 112

Hicks PE, Cannon JG 206, 231

Hiebel G, see Bonvallet M 53,106 Hieble JP, Pendleton RG 206,231 Hieble JP, see Steinsland OS 206,235 Hilbert RP, see Naitoh P 177, 192 Hildebrandt JR 36, 39, 40, 51, 67, 113 Hillyard SA 167, 170–173, 189 Hillyard SA, Galambos R 141, 171, 189 Hillyard SA, Hink RF, Schwent UL, Pieton TW 168, 189 Hillyard SA, see Courchese E 131, 132, 157, 162, 168, 185 Hillyard SA, see Donchin E 139,186 Hillyard SA, see Picton TW 169,193 Hillyard SA, see Squires KC 132, 167, 194 Hillyard SA, see Squires NK 131, 167, 168, 194 Hilton SM 25, 51, 62, 89, 96, 98, 99, 104, 113 Hilton SM, McAllen RM, Syper KM 49, 91, 113 Hilton SM, Spyer KM 39, 48, 50-52, 67, 88, 99, 113 Hilton SM, Spyer KM, Timms RJ 91, 113 Hilton SM, see Abrahams VC 52, 65, 97, 104 Hilton SM, see Coote JH 89, 98,99,108 Hink RF, see Hillyard SA 168, 189 Hirose K 32, 113 Hobart G, see Davis H 126, 185 Hockman CH, Talesnik J, Livingstone KE 88,90, 113 Hoefke W, see Fügner A 213, 230 Hoehne O, see Becker W 160, 184 Hoehne O, see Kornhuber HH 173, 190 Hökfelt B, see Andén NE 212, 213, 228 Hökfelt T, see Andén NE 212, 213, 228 Höller H, see Groll-Knapp E 179,187 Hömberg V, see Grünewald G 171, 174, 188

Hoffer BJ, Ratcheson R, Snider RS 101, 113 Hoffley E, see Donchin E 157, 186 Hoffman BB, Lean A de, Wood CL, Schocken DD, Lefkowitz RJ 221, 231 Hollister LW, see Kopell BS 178,190 Holmes R, see Clineschmidt BV 214, 229 Holst 8, 12, 14 Homma S, Miura M, Reis DJ 40,113 Hongo T, Ryall RW 58, 71, 113 Horeyseck G, Janig W, Kirchner F, Thamer V 53, 54,98,101,113 Honrung R, Presek P, Glossmann H 217–219, 222, 231 Hornung R, see Glossmann H 222,230 Horvath TB, see Friedman J 178,187 Houck K, see Otto D 163, 178, 192 Hoyer I, Zwieten PA van 212, 232 Hoyler E, see Davis JN 218, 229 Hsu CY, Knapp DR, Halushka PV 214, 215, 232 Huchet AM, see Mouillé P 206,209,*233* Hughes IE, see Harper B 226, 231 Humbertson AO, see Martin GF 69,117 Humphrey DR 36, 37, 39-42, 44-47, 50, 86, 113 Humphreys PW, Joels N, McAllen RM 89, 98, 113 Hunsperger RW 17 Huntsberger GE, see Roth WT 137, 193 Hunt GE, Atrens DM, Becker FT, Paxinos G 214, 232 Iizuka M, see Bagshaw RJ 89, 98,105 Iliukina WA 152, 182, 189 Illert M, Gabriel M 65, 68, 69.113 Illert M, Seller H 69, 70, 113 Illert M, see Seller H 33, 37, 42, 85, 96, 121 Ingram WR, Dawkins EA 27, 113

Iriuchijima J, Kumada M 74, 75, 80, 81, 93, 114 Irwin DA, Knott JR, McAdam DW, Rebert CS 171, 189 Irwin DA, see Knott JR 175, 190 Irwin DA, see Rebert CS 139, 171, 172, 193 Ise H, see Todo K 76, 79, 123 Ivins J, see Karrer R 163-165,190 Iwase K, see Becker W 160, 184 Iwase K, see Kornhuber HH 173, 190 Järvilehto T, Fruhstorfer H 159, 172, 173, 189 Jakobs KH, Saur W, Schultz G 214, 232 Jakobs KH, see Aktories K 226, 228 Jakobs KH, see Lasch P 215, 232 Janig W, Kümmel H 54, 114 Janig W, see Gregor M 53, 54, 112 Janig W, see Grösse M 53, 54, 112 Janig W, see Horeyseck G 53, 54,98,101,*113* Jankowska E, Roberts WJ 28.114 Jansen J, Brodal A 91, 114 Janssen PAJ, see Ennis C 211, 230 Janzen RW, see Speckmann EJ 145,194 Jarrott B, Louis WJ, Summers RJ 217, 219, 232 Jasper HH 139, 182, 189 Jasper HM, see Ferguson JH 146,187 Jauernig RA, Moulds RFW, Shaw J 215, 232 Jauernig RA, see Moulds RFW 215, 233 Jaussi 5 Jenkinson DH, see Butler M 206, 228 Jenkner G, see Groll-Knapp E 179, 187 Jenni 11 Jewett DL 74, 75, 79-81, 114 Jih F, see Katona PG 92, 114 Joels N, see Humphreys PW 89,98,*113* Johansson B, Johansson SR, Ljung B, Stage L 222, 232

99,110 Johansson SR, see Johansson B 222, 232 John ER, see Sutton S 127, 128, 131, 167, 169, 195 Johnson LC, see Naitoh P 177, 192 Johnson RN, see Baertschi AJ 86,105 Jones A, see Trzebski A 44, 123 Jones RT, see Callaway E 137, 184 Jong W de, Nijkamp P, Bohus B 48, 109 Jong W de, Palkovits M 48, 109 Jong W de, Zandberg P, Bohus B 48,109 Jong W de, Zandberg P, Palkovits M, Bohus B 48, 109 Jong W de, see Zandberg P 48, 49, *124* Jordan D 28, 31, 85, 104, 114 Jordan D, Khalid MEM, Schneiderman N, Spyer KM 80, 85, 88, 100, 114 Jordan D, Spyer KM 28-31, 34, 40, 50, 85, 86, 89, 93, 98, 99, 114 Jordan D, see Garcia M 28, 31, 34, 76, 77, 95, 111 Jordan D, see McAllen RM 26, 28-30, 39, 83, 118 Jouvet M, see Sakai K 34, 121 Judy WV, see Wilson MF 89, 98,124 Jung R, see Grünewald-Zuberbier E 136, 174, 188 Jus A, see Jus K 171, 189 Jus K, Kiljan A, Kubacki A, Rzepecki J, Wilczsak H, Jus A 171,*189* Jusseaume P, see Lelord G 174, 191 Kahn N, Mills E 62, 63, 114 Kahn N, see Katz RL 49, 114 Kalia M 32, 34, 114 Kamiya J, see Girton DG 141, 187 Kannan H, Yagi K 52, 114 Kanzow E, see Baumgarten R von 33,95,*106* Kapur H, Mottram DR 206, 232 Karahashi Y, Goldring S 145, 189

Johansson B, see Feigl E 89,

Karlin L 167, 189 Karlin L, Martz MJ 167, 189 Karlin L, Martz MJ, Mordkoff AM 146,190 Karliner J, see Barnes P 218, 228 Karplus, Kreidl 10 Karppanen H, see Ylitalo P 49,*124* Karrer R, Ivins J 163-165, 190 Karrer R, Warren C, Ruth R 164, 166, 167, 190 Karten HJ, see Katz DM 28, 114 Kastella KG, see Weiss GK 39, 123 Katona PG, Jih F 92, 114 Katona PG, Lipson D, Dauchot PJ 74, 75, 80, 81, 95,*114* Katona P, Poitras J, Barnett U, Terry B 74, 85, 80, 81, 93, 114 Katona PG, Tan KS 84, 114 Katz DM, Karten HJ 28, 114 Katz RL, Kahn N, Wang SC 49,114 Kaufman A, see Koizumi K 53-55, 57, 115 Kaufman MP, Hamilton RB, Wallach JH, Petrik GK, Schneiderman N 52, 79, 88,114 Kaufman MP, see Brickman AL 38, 52, 90, 106 Kaufman MP, see Gimpl MP 88,90,111 Keck W, see Richter DW 54, 56,96,120 Keith IC, Kidd C, Linden RJ, Snow HM 86, 89, 115 Keith IC, Kidd C, Penna PE 86, 89, 115 Kellaway P, see Low MD 140, 147, 149, 159, 164, 171, 191 Kent BB, Drane JW, Manning JW 51,115 Kerr FWL 28, 50, 76, 77, 115 Kerr FWL, Alexander S 115 Kezdi P 24, 25, 115 Kezdi P, Geller E 56, 115 Khalid MEM, see Jordan D 80, 85, 88, 100, 114 Kidd, Penna 89, 98 Kidd C, see Bennett JA 76, 77, 79, 106 Kidd C, see Donoghue S 29-31, 38, 109

Kidd C, see Fussey IF 71, 86, 110 Kidd C, see Keith IC 86, 89, 115 Kilbinger H, Wessler I 211, 232 Kiljian A, see Jus K 171, 189 Kimmel DB, see Kimmel DL 28.115Kimmel DL 28, 115 Kimmel DL, Kimmel DB 28,115Kimmel DL, see Culberson JL 27, 28, 108 Kirchheim HR 24, 74, 84, 115 Kirchner D, see Kirchner F 58, 65-67, 69-71, 115 Kirchner F, Kirchner D, Polosa C 58,65-67,69-71,115 Kirchner F, Wyszogrodski I, Polosa C 58, 65-67, 69, 70,115 Kirchner F, see Horeyseck G 53, 54, 98, 101, 113 Kirchner F, see Preiss G 55, 58,66,120 Kitamura T, see Miura M 39, 118 Kleberg HD, see Gold MS 227, 231 Klee MR 53, 115 Kleinrok Z, see Zebrowska-Lupina I 214, 236 Kleist 18 Klevans LR, Gebber GL 87, 88,115 Klevans LR, see Gebber GL 55, 87, 88, 111 Klinke R, Fruhstorfer H, Finkenzeller P 131, 169, 190 Klonoff H, see Low MD 178, 191 Klorman R 163, 164, 190 Knapp DR, see Hsu CY 214, 215, 232 Knapp E, Schmid H, Ganglberger JA, Haider M 174, 190 Knapp E, see Ganglberger JA 136, 148, 156, 182, 187 Knott JR, Irwin DA 175, 190 Knott JR, Peters JF 175, 190 Knott JR, see Irwin DA 171, 189 Knott JR, see McCallum WC 128, 192 Knott JR, see Miller LH 172, 192 Knott JR, see Peters JF 171, 193

142, 171, 172, 193 Knott JR, see Zimmermann GN 177,197 Kobinger W 213, 232 Kobinger W, Pichler L 206, 209, 213, 216, 232 Kobinger W, Walland A 212, 232 Kobinger W, see Pichler L 205, 206, 208, 209, 213, 233 Koch E 53, 115 Köhler W, Held R 126, 190 Köhler W, Held R, O'Connel DW 126,190 Koella WP, see Rothlin E 17 Koella, see Wyss OAM 14 Koepchen HP, Langhorst P, Seller H 42, 43, 115 Koepchen HP, Langhorst P, Seller H, Polster J, Wagner D 42,43,115 Koepchen HP, Lux HD, Wagner PH 92, 93, 115 Koepchen HP, see Seller H 33,96,122 Kötter V, Zerbst E, Schulze A 84,115 Koh ET, see Ricardo JA 32, 34, 53, 88, *120* Koizumi K, Sato A 99, 115 Koizumi K, Seller H, Kaufman A, Brooks C McC 53-55, 57,115 Kok A 169, 190 Koninckx N, see Dongier M 175, 176, 186 Koninckx N, see Timsit M 175, 176, 195 Koninckx N, see Timsit-Berthier M 176, 195 Konishi M, see Watanabe S 145,196 Kooi KA, Bagchi BK 137, 190 Kopell BS, Tinklenberger JR, Hollister LW 178, 190 Kopell BS, see Ford JM 187 Kopell BS, see Roth WT 137, 168, 178, 193 Korner PI 25, 116 Korner PI, see Daly M de B 87,109 Korner PI, see Reynoldson JA 213,*234* Kornhuber HH, Becker W, Träumer R, Hoehne O, Iwase K 173,190 Kornhuber HH, Deecke L 127, 130, 135, 147, 148, 173,190

Knott JR, see Rebert CS

Kornhuber HH, see Becker W 160,184 Kornhuber HH, see Deecke L 130, 134–136, 160, 161, 167, 177, 185, 186 Kornmüller, see Weisschedel E 18 Kosaka K 76, 116 Kosaka K, Yagita K 32, 116 Kosterlitz HW, Lydon RJ, Watt AJ 210, 232 Kraemer HC, see Roth WT 137, 193 Kreidl, see Karplus 10 Kreuzer J, see Kurtzberg D 162,190 Kriebel I, see Deecke L 160, 167,186 Krienke B, see Camerer H 54, 107 Kristeva R, see Becker W 174,184 Kristeva R, see Deecke L 160, 167, 186 Kroneberg G, see Deck R 210, 229 Kroneberg G, see Heise A 212, 231 Kroneberg G, see Oberdorf A 210, 233 Kubacki A, see Jus K 171, 189 Kümmel H, see Janig W 54, 114 Kuffler SW, Nicholls JG 145, 190 Kugler J, see Hess WR 21 Kuhar MJ, see Young WS 226,*236* Kuhnt U, see Creutzfeldt O 145, 185 Kumada M, Nakajima H 31, 36,116 Kumada M, Nogami K, Sagawa K 89, 116 Kumada M, Sagawa K 90, 116 Kumada M, Schramm LP, Altansberger RA, Sagawa K 89,98,116 Kumada M, see Iriuchijima J 74, 75, 80, 81, 93, 114 Kuno M, see Fernandez de Molina A 55, 58–60, 110 Kunze DL 74, 75, 81, 116 Kuo JS, see Lee TM 50, 77, 116 Kurtzberg D, Vaughan HG, Kreuzer J 162, 190 Kutas M, Donchin E 174, 190

Kutas M, see Donchin E 157, 186 Kuypers HGJM, Maisky VA 65,116 Kwa HY, see Timmermans PBMWM 209, 216, 235 Kylstra PH, Lisander B 89, 98,116 Kylstra PH, see Djojosugito AM 89, 98, 109 Laffont F, see Lelord G 174, 191 Lafit AB, see Bennett JA 76, 77,79,106 Lalley PM, see Groat WC de 26, 29, 36, 109 Lam E, see Timmermans PBMWM 209, 213, 235 Lam RL, Tyler HR 35, 36, 116 Lancry A 164, 190 Landgren S 84,116 Langer SZ 199, 203, 206, 210, 232 Langer SZ, Massingham R, Shepperson NB 226, 232 Langer SZ, see Cubeddu L 206, 229 Langer SZ, see Dubocovich ML 206, 207, 230 Langer SZ, see Scatton B 214,234 Langer SZ, see Starke K 210, 234 Langer SZ, see Story DD 227, 235 Langhorst P, Stroh-Werz M, Dittmar K, Camerer H 42-44,116 Langhorst P, Werz M 41-43, 116 Langhorst P, see Camerer H 54,107 Langhorst P, see Koepchen HP 42, 43, 115 Langhorst P, see Seller H 33, 96, 122 Langhorst P, see Stroh-Werz M 41, 42, 94, 122 Langhorst P, see Werz M 41, 124 Lansing RW, see Wilke JT 174, 196 Larrabee MG, see Bronk DW 53, 65, 107 Larrabee MG, see Pitts RF 58, 62, 96, 97, 120 Lasch P, Jakobs KH 215, 232 Lathrop GH, see Ellingson RJ 137, 186

Lauber W, see Bauer H 171, 184 Lauener H, see Scholtysik G 213, 234 Lawn AM 116 Laxson C, see Martin GF 69, 117 Layne RS, see Callaway E 137, 184 Lean A de, see Hoffman BB 221, *231* Lebedev VP, Petrov VI, Skobelev VA 59, 61, 116 Lebel W, see Constantine JW 216, 226, 229 Lecrubier Y, see Puech AJ 227, 233 Ledsome JR, see Burkhart SM 48.107 Lee H, see Wasman M 141, 196 Lee TM, Kuo JS, Chai CY 50,77,116 Lee WH, Fujiwara M 210, 232Lefèvre F, see Cavero I 203, 228 Lefèvre F, see Roach AG 209, 234 Lefèvre-Borg F, Roach AG, Cavero I 206, 209, 232 Lefèvre-Borg F, see Cavero I 206, 209, 228 Lefkowitz RJ, see Hoffman BB 221, 231 Lefkowitz RJ, see Tsai BS 214, 222, 226, *235* Lefkowitz RJ, see Wood CL 201, 221, 236 Leger L, see Sakai K 34, 121 Leifer LH, see Otto DA 142, 192 Leighton J, Butz KR, Parmeter LL 204, 206, 208, 232 Lelord G, Laffont F, Sauvage D, Jusseaume P 174, 191 Leonard CM, see Norgren R 32, 119Levy MN 74, 86, 116 Levy MN, Degeest H, Zieske H 86,116 Levy MN, Zieske H 86, 87, 116 Lewis PR, Shute CCD 146, 191 Lewis PR, see Shute CC 146, 194 Lewis SJ, see Roth WT 168, 193 Libretti A, see Bartorelli C 53,106

Liebman J, see Robson RD 206, 214, 234 Lifschitz K 168, 191 Lille F, see Hazemann PS 174, 189 Linden RJ, see Keith IC 86, 89,115 Lindgren P, see Elliason S 97, 110 Lindsay PH, see Squires KC 132, 167, 194 Lindsley DB 128, 191 Lindsley DB, see Skinner JE 154,194 Lipski J, Coote JH, Trzebski A 47, 48, 54, 58, 66, 96, *117* Lipski J, McAllen RM, Spyer KM 28-30, 33, 34, 38-40, 42, 44-48, 50, 54, 58, 64, 66, 85, 94, 96, 116, 117 Lipski J, McAllen RM, Trzebski A 39, 44, 45, 47, 85, 116 Lipski J, Trzebski A 28, 29, 34, 36, 39, 40, 42, 45-47, 50, 64, 67, 94, 116 Lipson D, see Katona PG 74, 75,80,81,95,114 Lisander B, Martner J 90, 91, 117 Lisander B, see Djojosugito A 101, 109 Lisander B, see Djojosugito AM 89,98,109 Lisander B, see Folkow B 89, 110 Lisander B, see Kylstra PH 89,98,116 Little R, Wennergren G, Öberg B 86, 117 Livingstone KE, see Hockman CH 88, 90, 113 Ljung B, see Johansson B 222, *232* Löfving B 50, 67, 117 Lofving B, see Feigl E 98, 99, 110 Loewy AD, Burton H 32, 33, 34,117 Loewy AD, see Saper C 65, 83, 101, 121 Loizou LA 34, 117 Lokhandwala MF, Coats JT, Buckley JP 205, 233 Lokhandwala MF, see Hardey DW 227, 231 Long CNH, see Beattie J 65, 106Loomis AL, see Davis H 126, 185 Lopes OU, Palmer JF 87-89, 93, 95, 100, 117

McAdam DW, see Irwin DA

Lorenz 8,12 Louis G, see Rossi EC 215, 234 Louis WJ, see Jarrott B 217, 219, 232 Loveless N, Sanford A 163, 165, 168, 191 Loveless NE, Sanford AJ 165, 191 Low MD 178, 191 Low MD, Borda RP, Frost JD Jr, Kellaway P 140, 147, 159, 164, 171, 191 Low MD, Borda RP, Kellaway P 149, 171, 191 Low MD, Coats AC, Rettig GM, McSherry JW 172, 175, 191 Low MD, Klonoff H, Marcus A 178, 191 Low MD, McSherry JW 143, 171, 191 Low MD, Stoilen L 164, 165, 191 Lox HJ, see Göthert M 206, 231 Lubin A, see Naitoh P 177, 192 Lucet B, see Mouillé P 206, 209, 233 Lüders H 162, 191 Lundholm L, see Wikberg J 210, 236 Lutzenberger W, see Birbaumer N 171, 184 Lux HD, see Koepchen HP 92,93,115 Lydon RJ, see Kosterlitz HW 210, 232 MacDonald A, see Docherty JR 216, 226, 229 MacDonald RL, see Cohen DH 53, 54, 107 Machado PJ, see Velasco M 152, 159, 196 Macleod VH, see Coote JH 51, 56, 57, 63, 65, 67-71, 108 Madjar H, Docherty JR, Starke K 216, 233 Magnes J, Moruzzi G, Pompeiano O 53, 117 Magnus WOC 18 Magoun 10, 12, 18 Maisky VA, see Kuypers HGJM 65,116 Maldonado H, see Velasco M 152, 159, 196 Malta E, McPherson GA, Raper C 211, 233

Mannard A, Polosa C 53-55, 58–60, 66, 117 Manning JW 49, 50, 117 Manning JW, see Adair JR 37,99,104 Manning JW, see Kent BB 51, 115 Marcus A, see Low MD 178, 191 Marczynski TJ 130, 191 Marczynski TJ, Hackett CJ, Sherry CJ, Allen SL 146, 191 Marguth H, Raule W, Schaeffer H 74, 80, 117 Mark VH, see Ervin FR 152, 187 Markwardt F, see Glusa E 215, 231 Marriott AS, see Drew GM 214,230 Marsh GR 162, 165, 191 Marsh GR, Thompson LW 159,165,191 Marsh GR, see Poon LW 157, 193 Marshall I, Nasmyth PA, Nicholl CG, Shepperson NB 204, 206, 233 Marshall JM 89,117 Marshall WS, see Fuller RW 213, 230 Martin GF, Humbertson AO, Laxson C, Parreton WM 69, 117 Martin TL, see Coote JH 68, 108 Martinetti MG, see Zappoli R 179,197 Martner J, see Lisander B 90, 91, 117 Marttila I, see Euler C von 33,64,110 Martz MJ, see Karlin L 146, 167, 189, 190 Mascher D, see Alanis J 64, 104 Massingham R, see Cambridge D 203, 228 Massingham R, see Dubocovich ML 207, 230 Massingham R, see Langer SZ 226, 232 Mayanagi Y, see Fukushima T 152, 187 McAdam DW, Rubin EH 172, 174, 191 McAdam DW, Seales DM 174, 191 McAdam DW, Whittaker HA 174, 191

171, 189 McAdam DW, see Rebert CS 171, 172, 193 McAllen RM 36, 38-40, 85, 86, 89, 98, 99, 104, 117 McAllen RM, Jordan D, Spyer KM 26, 28-30, 39, 83, 118 McAllen RM, Spyer KM 39, 40, 44, 45, 46, 50, 74, 75, 77-82, 87, 93-95, 118 McAllen RM, see Hilton SM 49,91,*113* McAllen RM, see Humphreys PW 89,98,113 McAllen RM, see Lipski J 28-30, 33, 34, 38-40, 42, 44-48, 50, 54, 58, 64, 66, 85, 94, 96, 116, 117 McAllen RM, see Spyer KM 87,95,122 McCaffrey TV, see Barman SM 65,105 McCall RB, see Gebber GL 59, 61, 62, *111* McCall RS, Gebber GL, Barman SM 42, 55, 58, 60, 61, 67, 68, 73, 118 McCall RS, see Gebber GL 55,58-63,65-68,111 McCallum WC 135, 149, 153, 161, 175, 176, 178, 191, 192 McCallum WC, Abraham P 176, 192 McCallum WC, Cummins B 177, 192 McCallum WC, Knott JR 128, 192 McCallum WC, Papakostopoulos D, Griffith HB 130, 145, 146, 153, 154, 160, 182, 192 McCallum WC, Walter WG 143, 172, 175, 176, 183, 192 McCallum WC, Walter WG, Winter A, Scotton L, Cummins B 177, 192 McCallum WC, see Abraham P 176, 177, 183 McCallum WC, see Thompson JW 179, 195 McCallum WC, see Walter WG 127, 132, 159, 171, 181, 196 McCarthy G, see Squires KC 167, 168, 194 McCloskey DI, see Davidson NS 74, 80, 81, 89, 93, 109

McCloskey DI, see Davis AL 96,109 McCloskey DI, see Gandevia SC 89.110 McCloskey DI, see Haymett BT 89,93,112 McCulloch MW, see Medgett IC 208, 233 McCulloch MW, see Rand MJ 208, 234 McDonald DA, see Hess WR 21 McGrath JC, see Docherty JR 209, 216, 226, 229 McGrath JC, see Flavahan NA 216,230 McPherson GA, see Malta E 211, 233 McShane WK, see Constantine JW 206, 209, 229 McSherry JW 145, 192 McSherry JW, Borda RP 145, 149, 173, 192 McSherry JW, see Low MD 143, 171, 172, 175, 191 McWilliams PN, see Bennett JA 76, 77, 79, 106 McWilliam PN, see Donoghue S 38, 109 Meares RA, see Friedman J 178, 187 Medgett IC, McCulloch MW, Rand MJ 208, 233 Meinbresse D, see Tecce JJ 172, 183, 195 Mengel E, see Werz M 41, 124 Merker R, see Starke K 202, 203, 207, 208, 234 Merrill EG 64, 95, 118 Mesulam MM 32, 118 Metcalf G, see Doxey JC 206, 230 Metral S, see Hazemann PS 174, 189 Mey JG de, Vanhoutte PM 216, 229 Meyers P, see Miach PJ 217-219, 221, 233 Miach PJ, Dausse JP, Cardot A, Meyer P 217–219, 233 Miach PJ, Dausse JP, Meyer P 221, 233 Middleton HH, see Middleton S 75,118 Middleton S, Middleton HH, Grundfest H 75, 118 Middleton S, Woolsley CN, Burton H, Rose JE 41, 42, 118 Miller FR, Bowman JT 77, 118

Miller LH, Veen W van, Sandman CA, Knott JR 172,192 Miller LH, see Peters JF 171, 193 Miller R 76, 118 Millman IE, see Ashton H 178, 183 Mills E, see Kahn N 62, 63, 114 Mitchell GAG, Warwick R 74,76,118 Miura M 39, 118 Miura M, Kitamura T 39, 118 Miura M, Reis DJ 36, 37, 39, 40, 42, 44, 45, 46, 48, 50, 90, 91, 118 Miura M, see Homma S 40, 113 Miyamoto J, see Alanis J 64, 104Mohlant M 118 Mollica A, see Chiurugi E 77, 78, 107 Mollica A, see Scheibel M 40, 121 Monakows 17 Monnier, Marcel 17 Montel H, Starke K, Weber F 227,233 Montel H, see Starke K 202, 203, 207, 208, 234 Monti JM, see Hawkins M 214, 231 Mordkoff AM, see Karlin L 146, 190 Morehead SD, see Wasman M 141, 196 Morest PK 32-34, 39, 119 Moriyasu N, see Tsubokawa T 152, 153, 182, 195 Morris BJ, Reid IA, Ganong WF 226, 233 Moruzzi G 90, 101, 119 Moruzzi G, see Magnes J 53, 117 Moruzzi G, see Scheibel ME 40,121 Moruzzi J 12, 13, 18 Mottram DR, see Kapur H 206, 232 Mouillé P, Huchet AM, Chelly J, Lucet B, Doursout MF, Schmitt H 206, 233 Mouillé P, Huchet AM, Lucet B, Chelly J, Schmitt H 206, 209, 233 Mouillé P, see Andréjak M 211, 228 Moulds RFW, Jauernig RA 215, 233

Moulds RFW, see Jauernig RA 215, 232 Müller-Büton H, see Stegemann J 84, 122 Müller-Schweinitzer E, see Scholtysik G 213, 234 Munzer RF, see Baertschi AJ 86,105 Murray JG, see Agostoni E 75,104 Myers FK, see Gunn CG 77, 112 Naitoh P, Hilbert RP 177, 192 Naitoh P, Johnson LC, Lubin A 177, 192 Nakajima H, see Kumada M 31, 36, 116 Nakao H, see Gellhorn E 51, 52, 87, 99, 111 Nakazato Y, see Seno N 210, 234 Nasmyth PA, see Marshall I 204, 206, 233 Nathan M 63, 119 Nathan MA, Reis DJ 63, 119 Nathan MA, see Reis DJ 48, 49,120 Nathan MA, see Smith OA 50, 96, 101, 122 Nathan MA, see Snyder DW 49, 122 Nauta WJH, see Glees P 27, 111 Navona C, see Zappoli R 177, 197 Neil E, Palmer JF 74, 93, 119 Neil E, see Heymans C 24, 26, 53, 73, 74, 86, 112 Nelson B, see Ellingson RJ 137, 186 Netz J, see Grünewald G 130, 171, 174, 177, 188 Netz J, see Grünewald-Zuberbier E 136, 174, 177, 188 Neuberger M, see Groll-Knapp E 179, 187 Neumann P, see Robertson TW 76,120 Newton P, see Abraham P 177, 183 Newton P, see Papakostopoulos D 141, 147, 193 Newton P, see Thompson JW 179, 195 Ngai SH, see Bolme P 98, 106 Nicholl CG, see Marshall I 204, 206, 233

Nicholl JG, see Kuffler SW 145,190 Nicoll RA, see Barker JL 52, 105 Niemczyk H, see Baust W 44, 106 Nijkamp P, see Jong W de 48, 109 Ninomiya I, see Wilson MF 89, 98, 124 Nishida I, see Okada H 74, 75,80,119 Nogami K, see Kumada M 89,116 Nomura Y, Oki K, Segawa T 214, *233* Norgren R 33, 119 Norgren R, Leonard CM 32, 119 Nosaka S 44, 119 Nosaka S, Yamamoto T, Yasunaga K 76, 77, 119 Nowlin JB, see Thompson LW 165,195 Oberdorf A, Kroneberg G 210, 233 Oberdorf A, see Deck R 210, 229 Oberholzer RJH 48, 119 Öberg B, Thoren P 86, 119 Öberg B, see Folkow B 89, 99,110 Öberg B, see Little R 86, 117 Obrist WD, see Waszak M 146, 171, 196 O'Connel DW, see Köhler W 126, 190 Offner F, see Cohen J 165, 185 O'Flaherty JJ, see Wurtz RH 146, 196 Ohga A, see Seno N 210, 234 Okada H, Okamoto K, Nishida I 74,75,80,119 Okamoto K, see Okada H 74, 75,80,119 Oki K, see Nomura Y 214, 233 O'Leary J, see Heinbecker P 75,112 O'Leary JL, Goldring S 126, 181, 192 O'Leary JL, see Rhoton AL 28,120 Oliver JR, see Daly M de B 87,109 Osselton JW, see Cooper R 133, 138, 140, 185 Otto D 130, 192

Otto D, Houck K, Finger H, Hart S 163, 178, 192 Otto DA, Leifer LH 142, 192 Otto D, see Donchin E 149, 186 Overton DA, see Straumanis JJ 137,171,195 Owsjannikow P 49, 119 Paasonen MK, see Ylitalo P 49.124 Paini M, Zappoli R 164, 193 Paintal AS 26, 36, 119 Palkovits M, Zaborsky L 32, 48,119 Palkovits M, see Jong W de 48,109 Palkovits M, see Zandberg P 48, 49, 124 Palmer C, see Cohen J 165, 185 Palmer JF, see Lopes OU 87-89, 93, 95, 100, 117 Palmer JF, see Neil E 74, 93, 119 Papakostopoulos D 130, 149, 192 Papakostopoulos D, Crow HJ 146, 148, 149, 173, 192 Papakostopoulos D, Fenelon B 159, 171, 192 Papakostopoulos D, Winter A, Newton P 141, 147, 193 Papakostopoulos D, see McCallum WC 130, 145, 146, 153, 154, 160, 182, 192 Papakostopoulos D, see Thompson JW 179, 195 Papakostopoulos D, see Weinberg H 159, 196 Papini M, see Zappoli R 177, 197 Pappini M, see Zappoli R 179,197 Parmeter LL, see Leighton J 204, 206, 208, 232 Parreton WM, see Martin GF 69,117 Pascual W, see Anrep GV 92, 93,105 Pasquinelli A, see Zappoli R 177, 197 Patil PN, see Ruffolo RR 226, 234 Paton WDM, Vizi ES 210, 211, 233 Paul DD, Sutton S 167, 172, 193

Paxinos G, see Hunt GE 214, 232 Pearce JW, see Calaresu FR 77,78,107 Pearce JW, see Smith RS 41, 42, 122 Pearson JE, see Cant BR 159, 184 Peiss CN 25, 49, 62, 119 Pelayo F, see Scatton B 214, 234 Pelletier CL, Clement DL, Shepherd JT 84, 119 Pendleton RG, see Hieble JP 206, 231 Penna PE, see Keith IC 86, 89,115 Perez-Gonzalez JF, Rojas JR 90,119 Perez-Gonzalez JF, see Coote JH 89,98,99,108 Perl E, see Fernandez de Molina A 55, 58–60, 110 Peroutka SJ, Greenberg DA, U'Prichard DC, Snyder SH 219, 221, 233 Perrot P, see Cavero I 209, 228 Persson B, see Andén NE 213, 228 Peters JF, Knott JR, Miller LH, Veen WJ van, Cohen S 171, 193 Peters JF, see Knott JR 175, 190 Peterson LH, see Bagshaw RJ 89, 98, *105* Peterson CH, see Trzebski A 44, 47, 123 Petras JM, Cummings JF 58, 61,*120* Petrik GK, see Brickman AL 38, 52, 90, 106 Petrik GK, see Hamilton RB 112 Petrik GK, see Kaufman MP 52, 79, 88, 114 Petrov VI, see Lebedev VP 59, 61, *116* Petsche O, see Hess WR 21 Pettinger WA, see Berthelsen S 199, 201, 210, 212, 226, 228 Pfurtscheller G, Aranibar A 183, 193 Phillips G, see Adrian ED 53, 58,104 Pichler L, Kobinger W 205, 208, 209, 233 Pichler L, Placheta P, Kobinger W 206, 213, 233

250

Pichler L, see Kobinger W 206, 209, 213, 216, 232 Picton TW, Hillyard SA 169, 193 Piercey MP, see Gootman PM 43,111 Pieton TW, see Hillyard SA 168, 189 Pitts LH, see Cohen DH 53, 54,107 Pitts RF, Bronk DW 58, 59, 62,97,120 Pitts RF, Larrabee MG, Bronk DN 58, 62, 96, 97, 120 Pitts RF, see Bronk DW 53, 65,107 Placheta P, see Pichler L 206, 213, 233 Pocock PV, see Thompson JW 179,195 Poitras J, see Katona P 74, 75, 80, 81, 93, 114 Polosa C 53, 55, 58, 59, 66, 120 Polosa C, see Kirchner F 58, 65-67, 69-71, 115 Polosa C, see Mannard A 53-55, 58-60, 66, 117 Polosa C, see Preiss G 54, 55, 58,66,120 Polosa C, see Wyszogrodski I 58,124 Polster J, see Koepchen HP 42, 43, 115 Pommier Y, see Andréjak M 211, 228 Pompeiano O, see Ghelarducci B 91, 111 Pompeiano O, see Magnes J 53,117 Poon LW, Thompson LW, Williams RB, Marsh GR 157.193 Porszasz J, Porszasz-Gibischer K 44,120 Porszasz-Gibischer K, see Porszasz J 44, 120 Porter R 35, 120 Potter EK, see Davis AL 96, 109 Potter EK, see Gandevia SC 89,110 Preiss G, Kirchner F, Polosa C 55, 58, 66, 120 Preiss G, Polosa C 54, 58, 66, 120 Preobrazhenski NN 120 Presek P, see Glossmann H 222, 226, 231 Presek P, see Hornung R 217-219, 222, 231

Pribram K, see Donchin E 149,186 Prichard BNC 25, 120 Przegalinski E, see Zebrowska-Lupina I 214, 236 Przybyla AC, Wang SC 43, 120Puech AJ, Lecrubier Y, Simon P 227, 233 Puiggari MS, see Gunn CG 77, 112 Quest JA, Gebber GL 89, 120 Ramon Y, see Cajal S 27, 30, 35,76,77,*107* Rand MJ, McCulloch MW, Story DF 208, 234 Rand MJ, see Medgett IC 208, 233 Ransom W 4, 10, 12 Raper C, see Malta E 211, 233 Rasche A, see Grünewald-Zuberbier E 136, 174, 177, 188 Ratcheson R, see Hoffer BJ 101, 113 Raule W, see Marguth H 74, 80,117 Rawlins MD, see Ashton H 178, 183 Rebert CS 145, 146, 149, 154, 181, 193 Rebert CS, Irwin DA 139, 193 Rebert CS, Knott JR 142, 193 Rebert CS, McAdam DW, Knott JR, Irwin DA 171, 172, 193 Rebert CS, Sperry KG 171, 193 Rebert CS, Tecce JJ 171 Rebert CS, see Irwin DA 171, 189 Redgate ES, see Gellhorn E 51, 52, 87, 99, 111 Redmond DE, see Davis M 214, 229 Redmond DE, see Gold MS 227, 231 Reid IA, see Morris BJ 226, 233 Reis DJ, Cuenod M 51, 120 Reis DJ, Doba N, Snyder DW, Nathan MA 48, 49, 120 Reis DJ, see Crill WE 29-31, 108

Reis DJ, see Doba N 48-50, 109 Reis DJ, see Homma S 40, 113 Reis DJ, see Miura M 36, 37, 39, 40, 42, 44, 45, 46, 48, 50, 90, 91, 118 Reis DJ, see Nathan MA 63, 119 Reis DJ, see Snyder DW 49, 122 Rethelyi M 59, 61, 120 Rettig GM, see Low MD 172, 175, 191 Reynoldson JA, Head GA, Korner PI 213, 234 Rhodes KF, Waterfall JF 206, 234 Rhoton AL, O'Leary JL, Ferguson JP 28, 120 Ricardo JA, Koh ET 32, 34, 53,88,120 Richter DW 104 Richter DW, Heyde F, Gabriel M 64,120 Richter DW, Keck W, Seller H 54, 56, 96, 120 Richter DW, Seller H 44, 47, 48,*120* Richter DW, see Seller H 33, 54, 56, 96, 122 Richter D, see Seller H 33, 96,122 Rieckesmann JM, see Göthert M 206, *231* Rijlant P 74, 120 Ritchie JM, see Douglas WW 26,110Ritter Q, see Donchin E 157, 186 Ritter W, Vaughan HG 127, 130, 167-169, 193 Ritter W, Vaughan HH, Costa LD 127, 128, 131, 132, 168, 193 Ritter W, see Vaughan HG 157, 196 Ritter W, see Vaughan HG Jr 127, 130, 135, 148, 160, 161,196 Roach A, see Cavero I 203, 228 Roach AG, Lefèvre F, Cavero I 209, 234 Roach AG, see Cavero I 206, 209, 228 Roach AG, see Day MD 25, 109 Roach AG, see Lefèvre-Borg F 206, 209, 232 Roberts WJ, see Jankowska E 28,114
Robertson D, see U'Prichard DC 217-219, 221, 235 Robertson TW, Wallach J, Schneiderman N, Neumann P 76, 120 Robson RD, Antonaccio MJ, Saelens JK, Liebman J 206, 214, 234 Rockstroh B, see Birbaumer N 171, 184 Rössler R, see Anrep GV 92, 93,105 Rojas JR, see Perez-Gonzalez JF 90,119 Ronchi O, see Zappoli R 179, 197 Rose JE, see Middleton S 41, 42, 118 Rosen S, see Bolme P 98, 106 Rosenthal F, see Coleridge HM 53,108 Rosing EL, see Ruffolo RR 222, 223, 225, 234 Rossi EC, Louis G, Zeller EA 215, 234 Rossi L, see Zappoli R 179, 197 Roth WT, Ford JM, Lewis SJ, Kopell BS 168, 193 Roth WT, Kopell BS, Tinklenberg JR, Huntsberger GE, Kraemer HC 137, 193 Roth WT, Tinklenberg JR, Kopell BS 178, 193 Roth WT, see Ford JM 187 Rothlin E 2 Rothlin E, Koella WP 17 Rouot BM, U'Prichard DC Snyder SH 222, 226, 234 Rousseau JC, see Bostem F 175, 184 Rousseau JC, see Timsit-Berthier M 175, 176, 195 Routtenberg A 181, 193 Rouxs W 2 Rowland V 126, 140, 141, 170, 181, 194 Rowland V, see Wasman M 141,196 Rubin EH, see McAdam DW 172, 174, 191 Rubinstein EH, see Folkow B 89,99,110 Ruchkin DS, Sutton S 169, 194 Ruchkin D, Sutton S, Tueting P 131, 169, 194 Rudomin P 85, 121 Ruffolo RR, Rosing EL, Waddell JE 222, 223, 225, 234

Ruffolo RR, Turowski BS, Patil PN 226, 234 Rusinov VS 169, 194 Ruth R, see Karrer R 164, 166, 167, 190 Ryall RW, see Hongo T 58, 71, 113 Rydin C, see Andén NE 212, 213, 228 Rzepecki J, see Jus K 171, 189

Saelens JK, see Robson RD 206, 214, 234 Sagawa K, see Kumada M 89, 90, 98, 116 Saito Y, see Akimoto H 53, 104 Sakai K, Touret M, Salvert D, Leger L, Jouvet M 34, 121 Salmoiraghi GC 41-43, 121 Salomon LE, see Harter MR 131, 189 Salvert D, see Sakai K 34, 121 Salzmann R, see Scholtysik G 213,234 Samodelov LF, Godehard E, Arndt JO 84, 121 Samonina GE, see Dugin SF 77,78,110 Sampson SR, see Biscoe TJ 29, 36-40, 44, 45, 47, 85, 106 Sander G, see Grünewald G 171, 174, 188 Sandman CA, see Miller LH 172,192 Sanford A, see Loveless N 163, 165, 168, 191 Saper C, Loewy AD, Swanson LW, Cowan WN 65, 83, 101, 121 Sato A, Schmidt RF 56, 69, 121 Sato A, see Coote JH 56, 66, 67, 69, 108 Sato A, see Fidone SJ 26, 29, 110 Sato A, see Koizumi K 99, 115 Satomi H, see Todo K 76, 79, 123 Saur W, see Jakobs KH 214, 232 Sauvage D, see Lelord G 174, 191 Savignano-Bowman J, see Tecce JJ 172, 183, 195

Scatton B, Pelayo F, Dubocovich ML, Langer SZ, Bartholini G 214, 234 Scatton B, see Cavero I 209, 228 Schaeffer H 74, 80, 121 Schaeffer H, see Baust W 44, 106Schaeffer H, see Marguth H 74,80,117 Schaeffer H, see Weidenger H 74, 80, 81, 123 Schaltenbrand, Bailey 155 Schaumann W, see Douglas WW 26,110 Scheibel ME, Scheibel AB, Mollica A, Moruzzi G 40, 121 Scheibel AB, see Scheibel ME 40, 121 Scheid P, see Deecke L 130, 160, 161, 185, 186 Schenkenberg 162 Scherrer H 68, 121 Schild HO, see Arunlakshana O 202, 228 Schlör, Stock 101 Schmid H, see Ganglberger JA 136, 148, 155, 182, *187* Schmid H, see Haider M 149, 152, 154, 173, 182, 189 Schmid H, see Knapp E 174, 190 Schmidt RF, see Sato A 56, 69, 121 Schmidt RF, Schönfuss K 56, 121 Schmidt G, see Deecke L 177,186 Schmitt G, see Deecke L 166, 167, 186 Schmitt H 212, 213, 234 Schmitt H, Schmitt H, Fernard S 212, 234 Schmitt H, see Andréjak M 211, 228 Schmitt H, see Delbarre B 199, 212, 214, 229 Schmitt H, see Mouillé P 206, 209, *233* Schmitt H, see Schmitt H 212, 234 Schnall AM, see Cohen DH 53.54.107 Schneiderman N 104 Schneiderman N, see Brickman AL 38, 52, 90, 106 Schneiderman N, see Garcia M 28, 76, 77, 95, *111* Schneiderman N, see Gimpl MP 88, 90, 111

Schneiderman N, see Hamilton RB 112 Schneiderman N, see Jordan D 80, 85, 88, 100, 114 Schneiderman N, see Kaufman MP 52, 79, 88, 114 Schneiderman N, see Robertson TW 76, 120 Schneiderman N, see Schwaber J 38, 42, 79, 121 Schnieden H, see Ennis C 211, 230 Schocken DD, see Hoffman BB 221, 231 Schönfuss K, see Schmidt RF 56,121 Scholtysik G, Lauener H, Eichenberger E, Bürki H, Salzmann R, Müller-Schweinitzer E, Waite R 213, 234 Schramm LP, Bignall KE 65, 121 Schramm LP, see Kumada M 89, 98, 116 Schulte FJ, Henatsch HD, Busch G 53, 121 Schultz G, see Aktories K 226, 228 Schultz G, see Jakobs KH 214,*232* Schulze A, see Kötter V 84, 115 Schwaber JS, Cohen DH 79,121 Schwaber J, Schneiderman N 38, 42, 79, *121* Schwartz M, see Shagass C 137, 194 Schweitzer A 87, 121 Schweitzer A, Wrights S 53, 121 Schwent UL, see Hillyard SA 168, 189 Scott J, see Harper B 226,*231* Scotton L, see McCallum WC 177,192 Scrutton MC, see Grant JA 214, 215, 231 Seales DM, see McAdam DW 174, 191 Sears T 95, 121 Segawa T, see Nomura Y 214, 233 Seller H 53-58, 60, 64, 121 Seller H, Illert M 33, 37, 42, 85, 96, 121 Seller H, Langhorst P, Richter D, Koepchen HP 33, 96, 122

Seller H, Richter DW 33, 54, 56,96,122 Seller H, see Amendt K 32, 33,104 Seller H, see Dembowsky K 69,71,109 Seller H, see Gabriel M 29, 30, 36, 37, 48, 85, 110 Seller H, see Illert M 69,70,113 Seller H, see Koepchen HP 42, 43, 115 Seller H, see Koizumi K 53-55, 57, 115 Seller H, see Richter DW 44, 47, 48, 54, 56, 96, 120 Seno N, Nakazato Y, Ohga A 210, 234 Sevelius G, see Gunn CG 77, 112 Shagass C, Schwartz M 137, 194 Shagass C, see Soskis DA 137, 194 Shagass C, see Straumanis JJ 137, 171, 195 Shaw J, see Jauernig RA 215, 232 Shaw JC, see Cooper R 133, 138, 140, 185 Sheehan P, see Greenberg DA 219, 222, 231 Shepherd JT 84, 122 Shepherd JT, see Pelletier CL 84,119 Shepperson NB, see Langer SZ 226, 232 Shepperson NB, see Marshall I 204, 206, 233 Sherry CJ, see Marczynski TJ 146, 191 Shute CC, Lewis PR 146, 194 Shute CCD, see Lewis PR 146, 191 Sigg B, see Bonvallet M 28, 106 Simon P, see Puech AJ 227, 233 Skinner JE 154, 194 Skinner JE, Lindsley DB 154, 194 Skinner JE, Yingling CD 154, 194 Skinner JE, see Yingling CD 154.197 Skobelev VA, see Lebedev VP 59,61,116 Skolnik P, see Davis JN 218, 229 Sloniec M, see Zebrowska-Lupina I 214, 236

Small IF, see Small JG 171, 176, 194 Small JG, Small IF 171, 176, 194 Smith DBD, Donchin E, Cohen L, Starr A 167, 194 Smith DB, see Gerbrandt LK 130, 135, 136, 160, 161, 187 Smith CFC, see Doxey JC 203, 204, 206, *230* Smith OA 25, 62, 65, 122 Smith OA, Nathan MA 50, 96, 101, 122 Smith OA, see Vito JL de 76, 109Smith RS, Pearce JW 41, 42, 122 Smolem JA, Truex RC 76, 122 Snider RS, see Hoffer BJ 101, 113 Snoddy HD, see Fuller RW 213, 230 Snow HM, see Keith IC 86, 89,115 Snyder DW, Gebber GL 54, 55, 59, 63, 68, 122 Snyder DW, Nathan MA, Reis DJ 49, 122 Snyder DW, see Gebber GL 89,111 Snyder DW, see Reis DJ 48, 49,120 Snyder SH, see Greenberg DA 219, 221, 222, 231 Snyder SH, see Peroutka SJ 219, 221, 233 Snyder SH, see Rouot BM 222, 226, 234 Snyder SH, see U'Prichard DC 217-221, 225-227, 235 Solandt DY, see Bronk DW 106 Solyom L, see Barbas H 176, 184 Soskis DA, Shagass C 137, 194 Sparks H, see Djojosugito A 101, 109 Spatz, Hess WR 18 Speckmann EJ, Caspers H 181, 183, 194 Speckmann EJ, Caspers H, Janzen RW 145, 194 Speckmann EJ, see Caspers H 144, 145, 185 Spence MV, see Wilkinson RT 173, 196 Spencer DB, see Wood CC 159,196

Sperry KG, see Rebert CS 171, 193 Spyer KM 40, 44, 46, 51, 52, 74, 82, 83, 87, 95, 122 Spyer KM, McAllen RM 87, 95, 122 Spyer KM, Wolstencroft JH 36-40, 91, 122 Spyer KM, see Garcia M 28, 31, 34, 76, 77, 95, 111 Spyer KM, see Ghelarducci B 91,111 Spyer KM, see Hilton SM 39, 48-52, 67, 88, 91, 99, 113 Spyer KM, see Jordan D 28-31, 34, 40, 50, 80, 85. 86, 88, 89, 93, 98, 99, 100, 114 Spyer KM, see Lipski J 28-30, 33, 34, 38-40, 42, 44-48, 50, 54, 58, 64, 66, 85, 94, 96, 116, 117 Spyer KM, see McAllen RM 26, 28–30, 39, 40, 44–46, 50, 74, 75, 77-83, 87, 93-95,118 Squires KC, Donchin E, Hering R, McCarthy G 167, 168, 194 Squires KC, Hillyard SA, Lindsay PH 132, 167, 194 Squires NK, Squires KC Hillyard SA 131, 167, 168, 194 Squires KC, see Squires NK 131, 167, 168, 194 Stage L, see Johansson B 222, 232 Stalvey LP, see Davis JN 218, 229 Starke K 201, 203, 204, 207, 212, 213, 234 Starke K, Borowski E, Endo T 203, 234 Starke K, Endo T, Taube HD 202, 204, 208, 235 Starke K, Langer SZ 210, 234 Starke K, Montel H, Gayk W, Merker R 202, 203, 207, 208, 234 Starke K, Stamm 209 Starke K, Taube HD, Borowski E 202, 235 Starke K, see Borowski E 202, 206, 228 Starke K, see Endo T 202, 230 Starke K, see Madjar H 216, 233 Starke K, see Montel H 227, 233

204, 235 Starke K, see Tanaka T 217, 218, 227, *235* Starke K, see Taube HD 209, 227, 235 Starke K, see Weitzell R 202, 236 Starr A, see Smith DBD 167, 194 Stegemann J, Müller-Büton H 84,122 Steinsland OS, Hieble JP 206, 235Stephenson RP 200, 223, 235 Steppeler A, Tanaka T, Starke K 204, 235 Stidl H, see Groll-Knapp E 179, 187 Stjärne L 208, 235 Stock G 90 Stoilen L, see Low MD 164, 165, 191 Storm von Leeuwen W 138, 194 Story DD, Briley MS, Langer SZ 227, 235 Story DF, see Rand MJ 208, 234 Straumanis JJ, Shagass C, Overton DA 171, 195 Strömbom U, Svensson TH 214, 235 Strömbom U, see Andén NE 213, 228 Stroh-Werz M, Langhorst P, Camerer H 41, 42, 94, 122 Stroh-Werz M, see Camerer H 54,107 Stroh-Werz M, see Langhorst P 42-44,116 Struppler A, see Hess WR 21 Studynka G, see Haider M 127, 130-132, 147, 168, 188 Summers RJ, see Blakeley AGH 206, 228 Summers RJ, see Jarrott B 217, 219, 232 Sutton S 169, 195 Sutton S, Barren M, Zubin J, John ER 127, 128, 131, 167, 195 Sutton S, Tuetting P, Zubin J, John ER 167, 169, 195 Sutton S, see Donchin E 139, 186 Sutton S, see Paul DD 167, 172, 193 Sutton S, see Ruchkin D 131, 169, 194

Starke K, see Steppeler A

Svensson T, see Andén NE 212, 213, 228 Svensson TH, see Strömbom U 214, *235* Swanson LW 83, 122 Swanson LW, see Saper C 65, 83, 101, 121 Symmes D, Eisengart MA 168, 195 Szentagothai J 76, 122 Szulczyk P 65, 70, 123 Szulczyk P, Trzebski A 65, 70.123 Takahashi I, Todo K 76, 79, 123 Takata H, see Todo K 76, 79, 123 Talesnik J, see Hockman CH 88, 90, 113 Tam RMK, see Chess GF 107 Tan KS, see Katona PG 84, 114 Tanaka T, Starke K 217, 218, 227,235 Tanaka T, see Steppeler A 204, 235 Tanaka T, see Weitzell R 202, 236 Taube HD, Starke K, Borowski E 209, 227, 235 Taube HD, see Endo T 202, 230 Taube HD, see Starke K 202, 204, 208, 235 Taylor DG, Gebber GL 53-55, 58-60, 63, 66-68, 123 Taylor DG, see Gebber GL 63, 65-67, 111 Tecce JJ 143, 172, 181, 195 Tecce JJ, Cole JO 172, 178, 195 Tecce JJ, Hamilton BT 172, 195 Tecce JJ, Savignano-Bowman J, Meinbresse D 172, 183, 195 Tecce JJ, see Rebert CS 171 Telford R, see Ashton H 178, 183 Tempest R, see Trzebski A 44,123 Terry B, see Katona P 74, 75, 80, 81, 93, 114 Thamer V, see Horeyseck G 53, 54, 98, 101, 113 Thomas MR, Calaresu FR 52, 77,78,89,1*23* Thomas MR, Ulrichsen RF, Calaresu FR 68, 123

Thompson HK, see Besson JM 146, 184 Thompson IW, see Ashton H 178, 183 Thompson JW, Newton P, Pocock PV, Cooper R, Crow H. McCallum WC. Papakostopoulos D 179, 195 Thompson LW, Nowlin JB 165,195 Thompson LW, see Marsh GR 159.165.191 Thompson LW, see Poon LW 157, 193 Thoren P, see Öberg B 86, 119 Timmermans PBMWM, Kwa HY, Zwieten PA van 209, 216,235 Timmermans PBMWM, Lam E, Zwieten PA van 209, 213, 235 Timmermans PBMWM. Zwieten PA van 216, 235 Timms RJ 90, 123 Timms RJ, see Hilton SM 91, 113 Timsit M, Koninckx N, Dargent J, Fontaine O, Dongier M 175, 176, 195 Timsit M, see Timsit-Berthier M 137, 195 Timsit-Berthier M, Delaunoy J, Koninckx N, Rosseau JC 176, 195 Timsit-Berthier M, Delaunoy J, Xhenseval B, Timsit M 137, 195 Timsit-Berthier M, Hausman J 164, 176, 195 Timsit-Berthier M, Rousseau JC, Delaunoy J 175, 176, 195 Timsit-Berthier M, see Dongier M 175, 176, 186 Tinklenberg JR, sæ Kopell BS 178, 190 Tinklenberg JR, see Roth WT 137, 178, 193 Todo K, Yamamoto T, Satomi H, Ise H, Takata H, Takahashi I 76, 79, 123 Torvik A 27, 32, 33, 123 Touret M, see Sakai K 34, 121 Tower SS, see Bronk DW 106 Träumer R, see Kornhuber HH 173,190 Trobiani S, see Wurster RD 58.124 Trolin G, see Andén NE 213, 228

Truex RC, see Smolem AJ 76, 122 Trzebski A, Peterson LH 44, 47,123 Trzebski A, Peterson LH, Attinger F, Jones A, Tempest R 44, 123 Trzebski A, see Lipski J 28, 29, 34, 36, 39, 40, 42, 45-47, 50, 54, 58, 64, 66, 67, 85, 94, 96, *116, 117* Trzebski A, see Szulczyk P 65,70,123 Tsai BS, Lefkowitz RJ 214, 222, 226, *235* Tsai BS, see Wood CL 201, 221, 236 Tsubokawa T, Moriyasu N 152, 153, 182, 195 Tueting P, see Donchin E 157, 186 Tueting P, see Ruchkin D 131, 169, 194 Tueting P, see Sutton S 167, 169, 195 Turowski BS, see Ruffolo RR 226,234 Tuttle RS, see Djojosugito AM 89,98,*109* Tuttle RS, see Folkow B 89, *110* Tyler HR, see Lam RL 35, 36,116 Udelnov MG, see Dugin SF 77,78,110 Üvnas B, see Elliason S 97, 110 Ulrichsen RF, see Thomas MR 68,123 U'Prichard DC, Charness ME, Robertson D, Snyder SH 217-219, 221, 235 U'Prichard DC, Greenberg DA, Snyder SH 217-220, 225, 227, *235* U'Prichard DC, Snyder SH 217-219, 221, 226, 235 U'Prichard DC, see Greenberg DA 219, 222, 231 U'Prichard DC, see Peroutka SJ 219, 221, 233 U'Prichard DC, see Rouot BM 222, 226, 234 Vallbo AB, see Hagbarth RE 53,112 Vanhoutte PM, see Mey JG de 216,*22*9

Vaughan HG 127, 147, 157, 158, 195

Vaughan HG, Ritter W 157, 196 Vaughan HG, see Kurtzberg D 162, 190 Vaughan HG, see Ritter W 127, 128, 130-132, 167-169, 193 Vaughan HG Jr 148, 195 Vaughan HG Jr, Costa LD 127, 148, 195 Vaughan HG Jr, Costa LD, Ritter W 127, 130, 135, 148, 160, 161, 196 Vaughan HG Jr, see Gilden L 127, 130, 147, 187 Veen W van, see Miller LH 172, 192 Veen WJ van, see Peters JF 171, 193 Velasco M, Velasco F, Maldonado H, Machado PJ 152, 159, 196 Velasco F, see Velasco M 152, 159, 196 Verworn M 2, 3 Verzar F 17 Vieth J, see Baust W 44, 106 Vito JL de, Clausing KW, Smith OA 76, 109 Vizi ES 201, 210, 211, 235 Vizi ES, see Paton WDM 210, 211, *233* Vogt O 10 Vyshatina AI, see Gurevitch MI 90,112 Wacholder 3 Waddell JE, see Ruffolo RR 222, 223, 225, 234 Wagner D, see Koepchen HP 42, 43, 115 Wagner H, see Groll-Knapp E 179, 187 Wagner PH, see Koepchen HP 92, 93, 115 Waite R, see Scholtysik G 213, 234 Walker JM, see Doxey JC 203, 204, 206, 230 Wallach JH, see Hamilton RB 112 Wallach JH, see Kaufman MP 52, 79, 88, 114 Wallach J, see Robertson TW 76,120 Walland A 205, 206, 235 Walland A, see Kobinger W 212, 232 Walter WG 130, 134, 147, 159, 163, 165, 171, 172, 175, 176, 178, 196

Winter AL, see Cooper R

Walter WG, Cooper R, Aldridge VJ, McCallum WC, Winter AL 127, 132, 159, 171, 181, 196 Walter WG, see Cohen J 168, 185 Walter WG, see Cooper R 147, 185 Walter WG, see McCallum WC 143, 172, 175-177, 183, 192 Walter WG, see Weinberg H 131, 169, 196 Wang SC, Borison HL 84-87, 123 Wang SC, Chai CY 49, 123 Wang SC, see Chai CY 49, 107 Wang SC, see Folkow B 89, *110* Wang SC, see Katz RL 49, 114 Wang SC, see Przybyla AC 43,120 Ward DG, Baertschi AJ, Gann DS 86, 123 Ward DG, see Baertschi AJ 86,105 Warner HR, Cox A 74, 123 Warren C, see Karrer R 164, 166, 167, 190 Warwick R, see Mitchell GAG 74, 76, 118 Warzel H, Brattström A 84, 123 Wasman M, Morehead SD, Lee H, Rowland V 141, 196 Waszak M, Obrist WD 146, 171,196 Watanabe S, Konishi M, Creutzfeldt O 145, 196 Waterfall JF, see Algate DR 206, 228 Waterfall JF, see Rhodes KF 206, 234 Watt AJ, see Kosterlitz HW 210, 232 Waud DR 200, 223, 236 Weaver LC, see Gebber GL 63, 65-67, 111 Weber F, see Montel H 227, 233 Weber WV, see Coote JH 55-57,98,108 Weeks RA, see Constantine JW 206, 209, 216, 229 Weidenger H, Hetzel R, Schaeffer H 74, 80, 81, 123 Weinberg H 173, 196 Weinberg H, Papakostopoulos D 159, 196

Weinberg H, Walter W, Cooper R, Aldridge V 131, 169, 196 Weinberg H, Walter WG, Crow HJ 169, 196 Weiner N, see Cubeddu L 206, 229 Weiss GK, Crill WE 99, 123 Weiss GK, Kastella KG 39, 123Weisschedel E, Kornmüller 18 Weisschedel E, see Bartorelli C 18 Weisschedel E, see Hess WR 21 Weitzell R, Tanaka T, Starke K 202,236 Wennergren G, see Little R 86,117 Werz M, Mengel E, Langhorst P 41, 124 Werz M, see Langhorst P 41-43,116 Wessler I, see Kilbinger H 211, 232 Westbury DR, see Coote JH 55, 58-60, 73, 89, 108 Westfall TC 201, 236 Whittaker HA, see McAdam DW 174, 191 Whitting SB, see Drew GM 214, 215, 230 Whitwam JG, see Fussey IF 71,86,110 Widdicombe JG 75, 79, 124 Wikberg J 210, 211, 236 Wikberg J, Andersson R, Lundholm L 210, 236 Wikberg JES 201, 210-212, 220, 224, 225, 236 Wilczsak H, see Jus K 171, 189 Wilke JT, Lansing RW 174, 196 Wilkinson RT, Haines E 172, 196 Wilkinson RT, Spence MV 173, 196 Williams RB, see Poon LW 157, 193 Williamson PD, see Wood CC 159,196 Wilson H, see Green GJ 210, 211,*231* Wilson MF, Ninomiya I, Franz GN, Judy WV 89, 98, 124 Winter A, see McCallum WC 177, 192 Winter A, see Papakostopoulos D 141, 147, 193

147, 185 Winter AL, see Walter WG 127, 132, 159, 171, 181, 196 Wiprich L, see Gregor M 53, 54,112 Wolotsky P, see Gootman PM 43,111 Wolstencroft JH, see Spyer KM 36-40, 91, 122 Wood CC, Allison T, Goff WR, Williamson PD, Spencer DB 159, 196 Wood CL, Arnett CD, Clarke WR, Tsai BS, Lefkowitz RJ 201, 221, 236 Wood CL, see Hoffman BB 221, *231* Woody CD, see Besson JM 146,184 Woolsley CN, see Middleton S 41, 42, 118 Wrights S, see Schweitzer A 53, 121 Wurster RD 58, 124 Wurster RD, Trobiani S 58, 124 Wurster RD, see Barman SM 65,105 Wurster RD, see Chung JM 58,107 Wurster RD, see Foreman RD 55,65,70,110 Wurster RD, see Geis GS 55, 65, 66, 70, 76, 79, 111 Wurtz RH, O'Flaherty JJ 146, 196 Wyman R, see Euler C von 64,110 Wyman RJ, see Euler C von 33,64,110 Wyss OAM, Koella 14 Wyss OAM, see Hess WR 20 Wyszogrodski I, Polosa C 58, 124 Wyszogrodski I, see Kirchner F 58, 65–67, 69, 70, 115 Xhenseval B, see Timsit-Berthier M 137, 195 Yagi K, see Kannan H 52, 114 Yagita K, see Kosaka K 32, 116 Yamamoto T, see Todo K 76, 79, 1*23* Yamamoto T, see Nosaka S 76,77,119 Yamashita H 44, 52, 124

## Author Index

Yasunaga K, see Nosaka S 76, 77, 119 Yates MS, see Green GJ 210, 211, 231 Yingling CD, Skinner JE 154, 197 Yingling CD, see Skinner JE 154, 194 Yhitalo P, Karppanen H, Paasonen MK 49, 124 Young WS, Kuhar MJ 226, 236

Zaborsky L, sæ Palkovits M 32, 48, 119 Zakharow ST, see Dugin SF 77, 78, 110 Zanchetti A, Zoccolini A 90,

*124* 

Zanchetti A, see Bartorelli C 53, 106 Zanchetti A, see Guazzi M 53, 112 Zandberg P, Palkovits M, Jong W de 48, 49, 124 Zandberg P, see Jong W de 48, 109 Zappoli R, Denoth F, Navona C 177, 197 Zappoli R, Giuliano G, Rossi L, Pappini M, Ronchi O 179, 197 Zappoli R, Papini M, Briani S, Benvenuti P, Pasquinelli A 177, 197

Zappoli R, Papini M, Denoth F, Pasquinelli A, Rossi L, Martinetti MG, Guerri S 179, 197

Zappoli R, see Papini M 164, 193

Zbrozyna A, see Abrahams VC 52, 65, 97, 104

Zbrozyna AW, see Coote JH 98, 108 Zebrowska-Lupina I, Przegalinski E, Sloniec M, Kleinrok Z 214, 236 Zeller EA, see Rossi EC 215, 234 Zerbst ES, see Kötter V 84, 115 Zieske H, see Levy MN 86, 87,116 Zimmermann GN, Knott JR 177, 197 Zoccolini A, see Zanchetti A 90,124 Zubin J, see Sutton S 127,

Zubin J, see Sutton S 127, 128, 131, 167, 169, 195 Zwieten PA van, see Hoyer I

212, 232 Zwieten PA van, see

Timmermans PBMWM 209, 213, 216, 235

257

## Subject Index

acetylcholine 95 - and  $\alpha$ -agonists, release of 210 activation, cortical 181 - mechanisms, general 180, 181 affektive Abwehrreaktion 12 affinity of a drug 200 - ratios,  $\alpha_1 / \alpha_2 = 220, 222, 224$ age dependency of slow potentials 162 ff agonist potencies versus affinities 221 - -receptor dissociation constants, functional determination of 200 Aktivierungssystem, aufsteigendes 12, 13 alcohol 178  $\alpha$ -adrenergic agonists, potency of 207, 211 --, post-/presynaptic potency 202-204, 206,208 - -, presynaptic and dopamine derivatives 206 ---- , selectivity of actions 203  $\alpha$ -adrenoceptors, agonist affinities 201 -, distribution of 209 -, functional studies 201 ff -, heterogeneity of 209 -, postsynaptic 203, 207, 209 -, potencies of agonists and antagonists 201 - 203-, radioligand binding 216 ff at sympathetic nerve-muscle junctions 201 - 208-, vascular 215  $\alpha_1$  - and  $\alpha_2$  -adrenoceptors, definitions 201, 203  $\alpha_2$  -adrenoceptors outside noradrenergic axons 209 ff  $\alpha_2$ -autoreceptors 213, 214 amino acids, anterograde transport of 65 --, tritiated 83 amphetamine 178 anterograde transport 32 antidiuretic hormone, release of 52 anxiety and contingent potentials 175 aortic nerve 26 ---, antidromic volleys in 30 - -, fibre size in 36 ---, input to the subthalamus 52 - -, projection of 31 -- stimulation 36, 38, 84, 85, 88, 90 --, responses to 48 area postrema destruction 49

arousal 47,167 -, autonomic 183 ascending cholinergic reticular system 146 attention 168 - and slow potentials 171, 172, 174 atropine 95,179 Augenmuskellähmungen 13 autonomic control, specificity in 55 averaging 141-144 -, alternatives to 144 azapetine 202, 203, 218, 219 baroreceptor afferents, antidromic activation 28 - denervation 54 - fibres, composition of 26 --, termination of 25-31 - function, test of 50 - inhibition, spinal 62 - inputs, interactions between 85 - - to medulla 44-48 -, presynaptic actions 85, 93 - to respiratory neurons 47, 48 -, site of inhibitory control 56, 57 - - to suprabulbar areas 52, 53 -- mediated inhibition 67 baroreceptor reflex, bulbospinal interneurones in 47 - - - - pathways in 72 – –, cardiac, inhibition 88 ff - -, central delay of 56 - -, components of 97, 100 --, descending pathways 61 - -, effect of cerebellar stimulation 90 ---, facilitation 85-88 ---, frequency limitation 85 --, hypothalamic components of 83 --, - control of 100 --, input-output characteristics of 24 –, interactions between baroreceptor inputs 83--85 - -, interneurones in 72, 73 - -, lesion studies 48-51 - --, neuronal circuity 25 - -, and olivary stimulation 101 - - pathway 31 ff ---, neuroanatomical studies 32-34 ---, electrophysiological studies 34 ff - -, resetting of 97

baroreceptor reflex, sympathetic 97 -, total central inhibition 97–102 --, vagal, facilitation 88 --, -, reflex time 81, 82 - -, -, respiratory influences 92-96, 100 - response, cardiac and vascular components 87 - 89- rhythmicity, central 42-44 - stimulation 41 ff baroreceptors and vagal output 74, 75, 80 f -, quantitative effects 84 -, receptor properties of 24,84 basal ganglia 156 Bereitschaftspotential 127, 129, 130, 134, 136, 143, 147, 166 Bewegungskoordination 13 B fibre axons 75, 78–80 binding assays 219, 220, 222 bipolar recordings 30 "blind-sac" preparation 41,44 blood platelets, a-adrenoceptors 214, 215 Blutviskosität 8 bradycardia, vagal 50, 77, 78, 80, 87, 88, 90 brain metabolism 147 - slices,  $\alpha$ -adrenoceptors 209 bulbospinal excitatory system 62-64 - inhibition 64 - respiratory neurones 64 - - pathways 48 butyrophenones 206 caffein 178 carbon monoxide 179 cardiac efferent neurones 78 - rhythm 53–56,60 – –, central neuronal 42, 43, 45 cardio-accelerator neurones 55 cardiovascular depression, central receptors responsible for 213 - neurones, central 43-45 - reflexes 53 - system and motor cortex 91 carotid occlusion 44 - –, effect of 50 - sinus, reflexes from 48–50 catecholamine-containing neurones 28, 51 central nervous system,  $\alpha$ -adrenoceptors 210, 212 - 214cerebellar influences on baroreceptor reflex 91 - stimulation 90,91 cerebellum, feedback loops 156 C-fibers, cardiac 78 chemoreceptor afferents 26 - -, input from 45 - excitation 44 - reflexes 87,93 - -, cerebellar influences 90 - stimulation 70,93 chlordiazepoxide 178 p-chlorophenylalanine 68

cholinergic mechanisms in DC potentials 145, 146 - neurones,  $\alpha_2$ -adrenoceptors on 210 cirazoline 207 clonidine 71, 202-204, 206, 208, 210, 211, 213, 215, 218, 219, 220 -, efficacy of 224 -, postsynaptic/presynaptic potency ratio 202, 208, 209, 220 -, sedative properties 213, 214 <sup>3</sup>H-clonidine, specific binding 216-220, 222 clozapine 202, 203, 218, 219 cognitive decision 170 - evaluation 167 commissural nuclei 27, 34 common reference derivation 140 conditioning, operant, and DC potentials 171 contingent negative variation 127 - - -, drug effects on 178, 179 --, resolution of 173 - positive variation 130 - potentials 132-134, 182 - see also DC, late and slow potentials - -, age dependency 163 - and  $\alpha$ -activity 183 - -, cortical location 159, 160 – –, electrogenesis 159 --, localization of 149, 173 --, mathematical models 134 - - and psychological variables 170-173 - –, reliability of 137 - -, subcortical 152-154 - responses 127 --, during brain operations 152 -- and readiness potentials 147 - -, significance of 147 corneoretinal potential 140 corynanthine 202-203, 218, 219 cranial nerves, projection of 26, 27 current spread 37, 38 DC negativity 183 - potentials 126, 127 -- see also slow late or contingent potentials - -, artifacts in 140, 141 - -, data analysis of 141 - -, event-related 126 - - related to mental activity 126 ---, recording system 138-140 decerebration 50, 51, 54 defence reaction 52, 89, 97 --- and baroreceptor reflex 97 ff - - and cerebellum 90, 91 degeneration studies 32 depressor area 40, 50-52, 63 - -, hypothalamic 39,88 ---, medullary 29,66 dihydroergotamine 206 <sup>3</sup>H-dihydroergokryptine binding 221, 222 5-, 6-dihydroxytryptamine 68 discrimination, sensory 172

dissociation constants, drug-receptor 200 distraction, effects of 172, 175, 177 - - arousal hypothesis 172 diving response 87 Doppelbildmessung 2 dorsolateral funiculus, descending pathways 69,70 drug-receptor interaction 200 dyslexia, slow potentials in 177, 178 EC<sub>20</sub> pre values 202, 207 EC<sub>50</sub> values 200, 202, 208 EC<sub>x</sub> values 207 EEG artifacts 140, 141 -, delta-theta oscillations 43, 44, 54 - electrodes 138 - recordings, methods of 139 electrocorticographical studies 148, 149 electrode location 139 . - potential, drifts 139 -, reversible 138 Elektrodenlokalisierung im Gehirn 10, 11 electrooculogram, elimination from records 140, 141 emitted potentials 169 endophylaktisches System 9 Entlastungsreflex 8 epileptiform discharges 146 ereismatische Motilität 14 ergotropes System 9 event-related potentials, mechanisms of 180, 181 evoked potentials 137, 138, 141, 145 - in baroreceptor reflex 35-38 --, early and late components 152 –, number and location of electrodes 139 --, thalamic 151, 152 – –, visual 178 E-waves 175 expectancy wave 127 - processes 171 extinction 171 fastigial nucleus, stimulation 90 field stimulation 211 flexor reflex, increase in strength 213 flurazepam 178 gallbladder,  $\alpha$ -receptors 210 galvanic skin response 140 glial activity, slow potentials 145 glossopharyngeal rootlets, central projection 27 guanethidine 98 guanfacine 206 habituation 169 Hämodynamik, Ökonomieprinzip 8 heart rate, baroreceptor control of 93 -, function of vagosympathetic systems 87 -, vagal control of 73, 74

hippocampus, stimulation of 88-90, 101 Hirnstammreizung 10-12 Hirnstrukturen, Reizung und Ausschaltung von 5, 7, 10horseradish peroxidase 83 ----- , spinal injection 65 -- studies 32-34 --, transport of 68 - -, - to NTS 28 --, - in vagus 76 H-reflex and contingent potentials 147 5HT-containing neurones 68 hypertension and CNS lesions 49 -, development of 25 hypoglossal nerve, current spread to 37 hypothalamus, baroreceptor input to 52, 53 -, depressor area in 51 -, effects on cardiac baroreceptor reflex 89 -, effect on olivary complex 101 -, effects on sympathetic activity 62 -, relation to midbrain 65 -, paraventricular nucleus 83 -, pathways to medulla 83 -, pathways to spinal cord 65 -- preoptic region, anterior 39 -, role in baroreceptor reflex 51, 62 --, stimulation of 87-89, 99, 100 imidazolines, affinity ratios for 220, 223, 224 imperative stimulus 171 indoramin 206 inferior olive and baroreceptor reflex 101 information and late potentials 167 --- -related potentials 129, 130, 169 initiation potentials 130 inspiratory neurones 33 Instinkthandlung 12 intermediolateral column, interneurones in 73 intestine,  $\alpha$ -receptors of 210 Kreislaufphysiologie, Hess' Arbeiten zur 7-9 labetalol 206, 211, 214 laryngeal input 87 late positive component 128 - potentials 128-132 - -, see also contingent, DC and slow p. -, age-dependency 162, 163 --- and contingent responses 148 --, information-related 167-169 - -, localization of 148 - -, significance of 146 - - in subcortical structures 151, 152 - -, surface distribution 157, 158 - -, thalamic 169 lateral reticular nucleus see under reticular n. lesions in the medulla 76,77 locus coerulus 34 LSD-25, E-wave 178 lung inflation 89, 92, 95 - stretch afferents 89

Marchi technique 12, 27 mediothalamic-frontocortical system 182 medulla, see also reticular formation medullary neurones, convergence onto 85 --, temporal facilitation 46 - stimulation 87, 89, 90 methoxamine 202-204, 210, 211, 214, 218, 219  $\alpha$ -methyldopa 213, 218, 219  $\alpha$ -methylnoradrenaline 202, 203, 205, 211, 212, 214, 218-220 metoclopramide 179 motor cortex and baroreceptor reflex 91 - potential 127, 129, 130, 135, 136, 156 - -, localized 148,149 motorische Funktionen 13,14 movement, anticipated 171 -, goal-directed 174 - execution potentials 130 - related potentials 130, 131, 134-137 --- and age 166 ff -- and force 174 ---, goal-directed 130, 136, 137 ---, localization of 149, 150 – – –, psychological aspects 173, 174 ---, significance of 147 ---, subcortical 154-156 naphazoline 203, 204, 208, 210, 211, 214, 218 - 220Nauta technique 27 neuraxis, transections of 51 neurophysin 83 neurotics, contingent potentials in 175, 176 nitrazepan 178 noradrenaline 90, 201, 202, 204, 208, 211, 218 - 220- cell bodies,  $\alpha$ -adrenoceptors on 212 -- containing neurones 68-71 - neurones, inhibition of 213 -, effects on preganglionic neurones 71 - release 207 - stores, depletion of 211 -, synthesis and utilization 213 nucleus alaris 36 - ambiguus 32, 35, 36, 40, 45, 46, 50 --, anatomical studies 75-77 --, inspiratory neurones of 95, 100 --, recording from 78-80 --, stimulation of 77, 78 - gigantocellularis 37 - intercalatus 32 - intermediomedialis 73 – parabrachialis 34 - tractus solitarii 27-29, 72 ---, baroreceptor input to 45, 47 - - , bulbospinal inhibitory pathway 47 - - - , connexions of 82, 83, 85 ---, convergence onto 42, 43, 84, 85 --, descending control function 66, 67 --, diencephalic projection 34

nucleus tractus solitarii, dorsomedial 42 ---, efferent projections 32-34 ---, evoked responses in 35-39 ---, inspiratory neurones in 95 ---, effects of lesions of 48, 49 --, intermediate portion 86 ---, interneurones in 39 ---, monosynaptic input 39, 40 — — and noradrenergic neurones 34 — — —, primary afferent depolarization in 99 – – –, projection to lateral subthalamus 53 - - -, pulse-synchronous activity 41, 42 - - -, response to sinus nerve input 45, 47 ---, stimulation of 85 ---, subnuclei of 33 - - -, ventrolateral region 30, 32, 33, 45 Nutritionsreflexe 8 orienting potentials 168 - response, central 127 oxygen availabiltiy 147 oxymetazoline 204, 208, 211, 218-220 parahypoglossal region 38, 40, 42, 45, 46, 50, 64 paramedian reticular nucleus see under reticular nucleus Parkinson's disease, Bereitschaftspotential in 177 phenoxybenzamine 203, 206, 213, 218, 219 phentolamine 206, 209, 211 phenylephrine 202-204, 206, 208, 210, 211, 214, 218, 219 piperoxan 209, 211, 213, 218, 219 platelet adenylate cyclase 214 polarization of electrodes 138 positive waves 128 positivity, premotor 135 postganglionic see under sympathetic post-R-wave histograms 42, 43, 60, 74 postwarning stimulus activity 163 potassium, local release of 145 potency ratio,  $\alpha_1/\alpha_2 = 220, 222, 224$ --, postsynaptic/presynaptic 202-204, 206, 208, 220 prazosine 201, 202, 203, 206, 209, 211, 215, 218, 219, 221 <sup>3</sup>H-prazosine, binding 216–220, 222 preganglionic see under sympathetic premotion negativity 130,174 – positivity 149, 156 preparatory potentials 130 pressor area, medullary 43, 49, 63, 66 presynaptic modulation 85 primary baroreceptor fibres, projections of 26 ff propriospinal pathways 65, 71 psychological variables and slow potentials 131 psychology and DC potentials 167 ff

Psychophysiologie, Hess' Beiträge zur 14, 15

psychotics, slow potentials in 176 pulse-synchronous activity, neuronal 41, 42 putamen 182 pyramidal tract neurones 53 radioligand binding 200 -, in physiological media 222 – – studies 216 ff raphe system 34,66 raphe-spinal neurones 67 Raumwahrnehmung 13 rauwolscine 201-203, 218, 219 reaction time 171 readiness potential 127, 129, 130, 143, 149, 156, 173, 174, 176, 177, 182 - -, see also Bereitschaftspotential receptor occupation 224 - theory 223 recurrent inhibitory pathway 59, 61 reference system 140 reinforcement and DC potentials 170, 171 reliability coefficients 137 - of potential measurements 137 respiratory drive, central 66, 92, 95, 96 - "gate" on baroreceptor reflex 92-95, 100 - neurones, baroreceptor input 47,48 --, intracellular recordings 48 - rhythm 53-55 resting potentials 130 reticular formation, dorsolateral 49 - -, lateral 35, 36, 38, 45 - -, medial 37, 40, 46, 50, 66 --, "unspecific" 43 ---, ventrolateral 68 - -, ventromedial 69, 70 - neurones, multisensorial inputs 43,44 - nucleus, lateral 32, 50, 51, 63, 64, 68, 70 - -, paramedian 37, 64, 67, 91 - system, unspecific 173 retrieval task 174 reward-contingent positive variations 146 schizophrenics, contingent potentials in 175 - 177Schlafexperimente 12,13 Schlaftheorie 5, 7, 13 septum 87 silent period 59 sinus arrhythmia 87, 88, 92 - nerve, afferent fibres in 26, 29 - -, antidromic volleys in 29 -, electrical stimulation of 26, 36, 37, 39, 47, 51, 52, 70, 81, 82, 84, 85, 93–95, 99 - -, termination of various afferents 30 sleep and  $\alpha$ -receptors 213, 214 - deprivation 177 -wakefulness cycle 53, 181 slow potentials, amplitude evaluation 142, 143 - -, see also DC potentials – -, changes of 181

slow potentials, characteristics and methodology 131-144 - -, clinical studies 174-177 - -, concepts of 181 -- related to conditioned responses 126, 127 –, electrogenesis and physiological significance 144 ---, event-related 128 - -, genesis of 145, 146 ---, hierarchical model of 179 ff --, late responses 127 ----, reliability of 137 --, terminology and definitions 127-131 --, thalamic 146, 151, 154 --, variability of 138 smoking and evoked potentials 178 smooth muscle,  $\alpha_2$  -receptors 215, 216 sodium and ligand binding 222 solitariospinal projection 33 somatosympathetic reflexes 69,70 spinal sympathetic reflex, early and late 56, 57 split brain experiments 173 stereo-electro-encephalographical studies 150-156 Stereotaxie in Narkose 10 stimulation of the medulla 77 stimulus-response curve at  $\alpha$ -receptors 224, 225 stria terminalis 34 strychnine 59 summation, temporal and spatial 39 supra-optic nucleus 52 sustained potentials 128 sympathetic activity, control of 47, 48, 62 - -, efferent, baroreceptor control of 53 ff --, -, baroreceptor inputs 98 ---, generating system 43 - -, integrative control of 71–73 --, postganglionic 53, 54 - - , - , latency to inhibition 56 --, -, power spectral analysis 54 --, - renal 98 - control 62 - interneurones 63 - neurones, cholinergic action 59 ---, functional distinction 54 - -, preganglionic, descending control of 66 - 73- -, -, firing patterns 58 - -, -, interneuronal control of 61, 63 --, -, intrinsic properties 59 --, -, location of 57, 58 - -, -, respiratory influence upon 96 --, -, spinal 55 - - , - , spinal reflexes to 69, 70 -, -, spontaneous activity 58, 59 - - , - , structural data 58 --, -, supraspinal influences onto 62 ff

Subject Index

sympathetic somatic and visceral reflexes 55-57 sympatho-excitatory control 50, 51 - effects 68-71 - pathways 62-64 - -, descending 65, 66 sympatho-inhibitory control, descending 66 ff - neurones, ventrolateral 68 - pathways 66 - response 50 synaptic transmission and α-adrenergic drugs 203, 206

teleokinetische Motilität 14 termination potentials 131 test-retest reliability 137 thalamocortical interactions 174 thalamotomy 153 thalamus, motor relay nuclei 155 -, slow potentials in 152-155 thymoxamine 179, 206 tolazoline 213, 218, 219 toxicology of slow potentials 178, 179 tractus solitarius 27 - - see also nucleus t.s. tramazoline 202-204, 206, 211, 218-220 trigeminal nucleus, spinal 27 trophotropes System 9,12 truncothalamic system 182

vagal afferents, unmyelinated cardiac 86
efferents, activity to heart 86, 87
-, cardiac rhythm of 81, 82
inputs, convergence of 42, 43
motoneurones, cardiac 80-82
-, -, cholinergic innervation 96
-, -, inhibition of 100
-, -, rhythm of 94
-, species differences 79
, motor nuclei 74, 75
neurones, antidromic activation 28, 31
-, bronchoconstrictor 79, 80

vagal neurones, cardiac 46 -, cardio-inhibitory 74–76, 78 - -, oesophageal-motor 79 --, preganglionic 74 ff, 84 --, -, baroreceptor input to 80-83 --, -, location of 75 ff --, -, respiratory control 87, 88 --, -, supramedullary inputs 83 --, respiratory rhythm 74 - nucleus, dorsal 35, 38 - preganglionic supply 50 - rootlets, central projection 27 --, fibre composition 26 vago/sympathetic systems 86,87 vagus, dorsal motor nucleus 50 ---, anatomical studies 75-77 ---, baroreceptor input to 46 --, recording from 78–80 --, stimulation 77 -, HRP transport in 76 -, stimulation of 35 variability, interindividual 138 vasoconstrictor neurones 54, 55 - tone and baroreceptor reflex 98 - -, reduction in 97, 98 vasodilatation, cholinergic 91 vasodilator fibres, cholinergic 97, 98 "vasomotor" center, medullary 24, 43, 62 wakefulness 181 warning stimulus 171 <sup>3</sup>H-WB 4101 binding 216-220, 222 xylazine 204, 206, 210, 214, 215, 218, 219 yohimbine 201-203, 206, 209, 210, 213, 215, 218, 219, 221 Zielbewegungspotentiale 130, 136 Ziel- und Stützmotorik 13 zona incerta 53, 88, 155

Zwischenhirn, Beeinflussung vegetativer Funktionen 5, 7, 10